



## Outcomes of Influence of Dimethylarginine Derivats, PAD Isoenzymes, or both as Hallmark on Citrullinisation Process on Autoantigens ACPA/ Anti CCP Antibodies overall for Diagnosis in Rheumatoid Arthritis

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### Abstract

**Introduction:** When establishing new diagnostic method for detection of the endothelial dysfunction in Rheumatoid Arthritis (RA), it is necessary to compare the diagnostic values with other laboratory variables.

**Aim:** To Assess Asymmetric Dimethylarginine (ADMA) in RA and study its relation to ACPA and different clinical and laboratory parameters of disease activity in early non treated RA patients. Also, to determine the association between the asymmetric dimethylarginine (ADMA) and anti-cyclic citrullinated peptide antibodies (ACPA, Anti-CCP<sub>2</sub>) of the second generation in RA and to explain their connection in the process of citrullination of autoantigens in RA. Also, to determine the association between ADMA and the acute phase reactants: Rheumatoid factor (RF), C-reactive protein (CRP), as well as the index of disease intensity (DAS<sub>28</sub>) in early diagnosis in non-treated RA patients and to determine whether the amount of ADMA changes in the course of the evolution of disease.

**Methods:** Using the ELISA method (DLD-Diagnostika-GMBH for ADMA and BioSystems S.A. Reagens & Instruments Costa Brava 30, Barseelona, Spain), the sera of 70 subjects have been examined (35 RA not-treated, 35 healthy control group).

**Results:** Of 35 patients with RA, presence of ADMA was detected in 13 patients (37.14%); RF was present in 17 patients (48.57%), while anti-CCP<sub>2</sub> antibodies were present in 23 patients (65.71%). Of 18 patients who were RF negative, ADMA was detected in 9 patients (50%), while 11 patients (31.43%) were anti-CCP<sub>2</sub> positive. In 17 RF positive patients with RA, ADMA was detected in 4 patients, while 12 patients (34.28%) were anti-CCP<sub>2</sub> and RF positive. In the healthy control group 8 patients (22.85%) showed ADMA positivity. There was moderate correlation between ADMA and anti-CCP<sub>2</sub> antibodies in the group of patients with RA (r=0.34).

**Conclusion:** There was an association between ADMA and anti-CCP<sub>2</sub> antibodies of the second generation in patients with RA.

