Histopathological Fate in the Inguinal Hernia Sac in the Children

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Abstract

Background/Purpose: Obliteration of the processus vaginalis during the fetal growth begins with a transient decrease in sympathetic tonus and then ends with smooth muscle cells (SMCs) undergoing apoptosis. Otherwise, an inguinal hernia (IH) occurs due to the defective obliteration. Although the mechanism in the formation of an inguinal hernia has been elucidated by many investigations, it has not been investigated whether proliferation in the IH sac cells which would lead to the benign or malignant process. In this study, we aimed to examine whether the proliferative changes in the cells of IH sac would be a histopathological precursor to any process.

Methods: We obtained fifty (male 25, female 25) samples of the IH sac from patients (study group) during operations. Control groups were formed from twelve parietal peritoneal samples (male 4, female 8) undergoing laparotomy for various reasons, and two processus vaginalis samples obtained from 2 boys. Samples were evaluated regarding mesothelial, smooth muscle, neuronal, adipose, and vascular tissue proliferation as well as inflammation and congestion. Histopathological evaluation was made with H&E staining. Immunohistochemical evaluation was performed with PCNA marker, respectively.

Results: There were significantly increased mesothelial, vascular, nerve, and SMC proliferation in the study groups compared with those of control groups (p < 0.05). The expression of HBME-1 and PCNA was significantly increased (p < 0.05) in control groups compared with those of controls.

Conclusions: We determined that the proliferation of SMCs increased in the IH sac as in previous studies. We also found the increased proliferation in the mesothelial cells together with the increased HBME-1 and PCNA expressions in the IH sac. These alterations may be an initiator of a benign or malignant process by which would be able to develop in the IH sac.

Keywords
Inguinal hernia, Sac, Proliferation, PCNA, HBME-1, Children

Introduction

Inguinal hernias in the children that require a semi-urgent operation due to the risk of incarceration can rarely lead to complications such as testis ischemia. From complications, the development of mesothelioma in a sac is an unusual process regarding the proliferation, which can be thought of as an endpoint during the developmental process of an IH sac [1-4].

Processus vaginalis (PV) forming with a peritoneal protrusion in the eighth week of pregnancy is wholly obliterated after the testis and ovary complete their descent to normal localization. If PV obliteration does not occur, it is inevitable that an inguinal hernia, hydrocele, and cord cysts or Nuck’s duct cyst form. PV obliteration occurs through apoptosis, namely programmed cell death. Smooth muscle cells (SMCs), which is a specific marker in the structure of fetal PV, are not found in both peritoneal area and PV in the postnatal period. Tanyel, et al. suggested that the obliteration of the PV takes place through a transient decrease in sympathetic tonus and an increase in parasympathetic tonus in this process [4-8].

During obliteration of PV, the intracellular CAMP increase that causes a trophic effect on SCMs via β-adrenergic receptor due to increased sympathetic tonus ceases critical apoptosis by preventing dedifferentiation of SCMs. Also, increased sympathetic tonus is associated with increased androgen effect, and increased androgens act trophic impact on SCMs. As the PV affected by deviations relating the programmed cell death in these complicated steps, this situation shows that PV is not just a simple peritoneal protrubance [9,10].

Tanyel, et al. asserted that myofibroblasts inducing...
apoptosis of SMCs in the fusion of mesothelial opposing layers in the PV lead to contribute to the obliteration [11]. In previous studies, immunohistochemical methods have been used to explain the indirect inguinal hernia mechanisms that occur in developmental aberrations in the PV [7-11]. The theories based on the absence of obliteration in the PV have focused on defective apoptosis through the persistence of fetal SMCs in the proliferative changes occurring in a hernia sac. Although the mechanism in the formation of an inguinal hernia has been elucidated by many investigations, it has not been investigated whether proliferation in the IH sac cells which would lead to the benign or malignant process. In this study, we aimed to examine whether the proliferative changes in the cells of IH sac would be a histopathological precursor to any process.

