



SHORT COMMUNICATION

Polycystic Ovary Syndrome: Genetic Contributions from the Hypothalamic-Pituitary-Gonadal Axis

Branavan Umayal¹, NV Chandrasekharan², WSS Wijesundera³ and Chandrika N Wijeyaratne^{1*}

¹Department of Obstetrics and Gynecology, Faculty of Medicine, University of Colombo, Sri Lanka

²Department of Chemistry, Faculty of Science, University of Colombo, Sri Lanka

³Department of Biochemistry and Molecular Biology, Faculty of Medicine, University of Colombo, Sri Lanka



*Corresponding author: Chandrika N Wijeyaratne, Department of Obstetrics and Gynecology, Faculty of Medicine, University of Colombo, PO Box 271, Kynsey Road, Colombo 08, Sri Lanka, Tel: 0094-77-7344480, Fax: 0094-11-2691581, E-mail: mandika59@hotmail.com

Abstract

Polycystic Ovary Syndrome (PCOS) is a common endocrine disorder, present in 5-13% women of reproductive age. The endocrine manifestations of PCOS include excess androgen production of ovarian and/or adrenal origin and arrested follicular development leading to chronic oligo- or anovulation. Long term health risks of women with PCOS include cardiovascular disease, type 2 diabetes mellitus and endometrial cancer. PCOS is diagnosed by chronic anovulation, polycystic ovaries on ultrasound and biochemical/clinical manifestations of hyperandrogenism. Phenotype expression is heterogeneous and varies throughout the woman's life cycle, making early confirmation difficult. South Asians with anovulatory PCOS manifest severe symptoms at a younger age, with greater insulin resistance and a higher prevalence of the metabolic syndrome than white Europeans, thereby reflecting their ethnic propensity to type 2 diabetes mellitus. PCOS appears to be a multigenic trait, although contributing genes remain undefined yet. Several studies have been carried out to identify the candidate genes and polymorphisms affecting the multiple biological pathways of PCOS. The main objective of this article is to review the role of genes regulating the Hypothalamus-Pituitary-Gonadal (HPG) axis - mainly *KISS1*, *GPR54* receptor gene, *GnRH* (Gonotropin Releasing Hormone), *GnRHR* (Gonotropin Releasing Hormone Receptor), *FSH* (Follicle Stimulating Hormone), *FSHR* (Follicle Stimulating Hormone Receptor), *LHβ* (Luteinizing Hormone beta subunit) and *LHCGR* (Luteinizing Hormone/Choriogonadotropin receptor) genes; with special focus on its association with PCOS.

Keywords

Hypothalamic pituitary gonadal axis, Polycystic ovary syndrome, South Asia

Introduction

Polycystic Ovary Syndrome (PCOS) is a common endocrine disorder, affecting women of reproductive age with prevalence varying between 5-13%. PCOS typically presents during adolescence with a wide spectrum of phenotypes that are characterized by features of anovulation (amenorrhoea, irregular cycles) combined with symptoms of androgen excess (hirsutism, acne, alopecia) and polycystic ovaries on ultrasound [1]. The characteristic biochemical abnormalities are elevation of serum androgen concentrations (particularly testosterone and androstenedione) and Luteinizing Hormone (LH) concentrations, but with normal or low levels of Follicle-Stimulating Hormone (FSH) [2]. Diagnosis of PCOS is based on the 'Rotterdam criteria', which require the presence of two of the three following features: polycystic ovaries, anovulation and androgen excess (clinical and/or biochemical) [3].

Current studies suggest that excess androgen production may induce the polycystic ovarian morphology, leading to the endocrine disruption of this disorder. In addition, presentation of PCOS in adolescence suggests that there is an underlying predisposition to the typical ovarian abnormalities that has origins before the onset of puberty. The basis of these two hypotheses was obtained from studies done by Abbott and colleagues on female Rhesus monkeys. In this study, the animals were exposed to high concentrations of testosterone *in uter-*

us and when they developed as adults showed typical features of PCOS such as hypersecretion of LH, ovarian hyperandrogenism, anovulation in relation to increased body weight and insulin resistance [4,5]. This shows that PCOS is induced by excess androgens and may arise by 'programming' of the hypothalamic-pituitary-ovarian axis by androgens in prenatal life [6].

PCOS is associated with metabolic abnormalities, central to which are insulin resistance and hyperinsulinemia, which carry an increased risk of developing type 2 diabetes in later life [2]. In addition, PCOS is one of the leading causes of female infertility. Lifestyle changes, such as losing weight, can trigger body changes that facilitate conception in women with PCOS [7].

Based on current research on PCOS there is strong evidence that genetic factors play a major role in its etiology. Despite several genetic studies dissecting the variants of genes from multiple biological pathways in its pathophysiology, the mode of inheritance of PCOS remains unclear [8]. Current findings favor PCOS as a complex endocrine disorder that results from the interaction of susceptible and protective genomic variants in several genes under the influence of environmental factors [9-11]. Candidate genes involved in steroid hormone metabolism, gonadotropin and gonadal hormones action, obesity and energy regulation, insulin secretion and action have been studied and implicated in the pathogenesis of PCOS. There may be several interlinking factors that affects expression of PCOS. A single cause for PCOS is unlikely [12]. Moreover, a study on phenotype genotype correlation in PCOS revealed that a PCOS genetic gradient resulted from genetic drifts due to a serial founder effect that occurred during ancient human migrations. The overall prevalence of the disease supports intra-locus sexual conflict as alternative to the natural selection of phenotypic traits in females [13].