### Keywords

Asymmetric dimethylarginine (ADMA), Rheumatoid arthritis, Rheumatoid factor

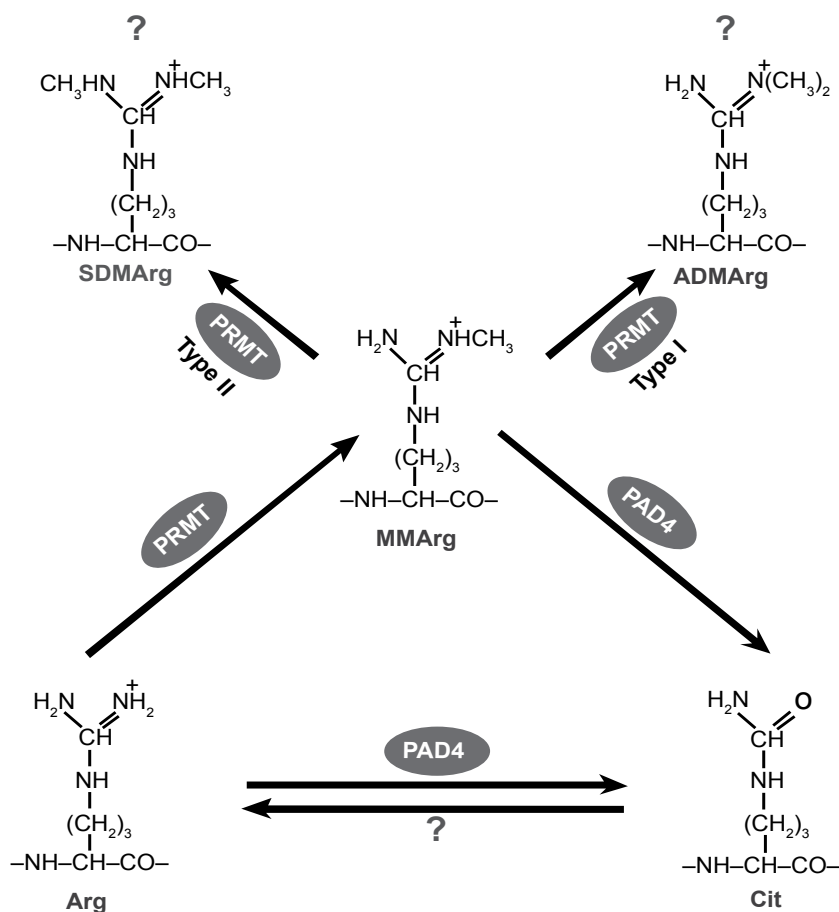
### Introduction

The association between arginine and citrulline, i.e. between asymmetric dimethylarginine (ADMA) as dimethyl derivative of the amino acid L-arginine and anti-cyclic citrullinated peptide antibodies (Anti-CCP<sub>2</sub>) of the second generation in patients with Rheumatoid Arthritis (RA) is the isoform of the enzyme Peptidylarginine Deiminase (PAD). Enzymatic deimination or protein citrullination is a process catalyzed by PAD enzymes which take part in the pathogenesis of RA. [1,2] In humans five PAD isoforms (PAD<sub>1-4</sub> and PAD<sub>6</sub>) are present. The greatest attention is paid to PAD<sub>2</sub> and PAD<sub>4</sub>, as potential candidates that could play role in the process of citrullination of autoantigens in RA. Both enzymatic forms are present in the rheumatoid synovium and fluid. [3-5] The polymorphism of PAD<sub>4</sub> is genetically associated with RA [6]. There is a difference in the tissue and cellular distribution of these two forms: PAD<sub>2</sub> is ubiquitously distributed in tissue (on the cytoplasmic level), while PAD<sub>4</sub> is more dominant in the hematopoietic cells (on nuclear level) [7-11]. In human neutrophils three different PAD isoforms (PAD<sub>2-4</sub>) are expressed. PAD targets are different cells' substrates on cellular or sub-cellular level. In the process of intracellular protein citrullination, different auto-antibodies recognize these substrates as citrullinated substrates. But, this explanation is probably partial, because the protein citrullination can take place also extracellularly. Cellular targets, identified as potential citrullinated autoantibodies with extracellular distribution are: filaggrin, vimentin, β-actin, collagen type I and II, gamma enolase etc [12-19]. Extracellular citrullination of fibrinogen is probably derived by PAD from damaged cells [4,20], but, so far it is not known in which conditions it occurs. Citrullinated autoantibodies play key role in the immune answer in RA. Few unique characteristics of protein citrullination enrolled in the pathogenesis of RA are identified: 1. The cytoplasmic content has autoantigenic features and is a target in the process of citrullination; 2. Although there are great number of proteins citrullinated by the active neutrophils, anti-CCP<sub>2</sub> recognises only small number of these molecules; 3. Citrulline activity of each PAD isoforms is characterized by the unique substrate specificity, independently of

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**Figure 1:** Metabolic aspects of dimethyl derivatives of the amino acid L-arginine and peptidylarginin deiminase (PAD).

their subcellular distribution. Only PAD<sub>2</sub> is capable of citrullination of native β/γ actine, while H<sub>3</sub> histones are only citrullinated by PAD<sub>4</sub> isoform. PAD<sub>4</sub> isoform which is cytoplasmic enzyme similar to PAD<sub>2</sub> is not able to citrullinate actine and H<sub>3</sub> histone; 4. Different reactive abilities of different sera in RA support the hypothesis that anti CCP<sub>2</sub> antibodies recognize the unique sequence [2,21]. The variations in the specificity of anti CCP<sub>2</sub> antibodies in different patients with RA are impressive. Probably all PAD isoforms share similar epitopes and their possible presence in the extracellular space indicate possible pass through the subcellular barriers.

### Some Aspects of Dimethylarginine Derivatives of the Amino Acid L-Arginine

There are 2 stereoisomers of the L-arginine: symmetrical (SDMA) and asymmetrical dimethylarginine (ADMA). ADMA interferes with L-arginine in the production of nitric oxide (NO), which plays key role in the normal endothelial function. Namely, NO is synthesized in the endothelial cells with the enzyme - endothelial nitric oxide synthetase (NOS). The physiological substrate (precursor) for NOS in this enzymatic process is L-arginine, converted in NO and L-citrulline. The NOS is inhibited by ADMA. The plasma level of ADMA is elevated in RA.

ADMA is synthesized by protein methylation mostly in the cellular nucleus. The methylation is catalyzed by the group of enzymes called protein arginine N-methyl transferases (PRMTs type I and II). Both PRMT subtypes have the ability to methylate monomethyl arginine (MMA); type 1 asymmetrically dimethylates arginine and creates ADMA, while type 2 catalyzes symmetrical dimethylation of arginine and creates SDMA (Figure 1).

#### Aim

The aim of this study is to determine the association between

ADMA and anti-cyclic citrullinated peptide antibodies (Anti-CCP<sub>2</sub>) of the second generation in patients with Rheumatoid Arthritis (RA) and to explain their connection in the process of citrullination of autoantigens in RA.

#### Material and methods

In the patients examined for this study, the diagnosis of the disease was established on the basis of revised diagnostic criteria for the classification of RA, suggested in 1987 by the American Association for Rheumatism (ARA) [22]. In order for a patient to be diagnosed with rheumatoid arthritis, patients must fulfill at least four out of seven criteria. Criteria from one to four are present for at least six month. In order to be included in the study every patient should fulfill at least 4 of the predicted 7 criteria. Criteria of diagnosis are related to American Colleague of Rheumatology (ACR).