Materials and Methods

Study subjects

We obtained fifty (male 25, female 25) samples of the hernia sacs (study groups) from the one-sided indirect inguinal hernia operations between January 2016 and January 2018. Standard procedure was high ligation of the IH sac and ligation via monofilament absorbable sutures during operation. Control groups were formed from twelve parietal peritoneum samples which were obtained from patients (male 4, female 8) undergoing laparotomy for various reasons (two processus vaginalis samples obtained from 2 boys undergoing contra-lateral exploration were included in the male control group). The histological features of each group of samples were examined by a pathologist blinded to the specific pathology. The study was approved by the Institutional Ethics Review Board for Clinical Research (2018/24/4).

Histopathological evaluation

Immediately following the surgical procedure, specimens were placed in a neutral buffered formaldehyde solution for histopathological evaluation. Following the fixation of the examples, paraffin sections were stained with Masson trichrome (HT15; Sigma, Milan, Italy) and hematoxylin-eosin (H&E) for microscopic evaluation. H&E-stained sections of the control groups and the study groups were evaluated regarding mesothelial, smooth muscle, neuronal, adipose, and vascular tissue proliferation. All specimens were also examined concerning inflammation and congestion.

Immunohistochemical evaluation

For the comparative evaluation of the specimens through immunohistochemical, the glass slides were washed thoroughly with distilled water for 20 minutes and exposed to antigen for a total period of 20 minutes, with a one-minute break at five-minute intervals, in a target retrieval solution, which was diluted by 1/10. The solution was left for twenty minutes at room temperature and then washed with distilled water. It was then kept for five minutes in phosphate-buffered saline (PBS). Then, it was incubated for one hour with 1/50 diluted ready-for-use PCNA antibody (Code: SC-56, Santa Cruz Biotechnology, Inc.). The rabbit anti-human HBME-1 (Hector Battifora mesothelial antigen-1) polyclonal antibodies were used as a marker of the mesothelial cells (diluted 1:50; cat. no. MA5-12220; Thermo Fisher Scientific, USA). The procedure was carried out in strict accordance with the instructions of rabbit anti-human HBME-1 monoclonal antibody kit. After it was incubated for about 15 minutes in a biotin solution, it was kept for 10 minutes in PBS. Then, it was incubated for about 15 minutes in a streptavidin-peroxidase solution. It was kept for five minutes in AEC (3 amino-9-etylcarbazole) chromogen after it was kept for 10 minutes in PBS. It was washed with distilled water. Cross-sections were then kept for five minutes in Mayer’s hematoxylin for opposite staining. Afterward, it was washed with tap water and enclosed with a mounting medium (Entellan, Merck Millipore, Darmstadt, Germany).

For immunostaining, a fully automated IHC device (Leica Bond-Max, Melbourne, Australia) was used. Histopathological and immunohistochemical evaluations were carried out using a light microscope (Olympus BX53, Tokyo, Japan).

Staining score

In the histopathological evaluation; the severity of mesothelial, smooth muscle, vascular and adipose tissue proliferation as well as inflammation and congestion were scored: 0 score = no proliferation, 1 score = mild (+), 2 scores = moderate (+++) and, 3 scores = strong (+++) proliferation. Immunohistochemical evaluation showed nuclear staining especially in specimens stained with PCNA antibody. PCNA staining score was used for evaluating the proliferative activity of the mesothelial and other cells. Positive staining with HBME-1 was detected by yellow-brown granular staining, mostly in the cytoplasm and less in the cell membrane. No staining with nuclear membrane HBME-1 was observed. In the microscopic evaluation, 4 high-power fields were determined for each slice at the 4 corners and the center to calculate the proportion of positive cells of 100 sample cells; mean positive score was obtained from the 4 calculation. The percentage of positive cells was categorized on a scale of score 0 = no staining, score 1 = weak staining (less than 10% focal involvement), score 2 = moderate staining (11-50% regional involvement), and score 3 = strong staining (greater than 50% diffuse involvement) [12].

