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ORIGINAL ARTICLE

Antimicrobial Resistance Patterns of *Pseudomonas aeruginosa* in a Vietnamese Tertiary Care Hospital

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Abstract

Background: Vietnam is one of the countries at high alert of antibiotic resistance. This study aimed to evaluate the antimicrobial susceptibility of *P. aeruginosa* isolates collected from a Vietnamese hospital in 2019.

Methods: A retrospective review was conducted of all reports of *P. aeruginosa* isolates from the records of the laboratory of Nhan Dan Gia Dinh Hospital between January 01 2019 to December 31 2019.

Results: Of 224 samples, the most common infection from which *P. aeruginosa* was isolated was pneumonia in hospitalized patients (49.1%) followed by skin and skinstructure infections (19,6%), and intra-abdominal infections (18.8%). The rates of isolates with MDR, XDR, and PDX were 7.1%, 20.5%, and 0.0%. Colistin was the most active agent overall (100.0%). Amikacin was the second most active agent, inhibiting 78.9% of all isolates. Other commonly used antipseudomonal β-lactams (cefepime, ceftazidime, imipenem, meropenem, and piperacillin-tazobactam) had susceptibilities for all isolates ranging from 68.0% to 70.9%. Ciprofloxacin and levofloxacin had overall 64.0%S and 61.8%S, respectively. Ticarcillin/Clavulanic acid was the least active antimicrobial tested with overall susceptibility of 38.1%.

Conclusions: The results of this investigation highlight the high rates of antibiotic resistance in *P. aeruginosa* in the hospital which will challenge optimizing empirical antimicrobial therapy for *P. aeruginosa* infections and require urgent antibiotic stewardship programs effectively.

Keywords

Antimicrobial, Multidrug resistance, *Pseudomonas aeruginosa*, Surveillance, Susceptibility

Abbreviations

ABR: Antibiotic Resistance; ASP: Antibiotic Stewardship Programs; VNMOH: Vietnam Ministry of Health; NDGDH: Nhan Dan Gia Dinh Hospital; CLSI: Clinical Laboratory Standards Institute; SSSI: Skin-Soft Tissue Infection; MDR: Multiple Drug Resistance; XDR: Extensively Drug-Resistant; PDR: Pan Drug-Resistant; NS: Non Sensible

Introduction

Antibiotic resistance (ABR) has been recognized as a global health issue. Pseudomonas aeruginosa is one of the most challenging organisms involved in a variety of infections. The World Health Organization (WHO) published a list of highly antibiotic resistant bacteria that are in need of priority for the research and development of new antibiotics. The list was divided into 3 levels, in which the most critical is the carbapenem-resistant *Pseudomonas aeruginosa* [1].

A geographic and temporal trend in resistant phenotypes of *P. aeruginosa* from \geq 400 medical centers worldwide over the 20 years of the SENTRY Antimicrobial Surveillance Program during the period from 1997 to 2016 found that the most common



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infection from which *P. aeruginosa* was isolated was pneumonia in hospitalized patients (44.6%) followed by bloodstream infection (27.9%), with a high rate of multiple drug resistance (MDR) (27.7% vs. 23.7, respectively) [2]. Regional variations in antibiotic resistance patterns for different organisms including *P. aeruginosa* also occur [2], which could be due to differences in antibiotic prescribing practices. Better understanding of global trends in antibiotic resistance for the organism is obtained through local and regional surveillance studies [3].

In Vietnam, *P. aeruginosa* has appeared as the most commonly isolated organisms in hospitals, causing 20-30% of all nosocomial infections [4]. In 2016, the Vietnam Ministry of Health (VNMOH) approved a national program in the effort of preventing the spread of ABR and a guideline on the implementation of "Antibiotic stewardship programs (ASP) in hospitals" [5]. ASP is a multifaceted, multidisciplinary team approach to combat antibiotic misuse. Of which periodic testing and evaluation of antibiotic resistance of bacterial agents is useful for physicians to detect resistance pattern and assist in the selection of an appropriate antibiotic for empiric treatment in a particular setting.

Therefore, this study aimed to determine the status of antimicrobial resistance to anti-pseudomonal agents and the magnitude of the multi-drug resistance to *P*. *aeruginosa* in one hospital.