In the study are included 70 patients, 35 patients with RA (28 women and 7 men), as well as 35 patients as healthy control group (18 women and 17 men). The mean age is 56, 68 years (±6.79) (40-65 years) in the group with RA and 46.2 years (±12.49) (29-65 years) in the healthy control group. The median duration of the disease in months is 43.97 (±45,23) in the interval of 1-168 months. Three patients were previously treated with oral steroids, while nobody was treated with NSAIDs. The others denied the use of other drugs such as arginine or nitroglycerin before the entrance in the study.

#### Inclusion criteria

In the study are included patients with RA aged 18-65 years, previously non-treated with NSAIDs or DMARDs.

#### Exclusion criteria

In the study are not included patients with diseases or conditions that could directly or indirectly influence the results, such as: 1. Patients with

previous history of spleen disease, thyroid disorders, hepatic damages, kidney, hematologic, neurologic and pulmonary disorders, autoimmune diseases, age <18 years; 2. Patients with Diabetes mellitus, acute infections, malignant diseases, febrile conditions ; 3. Patients with uric arthritis, urinary infections, SLE, mixed connective tissue disease, vasculitis; 4. Patients with previous history of blood transfusions, as well as overweight patients; 5. Patients treated with basic drugs for RA are excluded from the study; 6. Also, patients with the presence of hyperglycemia and elevated level of degradation products (serum urea and creatinin, urine creatinin), smokers, patients with arterial hypertension, hematologic and enzymatic disorders detected in 0 point, are excluded from the study; 7. Patients treated with cardiologic and hypolipidemic drugs, oral contraceptive drugs (estrogens, progesterone), vitamin E, nitroglycerin, arginine or analogues, and patients who received inhalational anesthetics one month before the start of the study are also excluded.

All participants voluntarily participated in the study, so the ethic criteria for the preparation of the study were fulfilled.

### clinical evaluation for disease activity

The clinical evaluation is made by the subspecialist in the field. Disease activity is evaluated using the DAS<sub>28</sub> index (Disease Activity Score (DAS<sub>28</sub>)). [23] The index uses mathematical formula in order to obtain unique composite quantitative score which consists of palpable painful joints (maximal number – 28), swollen joints (maximal number – 28), Westergren's Erythrocyte Sedimentation Rate (ESR) and patient's global evaluation for the disease activity (0-100 mm Visual Analogue Scale -VAS) as morning stiffness (in minutes). The DAS<sub>28</sub> index ranges from 0 to 10, and score below 3.2 qualifies the disease as low active.

### Laboratory evaluation

For clinical evaluation of RA several variables were taken into account: Complete Blood Count (CBC), differential blood count, reactants of the acute phase such as C-reactive protein (CRP), Rheumatoid Factor (RF) and Erythrocyte Sedimentation rate (ESR), anti CCP<sub>2</sub> antibodies, Alkaline Phosphatase (AP), Aspartate Aminotransferase (AST), Alanine Aminotransferase (ALT), Kreatinin Kinase (KK), Lactate dehydrogenase (LDH), serum urea and creatinin. The specimens are processed immediately (not frozen), respecting the rules of good laboratory practice.

### Determination of the activity of the serum Asymmetric Dimethylarginine : ELISA method (DLD Diagnostika-GMBH)-Enzyme Immunoassay

For quantitative determination of the endogen Asymmetric Dimethylarginine (ADMA) in plasma or serum

**Principle:** ADMA is connected in the solid phase of the micro titer plate. The ADMA standard, the specimens and positive control are pre-acylated in combination with rabbit anti-N-acyl-ADMA, and are incubated for 15-20 hours on 28°C. Later they are washed and the second antibody is added - anti rabbit IgG conjugated with peroxidase. They are incubated for 1 hour on room temperature. After washing, in order to get dyeing, tetramethylbenzidine is added as a substrate solution (TMB). After 20-30 minutes of the incubation it is stopped adding sulfuric acid. The absorbance is read on microtiter read on 450 nanometers. ADMA concentration is determined with the standard curve. Basically the amount of antibodies connected in the solid phase of ADMA is conversely proportional to the ADMA concentration in the specimens.

**Reference range:** ADMA in serum is 0.4-0.75 micromol/L.

CRP is determined by the agglutination test (Lateks CRP test) (BioSystems S.A. Reagens & Instruments Costa Brava 30, Barselona, Spain).

**Reference range:** <6 mg/L CRP in serum.

RF is determined by the agglutination test (Lateks CRP test) (BioSystems S.A. & Instruments Costa Brava 30, Barselona, Spain).

**Reference range:** <8 IU/ml in serum.

Anti CCP<sub>2</sub> antibodies are determined by the ELISA method (BioSystems S.A. Reagens & Instruments Costa Brava 30, Barselona, Spain).

**Reference range:** <20 IU/ml in serum.

Quantitative method for determination of the ESR is the Westergren method.

Reference range: 7-8mm for men, 11-16 for women.

### Statistical analysis

To test the significance of differences between two arithmetic means (proportions), the Student's t-test is used in comparison with the mean values of the determined numerical parameters between two groups, as well as Wilcoxon-matched test for independent specimens. The sensitivity and the prediction for the positive and negative test of the examined markers are determined with the test for sensitivity and specificity. The P-value in the range 0.05 and 0.1 is considered statistically significant. The data processing is made with the statistical package - Statistica 7.0.