Statistical analysis

In all statistical analysis of the study, SPSS (version 16.0; SPSS Inc, Chicago, USA) was used. All parameters were stated as mean ± S.D. Scores with ordinal data were compared between the groups by the Mann-Whit-
There was an increased proliferation score in FIH group compared to FC, which were also statistically significant (p < 0.05). There was no significant difference between FIH and MIH except for neuronal tissue proliferation histopathologically when both groups were compared with each other. Namely, the neuronal staining score in males was higher than that of females (p = 0.00). The histopathological staining scores of the study (MIH, FIH) and the control (MC, FC) groups were shown in Table 1.

The expression of HBME-1 and PCNA (Figure 2) was significantly increased (p < 0.05) in the MIH and FIH groups compared with those of controls. The comparative evaluation of staining score regarding inflammation and congestion between the study and control groups showed significant variation (p < 0.05) in favor of the study groups (Figure 1), whereas there were no significant differences between MIH and FIH groups (p > 0.05). Histopathological (H&E; inflammation, and congestion) and immunohistochemical (HBME-1 and PCNA) staining scores of the study (MIH, FIH) and control (MC, FC) groups were shown in Table 2.

Discussion
The PV composes of a process of the peritoneum

Figure 1: Histological findings (H&E, original magnification X 200); a) Normal peritoneum consisting of a superficial mesothelial layer and a submesothelial zone; b) Inguinal sac tissue with increased mesothelial proliferation, vascularity, inflammation and congestion; c) Increased smooth muscle proliferation; d) Marked neuronal tissue.
Table 1: The histopathological findings with scores in the study (inguinal hernia sac samples) and control (PV and peritoneal samples) groups.

<table>
<thead>
<tr>
<th>Type of tissue proliferation</th>
<th>MC score, Mean ± SD (n = 6)</th>
<th>FC score, Mean ± SD (n = 8)</th>
<th>MIH score, Mean ± SD (n = 25)</th>
<th>FIH score, Mean ± SD (n = 25)</th>
<th>p, MIH vs. MC</th>
<th>p, FIH vs. FC</th>
<th>p, MIH vs. FIH</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mesothelial</td>
<td>1.16 ± 0.40</td>
<td>1.12 ± 0.35</td>
<td>1.80 ± 0.57</td>
<td>1.96 ± 0.73</td>
<td>0.016</td>
<td>0.005</td>
<td>0.069</td>
</tr>
<tr>
<td>Smooth muscle</td>
<td>1.33 ± 0.51</td>
<td>1.37 ± 0.51</td>
<td>2.32 ± 0.69</td>
<td>2.16 ± 0.68</td>
<td>0.005</td>
<td>0.008</td>
<td>0.013</td>
</tr>
<tr>
<td>Neuronal</td>
<td>0.66 ± 0.81</td>
<td>0.62 ± 0.74</td>
<td>2.44 ± 0.66</td>
<td>1.36 ± 0.63</td>
<td>0.001</td>
<td>0.015</td>
<td>0.000</td>
</tr>
<tr>
<td>Vascular</td>
<td>1.16 ± 0.40</td>
<td>1.25 ± 0.46</td>
<td>2.36 ± 0.70</td>
<td>2.40 ± 0.60</td>
<td>0.002</td>
<td>0.000</td>
<td>0.088</td>
</tr>
<tr>
<td>Adipose</td>
<td>0.66 ± 0.10</td>
<td>0.25 ± 0.11</td>
<td>0.80 ± 0.20</td>
<td>0.76 ± 0.13</td>
<td>0.026</td>
<td>0.010</td>
<td>0.735</td>
</tr>
</tbody>
</table>

MC: male control group; FC: female control group; MIH: group of male inguinal hernia; FIH: group of female inguinal hernia.

Table 2: Inflammation, congestion and, expressed HBME-1 and PCNA scores of the groups.