We believe that focusing on the Hypothalamus-Pituitary-Gonadal (HPG) axis related genetic polymorphisms may shed fresh light on providing a more complete picture on the genetic basis of PCOS. Heritability can be studied by 4 methods - twin studies, family association studies, candidate gene studies and Genome-Wide Association Studies (GWAS). In this review we have analyzed the heritability based on the candidate gene studies related to the HPG axis and PCOS.

In our review, the role of genes regulating the HPG axis - mainly *KISS1*, *GPR54* receptor gene, *GnRH* (Gonadotropin Releasing Hormone), *GnRHR* (Gonadotropin Releasing Hormone Receptor), *FSHβ* (Follicle Stimulating Hormone beta subunit), *FSHR* (Follicle Stimulating Hormone Receptor), *LHβ* (Luteinizing Hormone beta subunit) and *LHCGR* (Luteinizing Hormone/Choriogonadotropin receptor) genes; with special focus on their association with PCOS were selected. Several studies have been carried out to identify the association of polymorphisms of these genes with polycystic ovary syndrome. However,

repeatability of results has remained low.

***KISS1* and *GPR54* Genes**

Female reproductive function depends on the proper development and regulation of the HPG axis. Kisspeptins are peptide products of *KISS1* gene that participate in the control of the HPG axis. Kisspeptin act via G protein-coupled receptor known as *GPR54* [14,15]. The *GPR54* - *KISS1* pathway has an essential role in the initiation and maintenance of mammalian fertility [16].

The *KISS1* gene is localized on chromosome 1q32 and consists of three exons, of which only part of the second and third exons are finally translated into a precursor 145 amino acid peptide, which is cleaved into three forms of kisspeptins containing 54, 14, or 13 amino acids. The three peptides exhibit the same affinity for their single receptor (*GPR54*) since they share a common C-terminal decapeptide. *GPR54* gene maps to chromosome 19p13.3 and includes five exons, encoding a 398 amino acid protein with seven hydrophobic trans-membrane domains [17].

The *KISS1* gene was originally identified in 1996 as a suppressor of metastasis in human malignant melanoma [18]. The role of Kisspeptin in reproduction was identified in 2003, which revealed the current understanding of the neuroendocrine regulation of human reproduction and the role of kisspeptin in puberty [19]. Kisspeptin signals directly to the GnRH neurons through the action on the kisspeptin receptor (*GPR54*) to release GnRH into the portal circulation, which in turn stimulates the secretion of LH and FSH from the gonadotrophs of the anterior pituitary [20]. GnRH secretion is deregulated in PCOS. Therefore, it can be postulated that altered patterns of kisspeptin inputs to GnRH neurons leads to dysregulated gonadotropin secretion in PCOS.

The most important function of the *KISS1/GPR54* system in the process of puberty makes it necessary to investigate the mutations and polymorphisms in the *KISS1* and *GPR54* genes and their association with PCOS. However, polymorphisms in *KISS1* and *GPR54* genes in relation to PCOS are not well studied. Therefore, further research should focus on identifying the polymorphisms in *KISS1* and *GPR54* genes and their expression levels in relation to PCOS and determine the association of these polymorphisms with clinical, biochemical and endocrine features in order to obtain a complete view of the scenario predisposing to PCOS. This will help expand our understanding of the basis of *KISS1* and *GPR54* genes in PCOS.

The *KISS1* gene encodes kisspeptin that signals directly to the GnRH neurons through the action on the kisspeptin receptor (*GPR54*) to release GnRH into the portal circulation, which in turn stimulates the secretion of LH and FSH from the gonadotrophs of the anterior pituitary by binding to its receptor GnRHR-1. LH and FSH act on gonads (by binding to their receptors LHCGR and