Table 1: Laboratory results in RA and control healthy group (M ± SD)

	Non-treated RA Group N° 35 (M ± SD)	RA <sup>sero-</sup> N° 18 (M ± SD)	RA <sup>sero+</sup> N° 17 (M ± SD)	Control healthy group N° 35 (M ± SD)
	Positive/ negative	Positive/ negative	Positive/ negative	Positive/ negative
<b>ADMA + &gt; 0.75 (micromol/L)</b>	13/22 1,096 (± 0,68) (0,25-3,1)	9/9 1,12 (± 0,58) (0,32-2,1)	4/13 1,07 (± 0,79) (0,25-3,1)	8/27 1,00 (± 0,50) (0,26-1,91)
<b>RF + 30 &gt; IU/ml</b>	17/18 346,15 (± 625,22) (0,00-1920)	0/18 0,00 (± 0,00) (0,00-0,00)	17/0 712,67 (± 743,72) (30-1920)	2/33 13,71 (± 38,73) (0,00-120)
<b>CRP + 12 &gt; mg/L</b>	14/21 46,86 (± 79,19) (0,00-384)	3/15 8,66 (± 24,62) (0,00-96)	13/4 87,31 (± 96,44) (0,00-384)	4/31 5,48 (± 12,80) (0,00-48)
<b>ESR + &gt; 16</b>	27/8 48,62 (± 39,81) (2,0-120)	13/5 43,94 (± 39,82) (2,0-120)	14/3 53,58 (± 40,39) (5,0-120)	4/31 9,42 (± 8,21) (2,0-44)
<b>ACPA antibodies + &gt; 1,26</b>	23/12 1,71 (± 0,69) (0,92-3,0)	11/7 1,56 (± 0,59) (0,93-2,6)	12/5 1,87 (± 0,77) (0,92-3,0)	1/34 0,95 (± 0,10) (0,90-1,38)
<b>DAS 28 + &gt; 3,2</b>	28/7 4,79 (± 1,56) (1,85-7,03)	13/5 4,56 (± 1,76) (1,85-7,03)	15/2 5,04 (± 1,33) (2,47-6,83)	0/35 0,00 (± 0,00) (0,00-0,00)
<b>Morning stiffness + &gt; 0 min</b>	26/9 43,20 (± 65,13) (0-300)	14/4 57,50 (± 81,40) (0-300)	12/5 28,05 (± 38,72) (0-120)	0/35 0,00 (± 0,00) (0,00-0,00)

**Table 2:** Some diagnostic performance of laboratory variables in rheumatoid arthritis

	Rheumatic factor RA untreated group Number of patients 35	Rheumatic factor RA group <sup>sero-</sup> Number of patients 18	Rheumatic factor RA group <sup>sero+</sup> Number of patients 17	C-reactive protein RA untreated group Number of patients 35	C-reactive protein RA group <sup>sero-</sup> Number of patients 18	C-reactive protein RA group <sup>sero+</sup> Number of patients 17	Sedimentation RA untreated group Number of patients 35	Sedimentation RA group <sup>sero-</sup> Number of patients 18	Sedimentation RA group <sup>sero+</sup> Number of patients 17
<b>Sensitivity %</b>	48,57	0	100	66,66	16,66	76,47	77,14	72,22	82,35
<b>Specificity %</b>	94,28	94,28	94,28	88,57	88,57	88,57	88,57	88,57	88,57
<b>Predictive values for positive test %</b>	89,47	0	89,47	77,77	42,85	76,47	87,09	76,47	77,77
<b>Predictive values for negative test %</b>	35,29	35,29	0	40,38	36,60	11,42	20,51	13,88	8,82
<b>Accuracy %</b>	71,42	62,26	96,15	64,28	64,15	84,61	82,85	83,01	86,53

	Asymmetric dimethyl arginine RA untreated group Number of patients 35	Asymmetric dimethyl arginine RA group <sup>sero-</sup> Number of patients 18	Asymmetric dimethyl arginine RA group <sup>sero+</sup> Number of patients 17	Disease activity score ( DAS 28 ) RA untreated group Number of patients 35	Disease activity score ( DAS 28 ) RA group <sup>sero-</sup> Number of patients 18	Disease activity score ( DAS 28 ) RA group <sup>sero+</sup> Number of patients 17
<b>Sensitivity %</b>	74,28	77,77	70,58	80	72,22	88,23
<b>Specificity %</b>	100	100	100	100	100	100
<b>Predictive values for positive test %</b>	100	100	100	100	100	100
<b>Predictive values for negative test %</b>	20,45	10,25	12,5	16,16	12,5	5,40
<b>Accuracy %</b>	87,14	92,48	90,38	90	90,56	96,15

## Results

Of the 35 patients with RA, 13 patients (37,14%) showed presence of ADMA. RF was present in 23 patients (65,71%). Of 18 RF negative patients, ADMA was detected in 9 patients (50%), while 11 patients (31,42%) were anti CCP<sub>2</sub> positive. In 17 RF positive patients ADMA was present in 4 patients; 12 patients (34,28%) were anti CCP<sub>2</sub> and RF positive. In the healthy control group 8 patients (22,85%) showed ADMA positivity. RF was present in 2 patients (5,71%), while 1 patient (2,85%) showed anti CCP<sub>2</sub> positivity. In 35 patients with RA, DAS<sub>28</sub>>3.2 was present in 28 patients (80%). In 17 seropositive RF patients, DAS<sub>28</sub>>3.2 was present in 15 patients (88,23%) (Tables 1,2).