<table>
<thead>
<tr>
<th></th>
<th>MC score, Mean ± SD (n = 6)</th>
<th>FC score, Mean ± SD (n = 8)</th>
<th>MIH score, Mean ± SD (n = 25)</th>
<th>FIH score, Mean ± SD (n = 25)</th>
<th>p, MIH vs. MC</th>
<th>p, FIH vs. FC</th>
<th>p, MIH vs. FIH</th>
</tr>
</thead>
<tbody>
<tr>
<td>Inflammation</td>
<td>0.16 ± 0.01</td>
<td>0.25 ± 0.16</td>
<td>1.08 ± 0.90</td>
<td>1.16 ± 0.94</td>
<td>0.022</td>
<td>0.010</td>
<td>0.799</td>
</tr>
<tr>
<td>Congestion</td>
<td>0.50 ± 0.14</td>
<td>0.62 ± 0.51</td>
<td>1.78 ± 0.89</td>
<td>1.64 ± 0.86</td>
<td>0.001</td>
<td>0.004</td>
<td>0.734</td>
</tr>
<tr>
<td>HBME-1</td>
<td>1.00 ± 0.00</td>
<td>1.00 ± 0.00</td>
<td>1.96 ± 0.61</td>
<td>1.88 ± 0.60</td>
<td>0.001</td>
<td>0.000</td>
<td>0.640</td>
</tr>
<tr>
<td>PCNA</td>
<td>1.00 ± 0.00</td>
<td>1.00 ± 0.00</td>
<td>1.88 ± 0.66</td>
<td>1.84 ± 0.62</td>
<td>0.003</td>
<td>0.001</td>
<td>0.843</td>
</tr>
</tbody>
</table>

MC: male control group; FC: female control group; MIH: group of male inguinal hernia; FIH: group of female inguinal hernia.

Figure 2: Immunohistochemical findings (original magnification X 200). In the control groups (a,b); a) Immunohistochemical slight staining of HBME-1 in mesothelial cells; b) Immunohistochemical slight staining of PCNA in mesothelial and stromal nuclear; c) Diffuse staining with HBME-1 of mesothelium and the submesothelial stromal area in the study group; d) Diffuse staining of mesothelium and stromal cells with PCNA in the study group.
developing inside the gubernacular mesenchyme as the caudal genito-inguinal ligament. In the gubernac-
ular mesenchyme; the outer side includes the cremas-
ter muscle. In addition, while the invaginating PV takes
form in the middle layer, the innermost is formed the
cord that joins to the caudal epididymis and testis [13].
Amount of the persisting SMCs in the sac is vital for the
clinical outcome. While the SMC is less present in hy-
drocele, the amount of SMC is the least in the sac asso-
associated with undescended testis. Additionally, there
is more the smooth muscle bundle in the hernia sac
[5,9]. Apoptosis of SMC is necessary for PV oblitera-
tion to take place. There, however, is scarce information in
the literature about the fate of other PV cell structures
other than SMC [5,7,9,11]. Tanyel, et al. suggested that
the least SMCs were detected in the sac associated with
undescended testis and hydrocele while the most SMCs
were detected in the inguinal hernia sac. In parallel,
they stated that the programmed cell death (PCD) was
more observed in the sacs with the least SMC and PCD
was higher in the cells such as mesothelial other than
the SMC [14]. In our study, the proliferation of meso-
thenial, nerve, vascular and adipose tissues in the sacs
increased in MIH and FIH groups compared to control
groups (p < 0.05; Figure 1 and Table 1). There were
significantly SMCs in the study groups while the SMCs
were not encountered in the control groups. There was
no significant difference between MIH and FIH groups
regarding SMC proliferation (p > 0.05).

As in previous studies, significant SMC prolifera-
tion in our histopathological evaluation with quantita-
tive scores through H&E staining was noticeable in the
IH sacs. Unlike previous studies [5,9], there was also
a marked increase in proliferation in cells other than
SMCs. That is, PCD was probably defective in these cells
or apoptosis did not occur after a certain period in these
cells. In our study, increased expression of mesothelial
HBME-1 which is the marker for mesothelial cells [15]
was demonstrated to be a significant increase immu-
nohistochemically in the MIH and FIH groups compared
to the peritoneal and PV samples. HBME-1 was present
mostly in the cytoplasm and less in the cell membrane.
Also, the expression of PCNA significantly increased in
the study groups compared to the control groups. PCNA
is an essential protein found in proliferating cells, and
it performs crucial roles in DNA replication, repair, and
control of cell proliferation [16]. Immunohistoche-
val evaluation showed the increased nuclear staining
with PCNA antibody in the study groups compared to
those of control groups. This result together with the
increased HBME-1 staining in the cytoplasm and cell
membrane has seemed to demonstrate an increase in
the proliferation of, especially mesothelial cells.