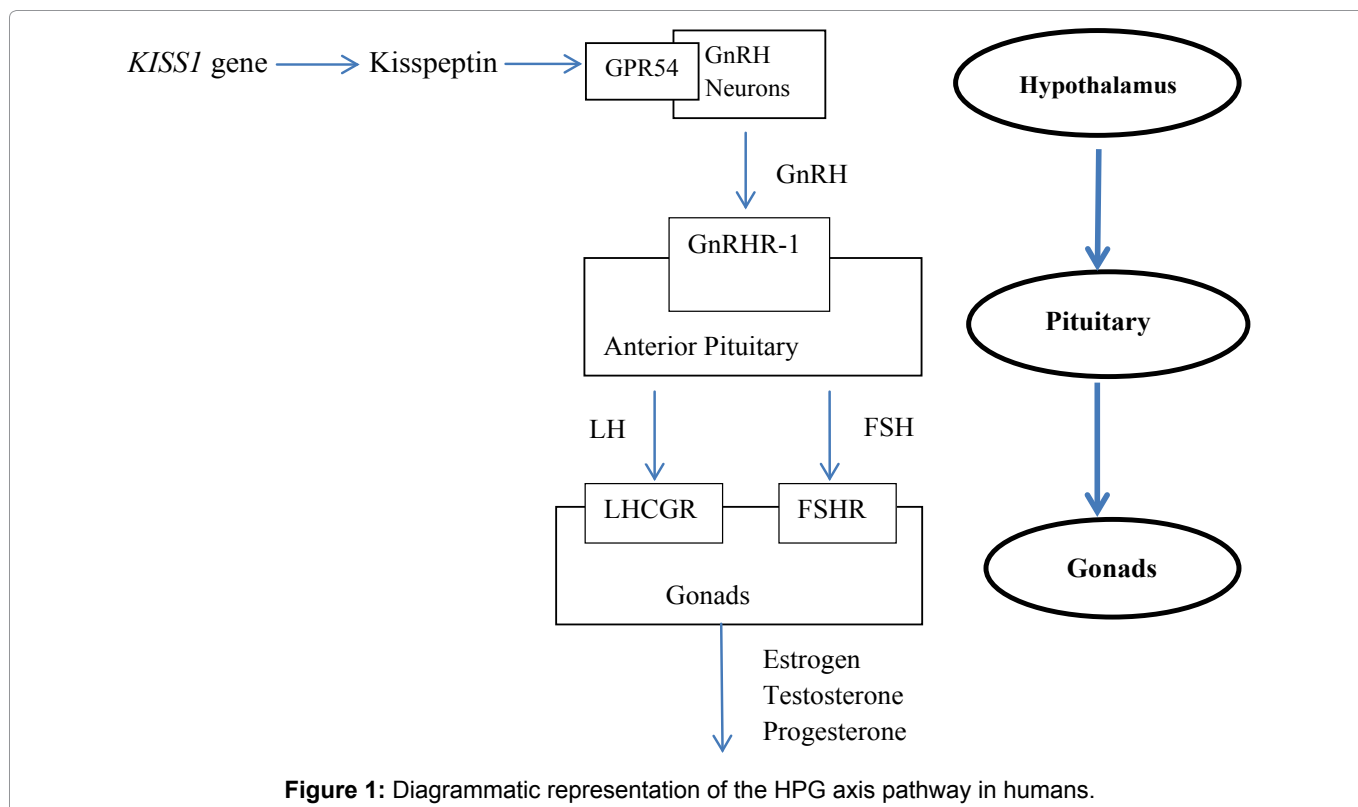


Figure 1: Diagrammatic representation of the HPG axis pathway in humans.

FSHR respectively) and stimulate the release of estrogen testosterone and progesterone (Figure 1).

GnRH and GnRHR Gene

The *GnRH* and its receptor (*GnRHR*) genes are important regulators of the HPG axis, and abnormalities in their function lead to impaired pubertal development and sexual maturation [21].

GnRH is a neurohormone consisting of 10 amino acids (decapeptide) that is produced in the arcuate nuclei of the hypothalamus [22]. GnRH stimulates the secretion of the two gonadotropins-LH and FSH by the anterior pituitary gland which stimulate spermatogenesis, folliculogenesis and ovulation. In addition, gonadal steroid hormones androgen, estrogen and progesterone participate in the negative feedback loop and inhibit GnRH and gonadotropin expression [23]. Four different *GnRHs* are reported to be expressed in various mammals. Of the four *GnRHs*, only *GnRHI* (mammalian *GnRH*) and *GnRHII* (chicken *GnRH II*) genes were identified in the human genome [24].

The *GnRH1* gene is located on chromosome 8p21.2. It spans about 5 kb and contains 3 exons. It encodes the GnRH1 precursor, which contains 92 amino acids, and it is subsequently processed to *GnRH1*, an active decapeptide. GnRH is the principal hormone regulating the pituitary gonadotropins, there by affecting the ovarian physiology. Lack of negative feedback regulation on GnRH pulse frequencies, can lead to excess secretion of LH; which in turn increases androgen biosynthesis in ovarian theca cells and results in hyperandrogenism, a key etiological factor in the pathogenesis of anovulation and infertility in PCOS women [25].

The *GnRHR* gene is located on chromosome 4q13.2. Its genomic sequence covers about 19 kb and it includes 3 exons. *GnRHR* gene encodes the receptor for GnRH1 hormone. This receptor is a member of the seven-transmembrane, G-protein Coupled Receptor (GPCR) family. It is expressed on the surface of pituitary gonadotrope cells as well as lymphocytes, breast, ovary, and prostate. Following binding of GnRH1, the receptor associates with G-proteins that activate phosphatidylinositol-calcium second messenger system and activation of the receptor leads to the secretion of LH and FSH [23].

So far, no major defects within *GnRH1* and *GnRHR* genes have been found in association with PCOS. However, a polymorphism in the first exon of *GnRH1* has been described, constituting an amino acid variation at codon 16 (Trp16Ser). A study by Valkenburg, et al. [26] showed distribution of the Trp16Ser alleles of *GnRH1* was similar in PCOS cases and controls and failed to identify any association with PCOS. Therefore, we can conclude that *GnRH1* and *GnRHR* genes mutations are uncommon in subjects with PCOS.

FSH and FSHR Genes

Follicle Stimulating Hormone (FSH) is a glycoprotein secreted by the anterior pituitary. It is a heterodimer consisting of common α -subunit and a hormone-specific β -subunit that contributes to the receptor binding specificity [27]. FSH secretion is regulated by GnRH and in turn regulates gonadal functions in males and females by activating their cognate receptors. In women, it plays a crucial role in the follicle development, oocyte maturation, steroidogenesis regulation, proliferation of granulosa cells and induces synthesis of the androgen-converting enzyme aromatase [28].