There is no statistical significance using Wilcoxon-matched test between ADMA in RA and the healthy control group for p<0.05 (p=0.555). 2. There is statistical correlation using Wilcoxon-matched test between ADMA in RA and age, duration of disease in months, DAS<sub>28</sub> index, RF and CRP, ESR, morning stiffness, in the same group for p<0.05: (ADMA vs age p=0.000; ADMA vs duration of disease in months p=0.000; ADMA vs DAS<sub>28</sub> p=0.000; ADMA vs RF p=0.018; ADMA vs CRP p=0.040; ADMA vs ESR p=0.000). 3. Although the presence of anti CCP<sub>2</sub> positive patients in DAS<sub>28</sub>>3.2 seropositive and seronegative patients is almost the same (10 vs 9 patients), the range of anti CCP<sub>2</sub> titre is higher in 10 RF seopositive patients with DAS<sub>28</sub>>3.2 than in RF seronegative patients with DAS<sub>28</sub>>3.2 (2.23 ± 0.61 vs 1.92 ± 0.45). There is no statistical correlation between these two groups (p=0.374) (Chart 1). 4. There is statistical correlation using Wilcoxon-matched test between anti CCP<sub>2</sub> in RA and the healthy control group for p<0.05 (p=0.000), as well as between anti CCP<sub>2</sub> in RA and DAS<sub>28</sub>, RF, CRP, ESR and morning stiffness in the same group for p<0.05 (anti CCP<sub>2</sub> vs DAS<sub>28</sub> p=0.000; anti CCP<sub>2</sub> vs RF p=0.018; anti CCP<sub>2</sub> vs CRP p=0.041; anti CCP<sub>2</sub> vs ESR p=0.000, anti CCP<sub>2</sub> vs morning stiffness p=0.000). There is moderate correlation between ADMA and anti CCP<sub>2</sub> antibodies in the group of patients with RA (r=0.34) (Chart 2).

## Discussion

The ADMA activity in serum is normally low. It is elevated in

endothelial dysfunction, because of the presence of the chronic inflammation in RA. Its presence is quantified by the degree of inflammation in RA which correlates with the disease activity. There is a strong correlation between ADMA and anti-CCP<sub>2</sub> antibodies from the second generation in early RA as a result of the disease activity and chronic inflammation [24-26].

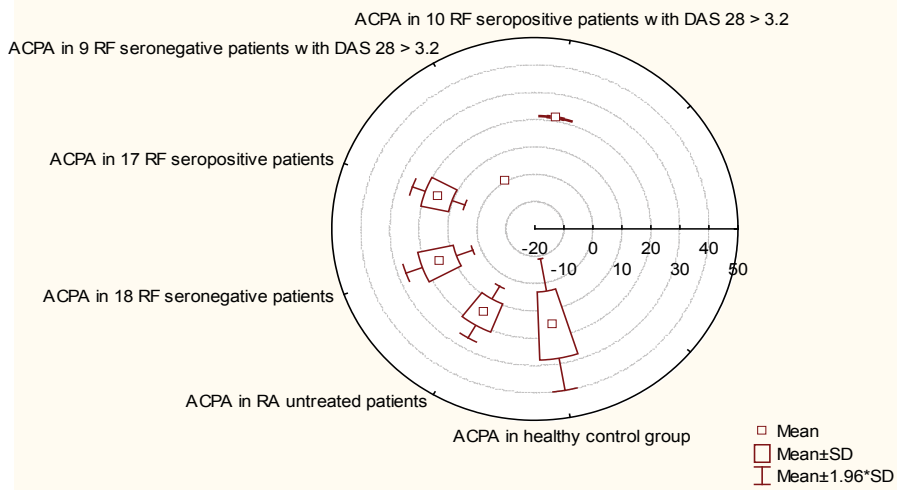
Seropositivity influences the ADMA induction (present also in our case) - seropositive RF patients with DAS<sub>28</sub>>3.2 have higher ADMA induction than seronegative RF patients with DAS<sub>28</sub>>3.2.

The association between ADMA and anti CCP<sub>2</sub> antibodies in patients with RA is confirmed also by other authors. They confirm the hypothesis that ADMA accumulation is associated with elevated titer of anti CCP<sub>2</sub> antibodies. Their conclusion is that excessive ADMA accumulation is associated with elevated titer of anti CCP<sub>2</sub> antibodies in patients with RA which duration is less than 3 years [25].

