A Case of Dedifferentiated Leiomyosarcoma of the Uterus
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Abstract
Dedifferentiation of leiomyosarcoma is a rare phenomenon that associates pleomorphic histology and loss of smooth muscle differentiation. Although the leiomyosarcoma is well known sarcoma in the uterus, and the dedifferentiated leiomyosarcoma is well recognized in the soft parts, there are only a few reports of dedifferentiation of leiomyosarcoma in the uterus. Herein we present a case of dedifferentiated uterine leiomyosarcoma and discuss its relation to the undifferentiated uterine sarcoma.

Keywords
Uterine leiomyosarcoma, Dedifferentiation, Undifferentiated uterine sarcoma, CD10

Abbreviations
MRI: Magnetic Resonance Imaging; SMA: Smooth Muscle Actin; CEA: Carcinoembryonic Antigen; ER: Estrogen Receptor; PgR: Progesterone Receptor

Introduction
Leiomyosarcoma is relatively rare disease of the uterine corpus accounting for 1-2% of all malignancies according to the World Health Organization (WHO) classification of tumours of female reproductive organs [1]. On the other hand, leiomyosarcoma is also important sarcoma of extra-uterine site and actually major sarcoma in the retroperitoneal and the pelvic areas [2]. Dedifferentiation is a rare phenomenon of soft tissue sarcomas including leiomyosarcoma and is extremely rare among uterine sarcomas [2-8]. Dedifferentiated tumors often exhibit pleomorphic high-grade histology resembling pleomorphic undifferentiated soft tissue sarcoma (USTS) that has been known as pleomorphic malignant fibrous histiocytoma (MFH) in the past [5,6,9,10]. Dedifferentiation is also demonstrated by loss of lineage-specific marker expression by immunohistochemistry. The uterine high grade sarcomas with unknown lineage are called as undifferentiated uterine sarcomas (UUS) and often demonstrate similar morphology with USTS. We present a case of uterine leiomyosarcoma with dedifferentiation and it’s highly pleomorphic dedifferentiated area showed reduced expression of smooth muscle markers.

Case Report
Clinical history
A 63-year-old multiparous female presented with bloody vaginal discharge lasting for 2 months. She had no particular medical history except for the uterine leiomyoma discovered 13 years before. Her cervical cytology was negative while endometrial cytology detected atypical cells. There was no elevation of tumor markers (CEA and CA19-9) in her serum. Hysteroscopy revealed a smooth-surfaced massive tumor occupying the uterine cavity. Abdominal MRI revealed 7.7 × 9.4 × 12.4 cm of round mass in the anterior uterine corpus. The lesion showed heterogeneous high intensity on T2 weighted image and was unevenly enhanced on dynamic contrast study. On the histological examination by biopsy, the tumor tissue was composed of spindle shaped atypical cells with blunt-ended nuclei and eosinophilic cytoplasm, arranged in irregular fascicular pattern intermingled with necrotic area. These tumor cells were positive for α-smooth muscle actin (α-SMA), calponin, desmin, estrogen receptor (ER) and progesterone receptor (PgR) but negative for CD10 by immunohistochemical (IHC) examination. MIB-1 index was around 50% and the tumor was diagnosed as leiomyosarcoma. Thereafter she was transferred to our hospital for further investigation and treatment, and second biopsy was performed. The tissue contained highly atypical cells including bizarre cells, multinucleated neoplastic giant cells and numerous mitotic figures against heavily necrotic backgrounds, rather resembled pleomorphic USTS or UUS than ordinary leiomyosarcoma observed in the previous biopsy. CT examination revealed multiple small nodules in bilateral lungs which were diagnosed as metastases. She underwent simple hysterectomy with bilateral salpingo-oophorectomy and has been receiving adjuvant chemotherapy.