The effect of FSH is mediated by binding to its receptor - FSHR (Follicle Stimulating Hormone Receptor), which is specifically situated on the granulosa cells of the ovary [29]. *FSHR* gene is located on chromosome 2p21 and comprises 10 exons and 9 introns.

The *FSH* and *FSHR* genes are necessary for female fertility. The importance of FSHR in the signaling transmission of FSH made *FSHR* gene one of the important candidate genes for PCOS [30]. More than 900 SNPs in the *FSH* and *FSHR* genes have been reported [31]. Mutations in *FSHR* gene can lead to arrest of follicle development at several phases of growth [32,33]. Genetic variants in the *FSHR* gene may have effects on the phenotype. These effects include variable development of secondary sex characteristics, primary amenorrhea, hypoplastic ovary, and high serum FSH levels [34,35]. The two most clinically relevant polymorphisms of *FSHR* gene are in exon 10. One polymorphism is located at codon 307 in the extracellular domain of the receptor, where alanine is replaced by threonine (A307T; rs6165). The other polymorphism is in the intracellular domain at codon 680, where asparagine is replaced by serine (N680S; rs6166) [34,36].

The association between *FSHR* gene polymorphisms and PCOS were examined by several studies but the results were contradictory. Gu, et al. reported Ser680Asn of *FSHR* gene polymorphism was associated with PCOS in Korean women, whereas the Ala307Thr was not [37]. Meanwhile Dolfin, et al. showed that the Ala307Thr of *FSHR* polymorphism was related to PCOS in Italian women [38]. In addition, Unsal, et al. found that the genotype frequencies of the Ala307Thr and Ser680Asn polymorphisms of *FSHR* were not different between cases and controls in Turkish adolescent girls [39]. Sudo, et al. reported a significant increase in the Ala307Thr frequency among Japanese women with PCOS when compared to normal subjects [40]. A significant association between the polymorphism Ala307Thr and PCOS was also reported by a recent study on Chinese women [41]. However, Mohiyiddeen, et al. did not find any association between the Ser680Asn polymorphism of *FSHR* gene and PCOS in a British population [42]. In addition, Valkenburg, et al. concluded that *FSHR* gene variants were strongly associated with the severity of PCOS clinical features, but not with disease risk [26]. Orio, et al. reported no significant relationship between various polymorphisms of *FSHR* gene and PCOS in Italian females [43]. Contradictory findings of the different studies may be due to the variation in sample size and ethnicity of the study population.

Importantly, most studies have focused on association of *FSHR* gene polymorphism with PCOS, whereas association of *FSH β* gene polymorphism with PCOS is less explored. Tong, et al. concluded that *FSH β* gene mutations were found to be uncommon in patients with PCOS. However, Accl polymorphism was found to be associated with the syndrome in some women, especially those with obesity and hyperandrogenism [44].

LHB and *LHCGR* Genes

Luteinizing Hormone (LH) is a member of the glycoprotein hormone family that includes human Chorionic Gonadotropin (hCG), FSH, and Thyroid Stimulating Hormone (TSH). They are α/β heterodimers with a common α -subunit and a unique β -subunit. The β -subunit confers biologic specificity [45,46]. The two LH subunits (α and β) are encoded by separate genes. The α subunit gene is in chromosome 6 and the β subunit gene is in chromosome 19 [47]. LH stimulates follicular development, steroidogenesis, and the formation of corpus luteum [48], and ovulation results from a surge in LH levels [49]. LH acts by binding to its high affinity receptor known as Luteinizing Hormone/Choriogonadotropin receptor/Choriogonadotrophin receptor (LHCGR) [48]. LHCGR is a G protein-coupled hormone receptor and is expressed in numerous tissues including the gonads, uterus [50], fallopian tubes [51], placenta and fetus [52]. Both LH and hCG are endogenous ligands for LHCGR [53]. LHCGR is encoded by LHCGR gene located on chromosome 2p21 and composed of 11 exons occupying about 700 kbp [54].

The relationship between LH signaling pathway and PCOS has not been clearly understood. However, abnormal LH signaling is believed to play a significant role in augmenting ovarian androgen production in PCOS leading to anovulation [55]. *LHB* and *LHCGR* gene mutations may change the structure or function of the LH and LHCGR, either activating or inactivating their bioactivity, which cause anovulation, amenorrhea, and polycystic ovaries in women [56]. Several studies have proven the genetic associations of *LHB* and *LHCGR* polymorphisms with PCOS, although the results of different populations and loci of polymorphisms showed inconsistencies.

There are 2 common mutations of *LHB* gene that were associated with PCOS; one in codon 8 and other in codon 15. Point mutation in codon 8 causes amino acid replacements from Trp to Arg; and in codon 15 from Ile to Thr [57]. Kurioka, et al. showed that these mutations were considerably associated with PCOS [58] and Tapanainen, et al. concluded that the presence of these mutations may help to diagnose the risk for PCOS particularly among obese women [59]. However, studies in a British population revealed that the incidence was not higher in women with PCOS, though it was increased in obese women with PCOS [60]. In contrast, Huhtaniemi, et al. reported that variant LH occurs with normal frequency in non-obese patients with PCOS but is under-represented in obese patients with PCOS [61]. These results show that the clinical significance of the variant LH with respect to PCOS is contradictory. The same mutations were reported from Finland [62] and a similar LH form was described from Japan [57], which suggests that this variant LH represents a universal polymorphism [62]. Worldwide carrier frequency of this common genetic variant has been analyzed and reported as a prevalence of 7% in U.S. Hispanics, 18% in England and 42% in Lapps of northern Finland [60,62,63]. However,

these studies also reported that the LH variant is less common in Asian countries.