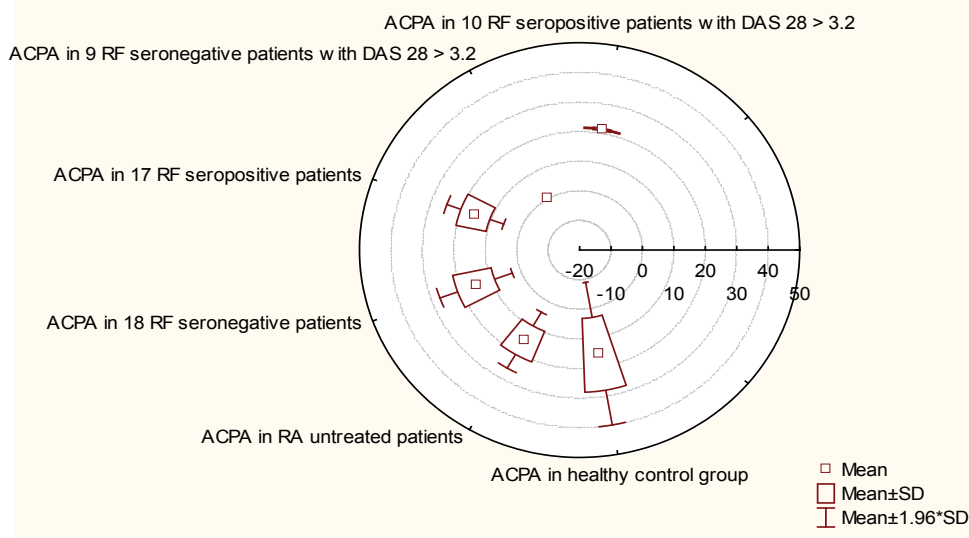
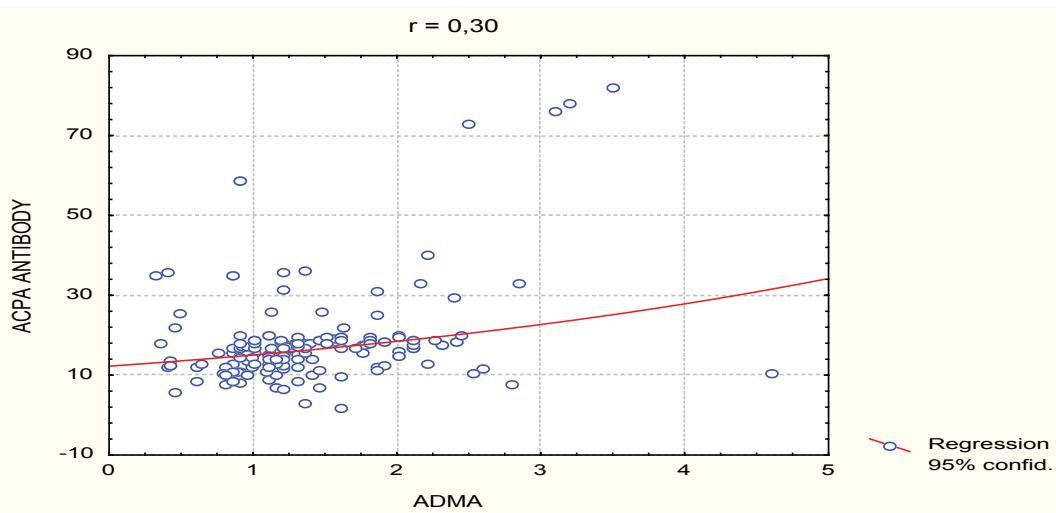
The association between anti CCP<sub>2</sub> antibodies and RF IgM in patients with RA and their predictive value is shown by other authors [27]. They conclude that anti CCP<sub>2</sub> antibodies and RF IgM are associated with the impaired endothelial function, independently from other cardiovascular risk factors. These autoantibodies could be reflection of the early atherosclerotic conditions and can induce increased risk of cardiovascular diseases.

The process of citrullination is described by other authors, also [28]. They demonstrate that human PAD<sub>4</sub> regulates the histone arginine methylation through conversion of methyl arginine in citrulline, releasing methylamine. The targets of PAD<sub>4</sub> are multiple areas in H<sub>3</sub> and H<sub>4</sub> histones that are methylated by the co-activators CARM<sub>1</sub> (H<sub>3</sub>Arg<sup>17</sup>) and PRMT<sub>1</sub> (H<sub>4</sub>Arg<sup>3</sup>). The decrease of the histone arginine methylation is secondarily associated with the increase in the process of citrullination. They point out that PAD<sub>4</sub> is mediated by the genetic expression through the regulation of arginine methylation and histone citrullination.

Other authors have almost equal perception [29]. They emphasize that PAD enzymes catalyze the conversion of arginine residues



**Chart 1:** Distribution of ACPA antibodies in Rheumatoid arthritis in all groups.



**Chart 2:** Correlation between ADMA and different parameter of RA. Pearson's coefficient of correlation (r) between the values of ADMA and ACPA antibodies in the group of patients with RA. There is a moderate correlation between ADMA and ACPA antibodies (r=0,30).

in proteins with citrulline residues. The citrulline is not a standard amino acid. It is not incorporated in proteins during translation, but is generated post-translationally by PAD enzymes. In normal conditions only nuclear histones possess this amino acid.