Gross findings
Resected uterus was longitudinally cut in the anterior wall and roughly lobulated polypoid tumor was exposed. After fixation in 10% formalin, the tumor was 14 × 10 × 10 cm in maximum dimension and felt elastic soft (Figure 1a). Stem of the tumor was attached to the uterine fundus. On sagittal slice, proximal part of the tumor looked solid with whitish cut surface containing necrotic foci and hemorrhages (Figure 1b). The whitish area lacked whorl pattern...
Figure 1: Gross findings of the dedifferentiated leiomyosarcoma.

a) Resected uterus was cut longitudinally in the anterior wall and the tumor was exposed. The polypoid tumor was lobulated and attached to the uterine fundus; b) Sagittal slice of the tumor (above) and horizontal slice of right side (below). On the sagittal slice, the fascicular area (FA) is to the left and the pleomorphic area (PA) to the right. Dashed line denotes assumed borderline between these areas; borderline in central necrotic area was postulated from morphology of tumor cells with coagulation necrosis. A leiomyoma was seen in the posterior uterine wall (arrowhead).

Figure 2: Histological finding of the dedifferentiated leiomyosarcoma in hematoxylin-eosin section (Original magnification: 200x [a-c] and 100x [d]).

a) Fascicular area. Tumor cells were spindle with eosinophilic cytoplasm, arranged in fascicular pattern. There were scattered numerous mitotic figures including atypical ones; b) Pleomorphic area. Tumor cells were highly pleomorphic with abundant bizarre cells. Some area was rich in hyalinized stroma and contained many hyalinized vessels as well; c) Polygonal or epithelioid tumor cells were also seen in some area; d) Interface of each area is shown (arrows). The fascicular area is to the left side and the pleomorphic area composed of eosinophilic epithelioid cells to the right. Transition between these components was abrupt.
which is characteristic of leiomyoma. About a half of mass distal to the fundus was tan tinted, heavily necrotic and hemorrhagic. The posterior and the lateral uterine wall contained usual leiomyomas. No remarkable change was found in the uterine cervix and uterine appendages.

### Histological findings

Entire sagittal slice was cut into sections for histological assessment. Microscopically, the proximal part of the tumor (whitish area) was composed of atypical spindle cells with brunt-ended elongated nucleus and eosinophilic cytoplasm, demonstrating high mitotic activity, as many as 50 mitoses per 10 high power fields on average (Figure 2a). There were occasional multinucleated giant cells and mild anisokaryosis, but tumor cells were arranged in the fascicular manner and each cell sustained some resemblance to smooth muscle cells, indicating well-differentiated state. This fascicular part contained occasional geographical necrosis. On the other hand, lower part of tumor (tan tinted area) consisted of cells with marked nuclear atypia, often bizarre in shape. Polygonal large cells rich in pale eosinophilic cytoplasm and occasional multinucleated huge cells as well as broad spindle cells were arranged in irregular fascicle like pleomorphic USTS (Figure 2b and Figure 2c). Occasional storiform arrangement was observed. Some of this pleomorphic area contained numerous hyalinized small vessels. There were heavy necrosis and hemorrhagic foci. An abrupt transition between the fascicular and the pleomorphic area was observed (Figure 2d). No heterologous elements were detected.

### Immunohistochemical examination

A tissue section containing both fascicular and pleomorphic area was selected for IHC examinations. Formalin-fixed paraffin-embedded tissues were 4-µm-cut and placed onto silane coated glass slides. Deparaffinized and pretreated tissue sections were processed by automatic immunostaining instrument, Histostainer36A (Nichirei bioscience Inc., Tokyo, Japan) according to the manufacturer’s instructions. Results of IHC study are summarized in table 1 and shown in figure 3. IHC data in normal myometrium contained in the section is also shown in table 1 as either positive or negative controls. Fascicular area of tumor was positive for several smooth muscle markers (α-SMA, desmin, h-caldesmon, calponin), whereas pleomorphic area was diffusely positive for α-SMA and only focally positive for calponin. Regarding hormone receptors, fascicular area was positive only for ER or for PgR. Pleomorphic area was negative either for ER or for PgR. Pleomorphic area was diffusely positive for CD10 and p16, both of which were absent in fascicular area. p53 was positive in both areas, whereas cytokeratin AE1/AE3, S-100, c-kit and CD34 were negative in both areas. From these findings, we finally concluded that the lower pleomorphic part of this tumor represents dedifferentiation from leiomyosarcoma.