The *LHCGR* gene has at least 300 known polymorphisms [45,64]. A study by Capalbo, et al. on S312N (G935A) polymorphism in exon 10 of the *LHCGR* gene revealed that this variant is strongly linked with PCOS in the Sardinian population [54]. The finding of this study was supported by two other studies by Ha, et al. [65] and Bassiouny, et al. [66]. They found that the G935A polymorphism of *LHCGR* gene is associated with PCOS in Hui Chinese and Egyptian populations. On the contrary, Almawi, et al. [67] and Valkenburg, et al. [26] reported that G935A polymorphism was not overtly associated with PCOS in Bahraini Arab and Caucasian populations. Furthermore, Thathapudi, et al. revealed that the GG genotype, rather than AA, conferred a significant risk for developing PCOS in South Indian women [68].

Eriksen, et al. found an association with rs13405728 variant in the *LHCGR* gene and PCOS in white Europeans of

Danish origin and have suggested that the gene products of the *LHCGR* gene is linked with the diagnosis of PCOS, despite ethnicity [69-71]. Valkenburg, et al. found that *LHCGR* 18insLQ insertion allele frequency was significantly lower in white Europeans with PCOS than controls [26].

Table 1 summarizes the major findings of the candidate genes of the HPG axis and their association with PCOS.

Conclusions

PCOS appears to be a multigenic trait, although the contributing genes broadly remain undefined. This review explored in-depth the possible associations between PCOS and genetic polymorphisms of the many genes linked to the HPG axis and found the reported associations to be conflicting. The discrepancy of findings is likely to be due to variations in study design, sampling technique and sample size, along with demographic and genetic differences among the study populations.

Table 1: Comparison of genotype associations with PCOS.

Gene	SNP	Mutation	A.A change	Sample size	Association (Yes/No)	Population
<i>KISS1</i>	rs4889	C/G	Pro → Arg	PCOS = 28 Controls = 30	Yes	Saudi Arabian Albalawi FS, et al. [70]
<i>GnRH1</i>	rs6185	G/C	Trp → Ser	PCOS = 518 Controls = 2996	No	Caucasian Valkenburg O, et al. [26]
<i>FSHR</i>	rs6165	G/A	Ala → Thr	PCOS = 235 Controls = 128	No	Korean Gu BH, et al. [37]
	rs6165			PCOS = 44 Controls = 50	No	Turkish Unsal T, et al. [39]
	rs6165			PCOS = 40 Controls = 66	Yes	Italian Dolfin E, et al. [38]
	rs6165			PCOS = 96 Controls = 426	Yes	Japanese Sudo S, et al. [40]
	rs6165			POF = 40 PCOS = 60 Controls = 90	Yes	Chinese Du J, et al. [41]
<i>FSHR</i>	rs6166	G/A	Ser → Asn	PCOS = 44 Controls = 50	No	Turkish Unsal T, et al. [39]
	rs6166			PCOS = 58 Controls = 80	No	UK Mohiyiddeen L, et al. [42]
	rs6166			PCOS = 235 Controls = 128	Yes	Korean Gu BH, et al. [37]
<i>FSHR</i>	Various polymorphism			PCOS = 50 Controls = 50	No	Italian Orio F Jr, et al. [43]
<i>FSHB</i>	rs6169	C/T	Tyr → Tyr	PCOS = 135 Controls = 105	No	Singapore Chinese Tong Y, et al. [44]
<i>LHB</i>	rs1800447/ rs3449826	T/C A/G	Trp → Arg Ile → Thr	PCOS = 130 Controls = 96	Associated with ↑ Testosterone levels	Brazilian Batista MC, et al. [71]
<i>LHCGR</i>	rs2293275	A/G	Asn → Ser	PCOS = 518 Controls = 2996	No	Caucasian Valkenburg O, et al. [26]
	rs2293275			PCOS = 198 Controls = 187	Yes	Sardinian Capalbo A, et al. [54]
	rs2293275			PCOS = 100 Controls = 60	Yes	Egyptian Bassiouny YA, et al. [66]

We propose the need to analyze polymorphisms in multiple candidate genes of PCOS to determine its exact genetic basis. Development of a genetic diagnostic tool that would help in screening multiple candidate genes simultaneously is suggested. Such a tool would also help elucidate which of these SNPs are present in the different phenotypic subgroups of affected women. Such an approach would help foster a better understanding of the genetic basis for the pathophysiology underlying PCOS in different subgroups and populations. This knowledge could then be leveraged to devise the most optimal screening and effective management strategy for an affected woman, depending on her phenotypic subgroup and ethnicity.

Funding

Our research is funded (financial and material support) by National Research Council of Sri Lanka (NRC grant no: 15-149).

Conflict of Interest

None.

Disclosure

No conflict of interest.