Material and Methods

Study design and setting

A retrospective review was conducted of all reports of *P. aeruginosa* isolates from the records of the laboratory of Nhan Dan Gia Dinh Hospital (NDGDH) between January 01, 2019 to December 31, 2019. NDGDH is a 1500-bed tertiary care referral hospital in Ho Chi Minh City, Vietnam. In cases where more than one *P. aeruginosa* was isolated from a single patient, the sample that is considered to be the primary source of infection is included (i.e. pulmonary excretions rather than blood in cases of pneumonia).

Isolation and identification

Submitted samples were analyzed using the traditional culture methods. Urine samples were inoculated onto blood agar and MacConkey agar. Sputum and tracheal aspirates were inoculated onto blood agar, MacConkey agar and chocolate agar. Blood samples were inoculated onto BACTEC Aerobic Plus vial, while cerebrospinal fluid and other body fluids were inoculated onto blood agar, MacConkey agar, trypticase soy broth and chocolate agar.

Identification of *P. aeruginosa* was based on routine biochemical methods that included the following reactions: Gram-negative bacilli, oxidase positive, unable to ferment sugars on triple sugar iron, motile, do not produce sulfide and indole, citrate positive, urease negative, lysine decarboxylase positive, lysine deaminase negative, able to grow at 42 °C, and pigmented.

Antibiotic susceptibility test

The investigated antibiotics were ticarcillin, ticarcillin/acid clavulanic, piperacillin, piperacillin/tazobactam, ceftazidime, cefepime, imipenem, meropenem, gentamicin, tobramycin, amikacin, ciprofloxacin, levofloxacin, and colistin. The *in vitro* susceptibility of these antibiotics was tested using the Kirby-Bauer disk diffusion method under the M02-A11 guidelines of the Clinical Laboratory Standards Institute (CLSI) and were confirmed by the automated VITEK 2 system (bioMérieux Inc., France). The subsequent results of the tests were then interpreted in accordance with the M100-S25 document.

Resistant phenotypes

Resistant phenotypes analysed using EUCAST criteria were as follows: MDR (NS to at least 1 antimicrobial in \geq 3 drug classes), extensively drug-resistant ([XDR] NS to at least 1 agent in all but \leq 2 drug classes), and pan drug-resistant (PDR), according to Magiorakos, et al. [6]. Ceftazidime-non sensible (NS) and meropenem-NS were determined according to EUCAST interpretive criteria.

Quality control

P. aeruginosa, with American type culture collection (ATCC) number 27853, was used as the reference strain for quality control of culture media, biochemical tests and susceptibility testing.

Ethical consideration

The study was approved by the Nhan Dan Gia Dinh Hospital's ethical review board with approval number 15/HĐĐĐ, on 06 January 2020. The study was conducted in a spirit of respecting the private information related to patients and health care providers. Information which was collected from routine data of drug charts was anonymized.

Statistical analysis

All data was analysed by using SPSS software version 20.0. The proportion of susceptible isolates was calculated as the sum of susceptible organisms (neither intermediately susceptible nor resistant) relative to the total number of organisms tested. The drug resistance pattern of *P. aeruginosa* with site of infection, specific antibiotics, and resistance phenotypes was summarized in terms of frequencies and percentages.

Results

Sample and infection type

Of 224 samples, the most common infection from which *P. aeruginosa* was isolated was pneumonia in

hospitalized patients (49.1%) followed by skin-soft tissue infection (SSSI) (19.6%), and intra-abdominal infection (18.8%) as shown in Table 1.

The rates of isolates with MDR and XDR were 7.1% and 20.5%. Pneumonia had a higher rate of isolates with MDR and XDR (11.8% and 21.8%, respectively) than BSIs (0.0% and 20.0%, respectively) as shown in Table 2. There was no PDR isolate. The frequency of XDR, ceftazidime-NS, and meropenem-NS isolates were highest in the urinary tract infection (61.1%, 76.0%, and 72.0%).