Patients who had no apparent history of asbestos ex-
posure, but mesothelioma developing from the Nuck’s
channel, inguinal hernia, and hydrocele sacs have been re-
ported in the literature [3,4,17,18]. As in PV, a compen-
satory cell proliferation does not develop after develop-
mental apoptosis or PCD [19]. If apoptosis does not occur
in SMCs via PCD in the PV, an inguinal hernia, hydrocele,
or abnormal testicular localization take place [5]. That is,
there is no proliferation increase in mesothelial cells in
obliterated PV as it was in our study. On the other hand,
in the present work, we showed not only an increase in
mesothelial cell proliferation via the increased in HBME-
1 and PCNA expression but also the increased in SMC
proliferation qualitatively in the IH sac. On the contrary,
some researchers have argued that both the IH sac and
the obliterated PV share similar histopathology except
for the amount of SMCs when examined qualitatively
by optical microscopy using H&E stain or indirect immu-
nohistopathological method [5,19]. In a previous study,
apoptotic nuclei have been detected within the vascular
structures and mesothelium, but not within the SMCs
in the hernia sac. Namely, the attempt of apoptosis
during the process of PV obliteration has been shown in
vascular and mesothelial cells. Based on the results
we had, therefore, it may be speculated that when the
obliteration of PV through SMCs apoptosis or PCD did
not occur, the mesothelial cell together with vascular
and neuronal cells underwent proliferation because of
incomplete apoptosis in the same failure process. Even
though inguinal hernia surgery in children is delay-
d due to some social reasons, any practitioner does not
encounter a malignancy that develops from the hernia
Sac. In the literature, however, mesothelioma case aris-
ning from IH sac have been encountered in patients who
were at adult ages [1-4,17,18]. Therefore, the increased
mesothelial proliferation in the IH sac would seem to be
a predecessor of such malignancy for IH patients at very
low rates theoretically in any period of their life.

The mechanism by which occurs PV obliteration via
SMC apoptosis is identical to be responsible for the inhi-
bition of testicular descent. That is, the process of failed
testis descent remains the physiologic pathway for the
obliteration of PV. While undescended testis may indi-
cate the decrease of SMC requiring for propelling the
testis through the PV via a premature activation of this
process, prolonged exposure of the cremaster muscle
to the parasympathetic tonus may cause to ascend an
undescended testis. The reason for sustained contrac-
tility of the cremaster muscle is the increase in the level
of cytosolic calcium [14].

Apoptosis of SM in the PV occurs through mitochon-
drial Ca2+ overload and then depletion of Ca2+ stores
with an increase in cytosolic Ca2+. This cascade is initiated
by an increase in Bax and Fas proteins regulating at the lev-
el of the mitochondrial membrane. Thus, the SMCs and
mesothelial cells are exposed to PCD, respectively; and
obliteration of the PV takes place [8,14]. Apoptosis in a
cell occurs through two interconnected pathways. The
extrinsic pathway is started by ligation of death-inducing
receptors. The intrinsic pathway is initiated by oxidative
stress, DNA damage, hypoxia or growth factor depriva-
tion. This pathway is controlled by the Bcl-2 superfamily of proteins at the level of the mitochondrial membrane [20]. The depletion of $Ca^{2+}$ from internal stores through inositol 1,4,5-trisphosphate receptors (IP3) via activation of phospholipase C by G-protein-linked signaling transduction takes part in this initial steps in PCD [21]. Insulin-like growth factor-I (IGF-I) carries out many of its antiapoptotic actions through regulation of mitochondrial membrane permeability via Bcl-2 proteins, thus working against apoptosis. While IGF-I increases Bcl-2 interacting mediator of cell death via the intrinsic apoptotic pathway, it blocks the translocation to the mitochondrial of the prosapoptotic protein Bax via the same apoptotic pathway [22,23]. In a recent study, it was detected IGF Receptor-1 (IGFR-1) in the cremaster muscle (CM) associated with both the undescended and the descended testis. The density of IGFR-1 was higher in the CM, CM’s nerves and vascular tissues of the descended testis group than that of the undescended testis group [24]. The gubernacular-CM complex is innervated by the genitofemoral nerve (GFN). Alteration and divisions of the genitofemoral nerve inhibit the descent of the testis [25,26]. The Bcl-2 family as a crucial key of neuronal cell fate plays a role in the antiapoptotic action at the level of varying from purely cytosolic to the predominantly mitochondrial membrane in the neuronal cell. The decreased activities of Bcl-2 cause the decreased sympathetic tonus and lose of the neuron. Defective innervation of a muscle also gives rise to an increase in Bax and loss of motor neuron. As the IGFR-1 promotes the decrease of Bax and Fas and the increase Bcl-2 proteins; the receptor deficiency leads to the increase in Bax and Fas and the decrease Bcl-2 [27]. This result may be indicative of the decreased expression of IGFR-1 in CM associated with undescended testis [24]. On the other hand, for this apoptotic pathway which is effective in the apoptosis of PV, it may be proposed that the activity of IGFR-1 is required. If there is a defect or density difference at the IGFR-1 receptor, there may be alterations in the apoptosis of PV and is the need for further investigations.