References

- Zawadski JK, Dunaif A (1992) Diagnostic criteria for polycystic ovary syndrome: towards a rational approach. *Polycystic Ovary Syndrome*. Boston: Blackwell Scientific Publications, 377-384.
- Franks S (1995) Polycystic ovary syndrome. *N Engl J Med* 333: 853-861.
- Rotterdam ESHRE/ASRM-Sponsored PCOS Consensus Workshop Group (2004) Revised 2003 consensus on diagnostic criteria and long-term health risks related to polycystic ovary syndrome. *Fertil Steril* 81: 19-25.
- Dumesic DA, Abbott DH, Eisner JR, Herrmann RR, Reed JE, et al. (1998) Pituitary desensitization to gonadotropin-releasing hormone increases abdominal adiposity in hyperandrogenic anovulatory women. *Fertil Steril* 70: 94-101.
- Eisner JR, Barnett MA, Dumesic DA, Abbott DH (2002) Ovarian hyperandrogenism in adult female rhesus monkeys exposed to prenatal androgen excess. *Fertil Steril* 77: 167-172.
- Abbott DH, Dumesic DA, Franks S (2002) Developmental origin of polycystic ovary syndrome - a hypothesis. *J Endocrinol* 174: 1-5.
- Moran LJ, Pasquali R, Teede HJ, Hoeger KM, Norman RJ (2009) Treatment of obesity in polycystic ovary syndrome: A position statement of the Androgen Excess and Polycystic Ovary Syndrome Society. *Fertil Steril* 92: 1966-1982.
- Morreale EHF, Ramirez LM, Millan JLS (2005) The Molecular-Genetic Basis of Functional Hyperandrogenism and the Polycystic Ovary Syndrome. *Endocr Rev* 26: 251-282.
- Diamanti-Kandaraki E, Piperi C, Spina J, Argyrakopoulou G, Papanastasiou L, et al. (2006) Polycystic ovary syndrome: the influence of environmental and genetic factors. *Hormones (Athens)* 5: 17-34.
- Xu Y, Li Z, Ai F, Chen J, Xing Q, et al. (2015) Systematic Evaluation of Genetic Variants for Polycystic Ovary Syndrome in a Chinese Population. *PLoS One* 10: e0140695.
- Dunaif A (2016) Perspectives in Polycystic Ovary Syndrome: From Hair to Eternity. *J Clin Endocrinol Metab* 101: 759-768.
- Jakubowski L (2005) Genetic aspects of polycystic ovary syndrome. *Endokrynol Pol* 56: 285-293.
- Casarini L, Brigante G (2014) The polycystic ovary syndrome evolutionary paradox: a genome-wide association studies-based, in silico, evolutionary explanation. *J Clin Endocrinol Metab* 99: 2412-2420.
- Lee DK, Nguyen T, O'Neill GP, Cheng R, Liu Y, et al. (1999) Discovery of a receptor related to the galanin receptors. *FEBS Lett* 446: 103-107.
- Oakley AE, Clifton DK, Steiner RA (2009) Kisspeptin signaling in the brain. *Endocr Rev* 30: 713-743.
- d'Anglemont de Tassigny X, Colledge WH (2010) The role of kisspeptin signaling in reproduction. *Physiology (Bethesda)* 25: 207-217.
- West A, Vojta PJ, Welch DR, Weissman BE (1998) Chromosome localization and genomic structure of the KiSS-1 metastasis suppressor gene (KISS1). *Genomics* 54: 145-148.
- Lee JH, Miele ME, Hicks DJ, Phillips KK, Trent JM, et al. (1996) KiSS-1, a novel human malignant melanoma metastasis-suppressor gene. *J Natl Cancer Inst* 23: 1731-1737.
- Seminara SB, Messager S, Chatzidaki EE, Thresher RR, Acierno JS Jr, et al. (2003) The GPR54 gene as a regulator of puberty. *N Engl J Med* 349: 1614-1627.
- Clarke IJ, Cummins JT (1985) GnRH pulse frequency determines LH pulse amplitude by altering the amount of releasable LH in the pituitary glands of ewes. *J Reprod Fertil* 73: 425-431.
- Achermann JC, Ozisik G, Meeks JJ, Jameson JL (2002) Genetic causes of human reproductive disease. *J Clin Endocrinol Metab* 87: 2447-2454.
- Schally AV, Arimura A, Baba Y, Nair RM, Matsuo H, et al. (1971) Isolation and properties of the FSH and LH-releasing hormone. *Biochem Biophys Res Commun* 43: 393-399.
- Kaiser UB, Conn PM, Chinn WW (1997) Studies of gonadotropin-releasing hormone (GnRH) action using GnRH receptor-expressing pituitary cell lines. *Endocr Rev* 18: 46-70.
- Neill JD (2002) GnRH and GnRH receptor genes in the human genome. *Endocrinology* 143: 737-743.
- Taylor AE, McCourt B, Martin KA, Anderson EJ, Adams JM, et al. (1997) Determinants of abnormal gonadotropin secretion in clinically defined women with polycystic ovary syndrome. *J Clin Endocrinol Metab* 82: 2248-2256.
- Valkenburg O, Uitterlinden AG, Piersma D, Hofman A, Themmen AP, et al. (2009) Genetic polymorphisms of GnRH and gonadotrophic hormone receptors affect the phenotype of polycystic ovary syndrome. *Hum Reprod* 24: 2014-2022.
- Pierce JG, Parsons TF (1981) Glycoprotein hormones: structure and function. *Annu Rev Biochem* 50: 465-495.
- Simoni M, Nieschlag E (1995) FSH in therapy physiological basis, new preparations and clinical use. *Reprod Med Rev* 4: 163-167.
- Simoni M, Gromoll J, Nieschlag E (1997) The follicle stimulating hormone receptor: biochemistry, molecular biology, physiology, and pathophysiology. *Endocrine Reviews* 18: 739-773.