Activities of specific antimicrobials are shown in Figure 1 for all isolates and in Figure 2 for infection type. Colistin was the most active agent overall (100.0%). Amikacin was the second most active agent, inhibiting 78.9% of all isolates. Tobramycin and gentamicin were slightly less active than amikacin and inhibited 74.1% and 71.9% of all isolates, respectively. Other

Table 1: Distribution of *Pseudomonas aeruginosa* isolated by infection and sample type antimicrobial susceptibility.

| Infection type/Sample | Frequency | Ratio |
|-------------------------------------|-----------|-------|
| Pneumonia | 110 | 49.1 |
| Sputum | 103 | 46.0 |
| Blood | 6 | 2.7 |
| Bronchoalveolar lavage | 2 | 0.9 |
| Pleural fluid | 1 | 0.4 |
| Skin - soft tissue infection (SSTI) | 44 | 19.6 |
| Pus | 26 | 11.6 |
| Skin/soft tissue drainage | 18 | 8.0 |
| Intra-abdominal infection | 42 | 18.8 |
| Abdominal fluid and pus | 42 | 18.8 |
| Urinary tract infection | 18 | 8.0 |
| Urine | 18 | 8.0 |
| Bloodstream infection (BSI) | 10 | 4.4 |
| Blood | 9 | 4.0 |
| Vessel catheter | 1 | 0.4 |
| Total | 224 | 100.0 |

commonly used antipseudomonal β -lactams (cefepime, ceftazidime, imipenem, meropenem, and piperacillintazobactam) had susceptibilities for all isolates ranging from 68.0% to 70.9%. Ciprofloxacin and levofloxacin had overall 64.0%S and 61.8%S, respectively. Ticarcillin/ Acid clavulanic was the least active antimicrobial tested with overall susceptibility of 38.1%.

The resistance patterns of *P. aeruginosa* to the antibiotics are almost identical among infection types (Figure 2). Colistin and aminoglycosides are still the most effective as compared to other antibiotics, and ticarcillin/acid clavulanic was still noted to have the highest resistance. However, resistance of *P. aeruginosa* is highest in urinary tract infection, followed by BSIs.

Discussion

Antimicrobial susceptibility

Resistance profile of *P. aeruginosa* in some studies in the literature is summarized in Table 3. Over the 20 years of SENTRY Program surveillance [2], the rates of resistant phenotypes for *P. aeruginosa* were highest in 2005-2008 and decreased in 2009-2016. The Asia-Pacific region had an overall lower frequency of MDR *P. aeruginosa* than Latin America and Europe. The region saw an increase in MDR *P. aeruginosa*, from 15.6% in 1997-2000 to 24.7% in 2005-2008 and decreased to 15.0% in 2013-2016. MDR *P. aeruginosa* in our study was lower (7.1%). XDR *P. aeruginosa* in 2013-2016 of SENTRY Program surveillance was 15.2% which was lower than one in our study (20.5%).

The frequency of Ceftazidime-NS, and meropenem-NS isolates were one third and were highest in the urinary tract infection. A study by Biedenbach, et al. [7] found that frequency of ceftazidime-NS, and meropenem-NS *P. aeruginosa* from 5 medical centers in Vietnam provided 529 *P. aeruginosa* isolates from patients with hospital-acquired or ventilator-associated pneumonia from 2012 to 2014 was 57,7% and 43.5%, respectively.

The resistance patterns of *P. aeruginosa* to the antibiotics are almost identical in many studies. Colistin and aminoglycosides are still the most

Table 2: Pseudomonas aeruginosa isolates 2019 stratified by infection type and percentage of isolates with resistant phenotypes.