### Discussion

Among soft tissue tumors, dedifferentiation is infrequent but well-known phenomenon such as dedifferentiated liposarcoma and pleomorphic rhabdomyosarcoma [2,11]. However, the dedifferentiation of leiomyosarcomas has only been described as occasional phenomenon, without particular designation as a subtype in WHO classification of tumours soft tissues and bone [2] and in Armed Forces Institute of Pathology (AFIP) atlas of tumors of the Soft Tissues [11]. Whereas, the dedifferentiation of uterine leiomyosarcomas have not even been mentioned in WHO classification of tumours of female reproductive organs [1]. To our knowledge, there are only five case reports of uterine leiomyosarcoma with dedifferentiation (Table 2) [4,7,8]. Dedifferentiated leiomyosarcomas are composed of pleomorphic atypical cells with high mitotic rate and heavy necrosis, and often exhibit abrupt transition from

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**Table 1: Results of immunohistochemical examination.**

<table>
<thead>
<tr>
<th></th>
<th>Fascicular area</th>
<th>Pleomorphic area</th>
<th>Myometrium</th>
</tr>
</thead>
<tbody>
<tr>
<td>α-SMA</td>
<td>Positive</td>
<td>Positive</td>
<td>Positive</td>
</tr>
<tr>
<td>Desmin</td>
<td>Positive</td>
<td>Negative</td>
<td>Positive</td>
</tr>
<tr>
<td>h-caldesmon</td>
<td>Positive</td>
<td>Negative</td>
<td>Positive</td>
</tr>
<tr>
<td>Calponin</td>
<td>Positive</td>
<td>Positive (focal)</td>
<td>Positive</td>
</tr>
<tr>
<td>CD10</td>
<td>Negative</td>
<td>Positive</td>
<td>Negative</td>
</tr>
<tr>
<td>ER</td>
<td>Positive</td>
<td>Negative</td>
<td>Positive</td>
</tr>
<tr>
<td>PgR</td>
<td>Negative</td>
<td>Negative</td>
<td>Positive</td>
</tr>
<tr>
<td>EMA</td>
<td>Negative</td>
<td>Positive (focal)</td>
<td>Negative</td>
</tr>
<tr>
<td>AE1/AE3</td>
<td>Negative</td>
<td>Negative</td>
<td>Negative</td>
</tr>
<tr>
<td>S-100</td>
<td>Negative</td>
<td>Negative</td>
<td>Negative</td>
</tr>
<tr>
<td>Cyclin D1</td>
<td>Negative</td>
<td>Negative</td>
<td>Negative</td>
</tr>
<tr>
<td>β-catenin</td>
<td>Negative</td>
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</tr>
<tr>
<td>p53</td>
<td>Variable</td>
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<td>Negative</td>
</tr>
<tr>
<td>p16</td>
<td>Negative</td>
<td>Positive</td>
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</tr>
<tr>
<td>c-Kit</td>
<td>Negative</td>
<td>Negative</td>
<td>Negative</td>
</tr>
<tr>
<td>CD34</td>
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<td>Negative</td>
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</tr>
</tbody>
</table>

**Abbreviations:** SMA: Smooth Muscle Actin, ER: Estrogen Receptor, PgR: Progesterone Receptor, EMA: Epithelial Membrane Antigen.

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**Figure 3:** Immunohistochemical examination of fascicular and pleomorphic area. (Original magnification: 400x).

Results in the fascicular area (FA) are shown in the upper column and those in the pleomorphic area (PA) are shown in the lower one. In the pleomorphic area, cells showing positivity for h-caldesmon in the figure are vascular smooth muscle cells.
well differentiated areas. These dedifferentiated areas often resemble pleomorphic USTS [5,6,9,10]. Dedifferentiation is usually proved by complete loss of lineage-specific markers in immunohistochemistry, and partial preservation of smooth muscle markers like in our case is distinguished as pleomorphic leiomyosarcoma by some authors [6,10].

Dedifferentiated and pleomorphic leiomyosarcomas of soft parts have been shown to be highly aggressive tumors presenting 50% to 65.2% of mortality and 89% of incidence of metastasis [6,10]. It has also been reported that the loss of myogenic differentiation in leiomyosarcoma could be a significant prognostic factor, accounting for an aggressiveness of these tumors [9].