30. Fu L, Zhang Z, Zhang A, Xu J, Huang X, et al. (2013) Association study between FSHR Ala307Thr and Ser680Asn variants and polycystic ovary syndrome (PCOS) in Northern Chinese Han women. *J Assist Reprod Genet* 30: 717-721.
31. Simoni M, Tuttelmann F, Michel C, Bockenfeld Y, Nieschlag E, et al. (2008) Polymorphisms of the luteinizing hormone/chorionic gonadotropin receptor gene: association with maldescended testes and male infertility. *Pharmacogenet Genomics* 18: 193-200.
32. Kuechler A, Hauffa BP, Koninger A, Kleinau G, Albrecht B, et al. (2010) An unbalanced translocation unmasks a recessive mutation in the follicle-stimulating hormone receptor (FSHR) gene and causes FSH resistance. *Eur J Hum Genet* 18: 656-661.
33. Casarini L, Pignatti E, Simoni M (2011) Effects of polymorphisms in gonadotropin and gonadotropin receptor genes on reproductive function. *Rev Endocr Metab Disord* 12: 303-321.
34. Aittomaki K, Lucena JL, Pakarinen P, Sistonen P, Tapanainen J, et al. (1995) Mutation in the follicle-stimulating hormone receptor gene causes hereditary hypergonadotropic ovarian failure. *Cell* 82: 959-968.
35. Achrekar SK, Modi DN, Desai SK, Mangoli VS, Mangoli RV, et al. (2009) Follicle-stimulating hormone receptor polymorphism (Thr307Ala) is associated with variable ovarian response and ovarian hyperstimulation syndrome in Indian women. *Fertil Steril* 91: 432-439.
36. Gromoll J, Simoni M (2005) Genetic complexity of FSH receptor function. *Trends Endocrinol Metab* 16: 368-373.
37. Gu BH, Park JM, Baek KH (2010) Genetic variations of follicle stimulating hormone receptor are associated with polycystic ovary syndrome. *Int J Mol Med* 26: 107-112.
38. Dolfen E, Guani B, Lussiana C, Mari C, Restagno G, et al. (2011) FSH receptor Ala307Thr polymorphism is associated to polycystic ovary syndrome and to a higher responsiveness to exogenous FSH in Italian women. *J Assist Reprod Genet* 28: 925-930.
39. Unsal T, Konac E, Yesilkaya E, Yilmaz A, Bideci A, et al. (2009) Genetic polymorphisms of FSHR, CYP17, CYP11A1, CAPN10, INSR, SERPINE1 genes in adolescent girls with polycystic ovary syndrome. *J Assist Reprod Genet* 26: 205-216.
40. Sudo S, Kudo M, Wada S, Sato O, Hsueh AJ, et al. (2002) Genetic and functional analyses of polymorphisms in the human FSH receptor gene. *Mol Hum Reprod* 8: 893-899.
41. Du J, Zhang W, Guo L, Zhang Z, Shi H, et al. (2010) Two FSHR variants, haplotypes and meta analysis in Chinese women with premature ovarian failure and polycystic ovary syndrome. *Mol Genet Metab* 100: 292-295.
42. Mohiyiddeen L, Salim S, Mulugeta B, McBurney H, Newman WG, et al. (2012) PCOS and peripheral AMH levels in relation to FSH receptor gene single nucleotide polymorphisms. *Gynecol Endocrinol* 28: 375-377.
43. Orio F Jr, Ferrarini E, Cascella T, Dimida A, Palomba S, et al. (2006) Genetic analysis of the follicle stimulating hormone receptor gene in women with polycystic ovary syndrome. *J Endocrinol Invest* 29: 975-982.
44. Tong Y, Liao WX, Roy AC, Ng SC (2000) Association of AccI polymorphism in the follicle-stimulating hormone beta gene with polycystic ovary syndrome. *Fertil Steril* 74: 1233-1236.
45. Themmen AP (2005) An update of the pathophysiology of human gonadotrophin subunit and receptor gene mutations and polymorphisms. *Reproduction* 130: 263-274.
46. Mutharasan P, Galdones E, Peñalver Bernabé B, Garcia OA, Jafari N, et al. (2013) Evidence for chromosome 2p16.3 polycystic ovary syndrome susceptibility locus in affected women of European ancestry. *J Clin Endocrinol Metab* 98: 185-190.
47. Fiddes JC, Talmadge K (1984) Structure, expression, and evolution of the genes for the human glycoprotein hormones. *Recent Prog Horm Res* 40: 43-78.
48. Dufau ML (1998) The luteinizing hormone receptor. *Annu Rev Physiol* 60: 461-496.
49. Ascoli M, Fanelli F, Segaloff DL (2002) The lutropin/choriogonadotropin receptor, a 2002 perspective. *Endocr Rev* 23: 141-174.
50. Ziecik AJ, Kaczmarek MM, Blitek A, Kowalczyk AE, Li X, et al. (2007) Novel biological and possible applicable roles of LH/hCG receptor. *Mol Cell Endocrinol* 269: 51-60.
51. Gawronska B, Paukku T, Huhtaniemi I, Wasowicz G, Ziecik AJ (1999) Oestrogen-dependent expression of LH/hCG receptors in pig Fallopian tube and their role in relaxation of the oviduct. *J Reprod Fertil* 115: 293-301.
52. Perrier d'Hauterive S, Berndt S, Tsampalas M, Charlet-Renard C, Dubois M, et al. (2007) Dialogue between blastocyst hCG and endometrial LH/hCG receptor: which role in implantation? *Gynecol Obstet Invest* 64: 156-160.
53. Hearn MT, Gomme PT (2000) Molecular architecture and biorecognition processes of the cystine knot protein superfamily: part I. The glycoprotein hormones. *J Mol Recognit* 13: 223-278.
54. Capalbo A, Sagnella F, Apa R, Fulghesu AM, Lanzone A, et al. (2012) The 312N variant of the luteinizing hormone/choriogonadotropin receptor gene (LHCGR) confers up to 2.7-fold increased risk of polycystic ovary syndrome in a Sardinian population. *Clin Endocrinol* 77: 113-119.
55. Balen AH (1993) Hypersecretion of luteinizing hormone and the polycystic ovary syndrome. *Hum Reprod* 8: 123-128.
56. Huhtaniemi IT, Themmen AP (2005) Mutations in human gonadotropin and gonadotropin-receptor genes. *Endocrine* 26: 207-217.
57. Furui K, Suganuma N, Tsukahara S, Asada Y, Kikkawa F, et al. (1994) Identification of two point mutations in the gene coding luteinizing hormone (LH) b-subunit, associated with immunologically anomalous LH variants. *J Clin Endocrinol Metab* 78: 107-113.
58. Kurioka H, Takahashi K, Irikoma M, Okada M, Ozaki T, et al. (1999) Diagnostic difficulty in polycystic ovary syndrome due to an LH-beta-subunit variant. *Eur J Endocrinol* 140: 235-238.
59. Tapanainen JS, Koivunen R, Fauser BC, Taylor AE, Clayton RN, et al. (1999) A new contributing factor to polycystic ovary syndrome: The genetic variant of luteinizing hormone. *J Clin Endocrinol Metab* 84: 1711-1715.
60. Rajkhowa M, Talbot JA, Jones PW, Pettersson K, Haavisto AM, et al. (1995) Prevalence of an immunological LH beta-subunit variant in a UK population of healthy women and women with polycystic ovary syndrome. *Clin Endocrinol (Oxf)* 43: 297-303.
61. Huhtaniemi I, Pettersson K, Haavisto AM, Anttila L, Jaatinen T, et al. (1994) A variant of luteinizing hormone due to polymorphism in its b-subunit gene occurs with normal frequency in nonobese, but is underrepresented in obese patients with polycystic ovarian syndrome [abstract]. *Eur J Endocrinol* 130: 65.
62. Haavisto AM, Pettersson K, Bergendahl M, Virkamäki A,