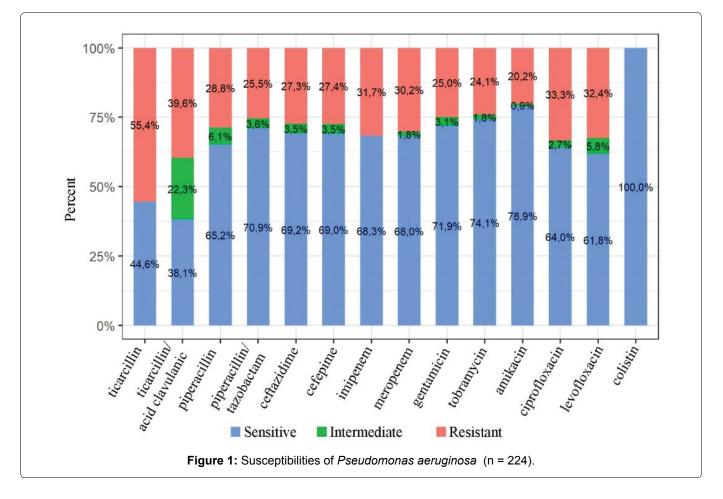
| Infection type/Resistant Phenotype | MDR | XDR | PDR | Ceftazidime -NS | Meropenem -NS |
|---------------------------------------|-------|-------|------|--------------------|------------------|
| Pneumonia (n = 110) | 11.8% | 21.8% | 0.0% | 29.6% | 35.5% |
| Skin-soft tissue (n = 44) | 2.3% | 13.6% | 0.0% | 20.8% | 16.0% |
| Intra-abdomen (n = 42) | 2.4% | 7.1% | 0.0% | 18.8% | 16.1% |
| Urinary tract (n = 18) | 5.6% | 61.1% | 0.0% | 76.0% | 72.0% |
| Bloodstream (n = 10) | 0.0% | 20.0% | 0.0% | 22.2% | 22.2% |
| Total (n = 224) | 7.1% | 20.5% | 0.0% | 30.8% | 32.0% |

MDR: Multidrug Resistant; EDR: Extensively Drug Resistant; PDR: Pan Drug Resistant; NS: Non-Sensible. Criteria as published by European Committee on Antimicrobial Susceptibility Testing (EUCAST) 2018.

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|-------------------------------|-----------------------|---------|---|--|------|--------|-----|----------------|----------------|-------------------|----------------|----------------------|----------------------|--------------------------|
| Authors | Period | Samples | Population | Settings | MDR | XDR F | PDR | Colistin NS | Amikacin NS | Ceftazidime NS | Cefepime NS | Mero- penem NS | Piper/ Tazo NS | Cipro- floxacin NS |
| Shortridge D, et al. [2] | 1997 <i>-</i> 2016 | 52022 | All | fic, | 24.9 | 17.6 0 | 0.1 | 0.6% | 7.0% | 22.5 | 20.7 | 23.9 | 26.9 | 27.1 |
| | 2013- 2016 | 16461 | | European, Latin American, and North American regions | 21.8 | 15.2 0 | 0.1 | | | 19.2 | | 22.6 | | |
| Biedenbach- DJ, et al. [7] | 2012- 2014 | 529 | hospital-ac- quired or ventila- tor-associ- ated pneu- monia | 5 medical centers in Vietnam | 1 | 1 | · | | 18.4 | 57.7 | 39.9 | 43.5 | 43.9 | 45.0 |
| Phu VD, et al. [8] | 2012- 2013 | 100 | Hospital- acquired infections | 15 ICU in 14 hospital in Vietnam | ı | 1 | 1 | | 1 | 1 | 1 | 55.7%* | 1 | 1 |
| Pfaller MA, et al. [9] | 2013- 2015 | 489 | All hospi- talized pa- tients | 14 medical centers located in 7 countries in the APAC region (minus China, Australia and New Zealand) | 27.4 | 1 1 | | 1.6% | 7.8% | 25.8% | 20.9% | 28.3% | 27.9% | 26.0%* |
| | | 108 | | Korean | | | | | | 38.9% | | 46.3% | 42.6% | |
| | | 37 | | India | | | | | | 32.4% | | 33.3% | 40.5% | |
| | | 138 | | Thailand | | | | | | 27.5% | | 33.3% | 29.0% | |
| | | 63 | | Singapore | | | | | | 20.6 | | 17.5 | 19.0% | |
| | | 21 | | Taiwan | | | | | | 19.0% | | 19.0 | 25.0% | |
| | | 06 | | Malaysia | | | | | | 14.4 | | 11.1 | 14.4 | |
| | | 32 | | Hong Kong | | | | | | 12.5 | | 15.6 | 15.6 | |
| Sader HS, et al. [10] | 2012- 2015 | 7452 | AII | 79 U.S. medical centers | 15.5 | 9.4 | 0 | 0.7 | 3.0 | 15.7 | 14.6 | 18.0 | 19.4 | 22.5 |
| Our study | 2019 | 244 | AII | A hospital in Vietnam | 7.1 | 20.5 0 | 0.0 | 0.0 | 21.1 | 30.8 | 30.9 | 32.0 | 29.1 | 36.0 |
| - | - | 5 | | | | | | | | | | | | |

Table 3: Non-susceptibilities of Pseudomonas aeruginosa of different studies.