Some studies have demonstrated that IGF1 is required for the development and functioning of the LCs in the developmental process of the testis. Based on this information it can be said that the Leydig cells in which insulin-like hormone 3 (INSL-3) requiring for the testis descent is produced need IGF1; otherwise, the production of INSL-3 becomes defective [28]. Therefore, the low density of IGFR-1 in the gubernacular-CM complex may probably be effective in the etiology of undescended testis [24]. According to mentioned above information, the PV obliteration requires a decrease in Bcl-2 and an increase in Bax for the SMCs apoptosis. IGFR-1 has a significant effect that triggers this event, and this act is accomplished through its receptor. Therefore, if PV and CM with the same mesenchymal origin are thought to share the same fate, the contribution of IGFR-1 to obliteration in the PV is likely.

In our study, there was no significant difference between FIH and MIH except for neuronal tissue proliferation histopathologically when both groups were compared. The neuronal staining score in males was higher than that of females ($p = 0.00$). This difference may be due to the sympathetic tonus being sexually dimorphic which is in favor of the male [10].

In a previous study, Nishioka, et al. showed that mesenchymal cells expressing vimentin appeared before mesothelial regeneration on the fibrotic peritoneum. Also, they found that HBME-1-positive cells were originated from vimentin-positive cells [15]. In another study, it has been suggested that immature SMCs known myofibroblast express vimentin, as an intermediate filament in the apoptosis of SMC which plays a role in the obliteration of the PV. Vimentin has been recognized, among other intermediate filaments, as an indicator of undifferentiated SMCs [9,11]. That is, if cells expressing vimentin represent both mesothelial and SMC precursor cells, vimentin-positive cells in the inguinal hernia sac may represent both. This result may be indicative of increased HBME-1 expression with a marker of mesothelial proliferation in the hernia sac as in our study. Also, increased PCNA expression in a hernia sac supports increased mesothelial cell proliferation.

Conclusions

The most popular etiological reason in the indirect IH is the defect in the PV obliteration. Defective apoptosis through the persistence of fetal SMCs in the proliferative changes occurring in the hernia sac gives rise to the defective obliteration of the PV. In our study, we determined that the SMCs increased in the hernia sac. This detection was supporting previous investigations and parallel with that of theirs. As it has been found in some recent works, PV and CM with the same mesenchymal origin are thought to share the same fate, the contribution of IGFR-1 to obliteration in the PV is likely. As a result of this defective apoptotic pathway, we also found an increase in proliferation of the mesothelial cells with the increased HBME-1 and PCNA expressions. There are mesothelioma cases developed in the hernia sac in advanced adult ages in the literature. Therefore, we suppose that the increase in mesothelial proliferation in the IH sac would seem to be a predecessor of such malignancy.

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Conflicts of Interest

None.

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References