Other authors also agree with the principle of auto-antigenicity in RA [30]. They relate auto-antigenic citrullination and different PAD enzymes in RA, emphasizing that PAD enzymes have intrinsic capacity for selection of protein targets. Such specificity could play role in auto-antigenic selection in RA. Other authors [31-33] conclude that citrullination of the proteins is enabled with the enzymatic conversion (by PAD enzyme) from proteins that contain arginine residues towards citrulline residues. They agree that PAD enzymatic activity fulfill the criteria as additive marker in monitoring disease progression, together with anti CCP<sub>2</sub> and RF. They emphasize that the PAD method for detection is rapid, sensitive and relatively cheap.

They suggest that citrullination and methylation of the arginine residues is a process of competition. It is theoretically reversible in regard of methylation of arginine residues through conversion of mono-methyl-peptidyl-arginine (MMA) in citrulline. But, contradictory results are shown in terms of the capacity of the PAD enzymatic activity towards citrullination of MMA. In regard of the reversibility of the arginine methylation, in general, the protein methylation is one-way process and the methylated arginine residues rest as an integral part of the proteins so far, until they are degraded in the process of proteolysis. The protein methylation, generally, is considered as an irreversible process, due to the lost of demethylating enzymes in the process of conversion.

PAD<sub>4</sub> is not able to citrullinate directly ADMA, but, it interferes in ADMA synthesis via citrullination of MMA as an intermediary product in ADMA synthesis.

## Conclusion

ADMA is indirectly linked between autoantigenic citrullination and peptidyl-arginines' enzymatic isoforms in RA. Determination of ADMA together with other biological parameters contributes in the early diagnosis of RA.