Oda, et al. reported that pleomorphic leiomyosarcoma accounts for 8.6% of all leiomyosarcoma [10], whereas Nicolas, et al. implied higher incidence, for such tumor with tiny differentiated area might have been misdiagnosed as pleomorphic MFH [6]. Just like Demicco, et al. assumed that many of USTSs would be actually dedifferentiated leiomyosarcomas [9], those tumors reported as uterine MFH or UUS might have been dedifferentiated leiomyosarcoma. Indeed, the pleomorphic area of the present case was hardly distinguishable from pleomorphic UUS, since UUS can be positive for CD10 and focally positive for myogenic markers [1].

UUS has now been considered to arise from heterogeneous precursors and to be a diagnosis of exclusion [12] like USTS in soft tissues, requiring extensive histopathological examination for final diagnosis. UUS is known to display transition from endometrial stromal tumors of lower grade, suggesting some UUS are dedifferentiated endometrial stromal tumors, which is also supported by experiments in genetic level [13]. Kurihara, et al. demonstrated that transcripts of JAZF1-JJAZ, a fusion gene detected in 50% of low-grade sarcoma shows CD10 expression than restricted phenomenon to Müllerian derived tumors. Therefore, it may be rather generalized event that high-grade sarcoma shows CD10 expression than restricted phenomenon to Müllerian derived tumors.

Consent

Written informed consent was obtained from the patient for publication of this Case Report and any accompanying images. A copy of the written consent is available for review by the Editor-in-Chief of this journal.

References


Table 2: Dedifferentiated leiomyosarcoma of the uterus.

<table>
<thead>
<tr>
<th>Year</th>
<th>Author</th>
<th>Age</th>
<th>Size</th>
<th>Therapy</th>
<th>Metastasis</th>
<th>Status (*)</th>
<th>SMA</th>
<th>Des</th>
<th>Calp</th>
<th>h-Cal</th>
<th>CD10</th>
<th>CK</th>
<th>ER</th>
<th>PgR</th>
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<td>8 cm</td>
<td>SH+USO</td>
<td>lung, liver</td>
<td>Dead(11m)</td>
<td>epithelioid cells, OLGC</td>
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<td>-</td>
<td>NA</td>
<td>NA</td>
<td>-</td>
<td>NA</td>
<td>-</td>
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<tr>
<td>2007</td>
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<td>48</td>
<td>14 cm</td>
<td>UAE+SH</td>
<td>lung, brain</td>
<td>Dead(20m)</td>
<td>bizarre cells</td>
<td>+</td>
<td>-</td>
<td>NA</td>
<td>+</td>
<td>NA</td>
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<td>-</td>
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<tr>
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<td>Chen [5]</td>
<td>59</td>
<td>5 cm</td>
<td>TBH+BSO</td>
<td>none</td>
<td>Dead(28m)</td>
<td>NA</td>
<td>+</td>
<td>NA</td>
<td>NA</td>
<td>NA</td>
<td>-</td>
<td>NA</td>
<td>-</td>
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<tr>
<td>2012</td>
<td>Chen [5]</td>
<td>60</td>
<td>7 cm</td>
<td>excision</td>
<td>NA</td>
<td>Dead(12m)</td>
<td>NA</td>
<td>+</td>
<td>NA</td>
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<td>NA</td>
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<tr>
<td>2012</td>
<td>Rawish [7]</td>
<td>48</td>
<td>16 cm</td>
<td>SH+BSO+CT</td>
<td>serosa, RP</td>
<td>Alive(8m)</td>
<td>osteoids</td>
<td>-</td>
<td>NA</td>
<td>-</td>
<td>NA</td>
<td>focal</td>
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<tr>
<td>2016</td>
<td>Nosaka [6]</td>
<td>63</td>
<td>12.4 cm</td>
<td>SH+CT</td>
<td>lung</td>
<td>Alive(9m)</td>
<td>bizarre cells</td>
<td>+</td>
<td>focal</td>
<td>-</td>
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