*Carbapenem resistance; #Levofloxaci



effective as compared to other antibiotics, followed by antipseudomonal β -lactams (cefepime, ceftazidime, imipenem, meropenem, and piperacillin-tazobactam), then quinolones. However, the rates of antibiotic -NS *P. aeruginosa* in our study are higher in ones in SENTRY Program surveillance.

Pneumonia had a higher rate of isolates with MDR and XDR than BSIs while the frequency of XDR, ceftazidime-NS, and meropenem-NS isolates were highest in the urinary tract infection. A study by Biedenbach, et al. [7] found that the rates of isolates NS antipseudomonal β -lactams and quinolones of Vietnamese patients with hospital-acquired or ventilator-associated pneumonia was very high and in range of 40.0%-60.0%. Similarly, carbapenem resistance was most common in *Pseudomonas aeruginosa* (55.7%) in Vietnamese patients in Intensive Care Units with hospital-acquired infections [8].

Compared to the results of a study by Pfaller, et al. in the period 2013-2015 [9] and Sader, et al. in the period 2012-2015 [10], our frequency of antibiotic-resistant *P. aeruginosa* were lower than Korea and India; and similar to Thailand; and higher than Singapore, Taiwan, Malaysia, Hong Kong, and United States.

Limitations

Studies in the Asia-Pacific region have shown an increasing prevalence of metallo- β -lactamases and carbapenemases in *P. aeruginosa*, particularly the

ST235 clone, which may explain the increase in MDR [11]. However, strain typing was not performed in this study, it is unknown whether the decrease in resistance is due to the prevalence of ST235 or other causes.

Conclusion

The results of this investigation highlight the high rates of antibiotic resistance in *P. aeruginosa* in our hospital which will challenge optimizing empirical antimicrobial therapy for *P. aeruginosa* infections and require urgent ASP effectively.

Acknowledgements

No.

Sources of Support

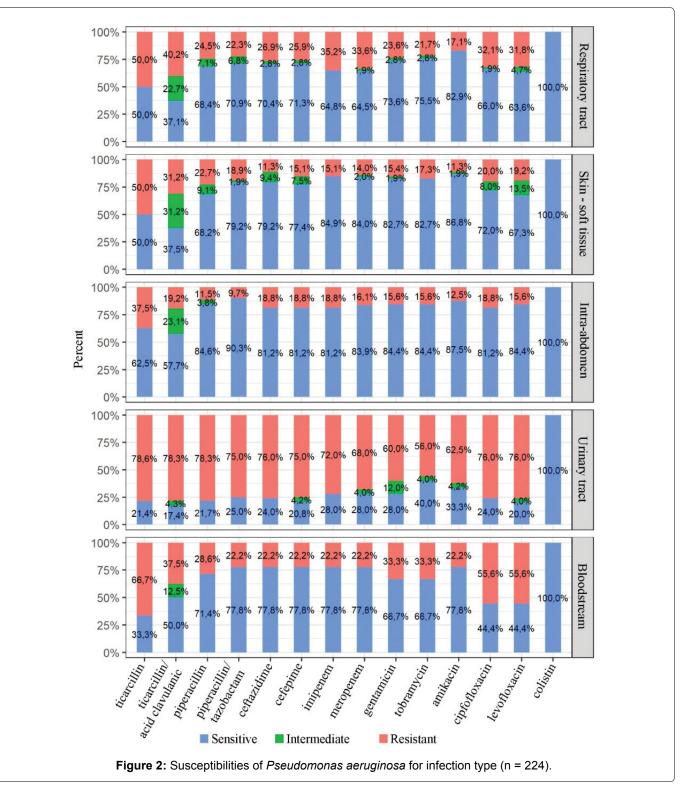
There is no funding available for this study.

Conflicts of Interest

All authors don't have any conflicts of interest to declare.

Statement of Equal Authors' Contribution

Pham HT: Conceptualization, Methodology, Writing- Reviewing and Editing, Supervision; Bui QH: Data curation, Formal analysis Writing- Original draft preparation; Nguyen HH: Conceptualization, Writing-Reviewing and Editing; VoTH: Conceptualization, Methodology, Writing- Reviewing and Editing, Supervision.



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