## References

- Vossenaar ER, Zendman AJ, van Venrooij WJ, Puijntjens GJ (2003) PAD, a growing family of citrullinating enzymes: genes, features and involvement in disease. *Bioessays* 25: 1106-1118.
- Wegner N, Lundberg K, Kinloch A, Fisher B, Malmström V, et al. (2010) Autoimmunity to specific citrullinated proteins gives the first clues to the etiology of rheumatoid arthritis. *Immunol Rev* 233: 34-54.
- Foulquier C, Sebbag M, Clavel C, Chapuy-Regaud S, Al Badine R, et al. (2007) Peptidyl arginine deiminase type 2 (PAD-2) and PAD-4 but not PAD-1, PAD-3, and PAD-6 are expressed in rheumatoid arthritis synovium in close association with tissue inflammation. *Arthritis Rheum* 56: 3541-3553.
- Chang X, Yamada R, Suzuki A, Sawada T, Yoshino S, et al. (2005) Localization of peptidylarginine deiminase 4 (PAD4) and citrullinated protein in synovial tissue of rheumatoid arthritis. *Rheumatology (Oxford)* 44: 40-50.
- Kinloch A, Lundberg K, Wait R, Wegner N, Lim NH, et al. (2008) Synovial fluid is a site of citrullination of autoantigens in inflammatory arthritis. *Arthritis Rheum* 58: 2287-2295.
- Suzuki A, Yamada R, Chang X (2003) Functional haplotypes of PADI4, encoding citrullinating enzyme peptidylarginine deiminase 4, is associated with rheumatoid arthritis. *Nat Genet* 34: 395-402.
- Nakashima K, Hagiwara T, Yamada M (2002) Nuclear localization of peptidylarginine deiminase V and histone deimination in granulocytes. *J Biol Chem* 277: 49562-49568.
- Mastrorandi FG, Wood DD, Mei J (2006) Increased citrullination of histone H3 in multiple sclerosis brain and animal models of demyelination: a role for tumor necrosis factor-induced peptidylarginine deiminase 4 translocation. *J Neurosci* 26: 11387-11396.
- Harris ML, Darrach E, Lam GK, Bartlett SJ, Giles JT, et al. (2008) Association of autoimmunity to peptidyl arginine deiminase type 4 with genotype and disease severity in rheumatoid arthritis. *Arthritis Rheum* 58: 1958-1967.
- Andrade F, Darrach E, Gucek M, Cole RN, Rosen A, et al. (2010) Autocitrullination of human peptidyl arginine deiminase type 4 regulates protein citrullination during cell activation. *Arthritis Rheum* 62: 1630-1640.
- Goëb V, Thomas-L'Ottelier M, Daveau R, Charlonet R, Fardellone P, et al. (2009) Candidate autoantigens identified by mass spectrometry in early rheumatoid arthritis are chaperones and citrullinated glycolytic enzymes. *Arthritis Res Ther* 11: R38.
- Schellekens GA, de Jong BA, van den Hoogen FH, van de Putte LB, van Venrooij WJ (1998) Citrulline is an essential constituent of antigenic determinants recognized by rheumatoid arthritis-specific autoantibodies. *J Clin Invest* 101: 273-281.
- Girbal-Neuhaus E, Durieux JJ, Arnaud M, Dalbon P, Sebbag M, et al. (1999) The epitopes targeted by the rheumatoid arthritis-associated antifilaggrin autoantibodies are posttranslationally generated on various sites of (pro) filaggrin by deimination of arginine residues. *J Immunol* 162: 585-594.
- Vossenaar ER, Radstake TR, van der Heijden A, van Mansum MA, Dieteren C, et al. (2004) Expression and activity of citrullinating peptidylarginine deiminase enzymes in monocytes and macrophages. *Ann Rheum Dis* 63: 373-381.
- Vossenaar ER, Després N, Lapointe E, van der Heijden A, Lora M, et al. (2004) Rheumatoid arthritis specific anti-Sa antibodies target citrullinated vimentin. *Arthritis Res Ther* 6: R142-150.
- Kinloch A, Tatzer V, Wait R, Peston D, Lundberg K, et al. (2005) Identification of citrullinated alpha-enolase as a candidate autoantigen in rheumatoid arthritis. *Arthritis Res Ther* 7: R1421-1429.
- Matsuo K, Xiang Y, Nakamura H, Masuko K, Yudoh K, et al. (2006) Identification of novel citrullinated autoantigens of synovium in rheumatoid arthritis using a proteomic approach. *Arthritis Res Ther* 8: R175.
- Suzuki A, Yamada R, Ohtake-Yamanaka M, Okazaki Y, Sawada T, et al. (2005) Anti-citrullinated collagen type I antibody is a target of autoimmunity in rheumatoid arthritis. *Biochem Biophys Res Commun* 333: 418-426.
- Burkhardt H, Sehnert B, Bockermann R, Engström A, Kalden JR, et al. (2005) Humoral immune response to citrullinated collagen type II determinants in early rheumatoid arthritis. *Eur J Immunol* 35: 1643-1652.
- Takizawa Y, Suzuki A, Sawada T, Ohsaka M, Inoue T, et al. (2006) Citrullinated fibrinogen detected as a soluble citrullinated autoantigen in rheumatoid arthritis synovial fluids. *Ann Rheum Dis* 65: 1013-1020.
- Snir O, Widhe M, von Spee C, Lindberg J, Padyukov L, et al. (2009) Multiple antibody reactivities to citrullinated antigens in sera from patients with rheumatoid arthritis: association with HLA-DRB1 alleles. *Ann Rheum Dis* 68: 736-743.
- Arnett FC, Edworthy SM, Bloch DA, McShane DJ, Fries JF, et al. (1998) The American Rheumatism Association 1987 revised criteria for the classification of rheumatoid arthritis. *Arthritis Rheum* 31: 315-324.
- Balsa A, Carmona L, González-Alvaro I, Belmonte MA, Tena X, et al. (2004) Value of Disease Activity Score 28 (DAS28) and DAS28-3 compared to American College of Rheumatology-defined remission in rheumatoid arthritis. *J Rheumatol* 31: 40-46.
- Vaudo G, Marchesi S, Gerli R, Allegrucci R, Giordano A, et al. (2004) Endothelial dysfunction in young patients with rheumatoid arthritis and low disease activity. *Ann Rheum Dis* 63: 31-35.
- Surdacki A, Martens-Lobenhoffer J, Wloch A, Glusko P, Rakowski T, et al. (2009) Plasma asymmetric dimethylarginine is related to anticitrullinated protein antibodies in rheumatoid arthritis of short duration. *Metabolism* 58: 316-318.
- Turiel M, Tomasoni L, Sitia S, Cicala S, Gianturco L, et al. (2010) Effects of long-term disease-modifying antirheumatic drugs on endothelial function in patients with early rheumatoid arthritis. *Cardiovascular Therapeutics* 28: 53-64.
- Hjeltne G, Hollan I, Førre Ø, Wiik A, Mikkelsen K, et al. (2011) Anti-CCP and RF IgM: predictors of impaired endothelial function in rheumatoid arthritis patients. *Scand J Rheumatol* 40: 422-427.
- Wang Y, Wysocka J, Sayegh J, Lee YH, Perlin JR, et al. (2004) Human PAD4 regulates histone arginine methylation levels via demethylination. *Science* 306: 279-283.
- Rajmakers R, Zendman AJ, Egberts WV, Vossenaar ER, Raats J, et al. (2007) Methylation of arginine residues interferes with citrullination by peptidylarginine deiminases in vitro. *J Mol Biol* 367: 1118-1129.
- Darrach E, Rosen A, Giles JT, Andrade F (2012) Peptidylarginine deiminase 2, 3 and 4 have distinct specificities against cellular substrates: novel insights into autoantigen selection in rheumatoid arthritis. *Ann Rheum Dis* 71: 92-98.
- Basu PS, Majhi R, Ghosal S, Batabyal SK (2011) Peptidyl-arginine deiminase: an additional marker of rheumatoid arthritis. *Clin Lab* 57: 1021-1025.
- Zhang Y (2004) Molecular biology: no exception to reversibility. *Nature* 431: 637-639.
- Cuthbert GL, Daujat S, Snowden AW, Erdjument-Bromage H, Hagiwara T, et al. (2004) Histone deimination antagonizes arginine methylation. *Cell* 118: 545-553.