- Huhtaniemi I (1995) Occurrence and biological properties of a common genetic variant of luteinizing hormone. *J Clin Endocrinol Metab* 80: 1257-1263.
63. Nilsson C, Pettersson K, Millar RP, Coerver KA, Matzuk MM, et al. (1997) Worldwide frequency of a common genetic variant of luteinizing hormone: an international collaborative research. *Fertil Steril* 67: 998-1004.
64. Atger M, Misrahi M, Sar S, Le Flem L, Dessen P, et al. (1995) Structure of the human luteinizing hormone-choriogonadotropin receptor gene: unusual promoter and 5' non-coding regions. *Mol Cell Endocrinol* 111: 113-123.
65. Ha L, Shi Y, Zhao J, Li T, Chen ZJ (2015) Association study between polycystic ovarian syndrome and the susceptibility genes polymorphisms in Hui Chinese women. *PLoS One* 10: e0126505.
66. Bassiouny YA, Rabie WA, Hassan AA, Darwish RK (2014) Association of the luteinizing hormone/choriogonadotropin receptor gene polymorphism with polycystic ovary syndrome. *Gynecol Endocrinol* 30: 428-430.
67. Almawi WY, Hubail B, Arekat DZ, Al-Farsi SM, Al-Kindi SK, et al. (2015) Luteinizing hormone/choriogonadotropin receptor and follicle stimulating hormone receptor gene variants in polycystic ovary syndrome. *J Assist Reprod Genet* 32: 607-614.
68. Thathapudi S, Kodati V, Erukkambattu J, Addepally U, Qurratulain H (2015) Association of Luteinizing Hormone Chorionic Gonadotropin Receptor Gene Polymorphism (rs2293275) with Polycystic Ovarian Syndrome. *Genet Test Mol Biomarkers* 19: 128-132.
69. Eriksen MB, Brusgaard K, Andersen M, Tan Q, Altinok ML, et al. (2012) Association of polycystic ovary syndrome susceptibility single nucleotide polymorphism rs2479106 and PCOS in Caucasian patients with PCOS or hirsutism as referral diagnosis. *Eur J Obstet Gynecol Reprod Biol* 163: 39-42.
70. Albalawi FS, Daghestani MH, Daghestani MH, Eldali A, Warsy AS (2018) rs4889 polymorphism in KISS1 gene, its effect on polycystic ovary syndrome development and anthropometric and hormonal parameters in Saudi women. *J Biomed Sci* 25: 50.
71. Batista MC, Duarte Ede F, Borba MD, Zingler E, Mangusi-Gomes J, et al. (2014) Trp28Arg/Ile35Thr LHB gene variants are associated with elevated testosterone levels in women with polycystic ovary syndrome. *Gene* 550: 68-73.