



RESEARCH ARTICLE

Plastic Additive Bisphenol A: Toxicity in Surface- and Groundwater Crustaceans

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Abstract

Bisphenol A (BPA) is being considered by the European Union as a substance of very high concern, occurring worldwide due to its wide application in many plastic products, building materials, coatings and epoxy resins. The toxicity of BPA in groundwater invertebrates is insufficiently understood.

Both acute (24 h) and chronic (28 d) toxicity was compared in *Daphnia magna* and ecologically important crustaceans in surface water (*Gammarus fossarum*, *Eucyclops serrulatus*) and groundwater (*Niphargopsis casparyi*, *Proasellus slavus*). Survival and spontaneous locomotory activity was recorded in real-time in the Multispecies Freshwater Biomonitor (MFB) and the new Microimpedance Sensor System (MSS) for Meiofauna.

The benthic copepod *E. serrulatus* was the most sensitive test species, followed by the groundwater species *N. casparyi* and *P. slavus* with *G. fossarum* and *D. magna* being the least sensitive species in acute tests. In chronic exposures the most sensitive species was *P. slavus*, whereas the amphipods and *D. magna* showed overall similar sensitivity. *G. fossarum* showed slightly more sensitive behavioral responses compared to *N. casparyi*, while *D. magna* behavior was affected by an age-dependent increase, masking potential negative effects of BPA on locomotory activity. Feeding activity of *G. fossarum* was more sensitive than molting and reproduction of *D. magna*. *G. fossarum* can be recommended for toxicity evaluation of groundwater habitats, as no standard test procedures and toxicity data for groundwater species are available. The organisms might also be used as biomonitors for landfill leachates, wastewater treatment plants, sewage plants and paper recycling plants.

Keywords

Bisphenol A, *Gammarus fossarum*, *Niphargopsis casparyi*, *Proasellus slavus*, *Eucyclops serrulatus*, *Daphnia magna*

Introduction

In 2016, REACH classified BPA as substance of very high concern, due to its toxic effects on reproduction (svhc: substances of very high concern, category 1B), which is being implemented from 1 March 2018 in Europe [1]. Moreover, BPA has been placed on the watch list of the European Water Framework Directive as xeno-estrogen micropollutant (www.umweltbundesamt.de).

The synthetic phenol 2,2 Bis (4-hydroxyphenyl)-propane (Bisphenol-A, BPA) is being used as additive in the production of plastic materials (mostly polycarbonate), phenol and epoxy resins, polyesters and polyacrylates [2]. It can be found in many products of daily life, such as CDs, polycarbonate bottles, thermal receipts, coatings of food tins and drinking water pipes, resin based dental sealing and epoxy glues [3]. BPA is being produced in huge amounts worldwide (e.g. 410.000 t/y in Germany) and occurs ubiquitous, even in dust and human urine samples [4].

BPA is released into the environment through sewage treatment effluents, landfill leachates and natural degradation of polycarbonate plastics [5]. The negative effects of BPA in vertebrates are various and include disruptions to the reproductive, immune and central nervous systems [6] as well as inducing developmental disorders in the brain and carcinogenesis, whereby numerous signaling pathways can be affected [2]. Moreover, endocrine disruption [7] and impairment of the glucose metabolism have been reported in zebra-fish *Danio rerio* larvae [8] as well as reduced fish body length, induced oxidative stress and altered gene ex-

pression of immune-related genes in zebrafish [9]. The replacement products such as Bisphenol F and Bisphenol S [10] and degradation products such as BPA-mono/dimethyl ethers are toxic, too [11].

Wastewater treatment plants have a varying capacity to reduce BPA by 61-98%, however, the concentrations of degradation/transformation products may rise [3]. In surface waters, BPA concentrations range between 0.01 - 2.4 µg/L, most of the small German streams showed values below 0.05 µg/L, whereas large rivers like the Danube and Main can reach up to 130 µg/L resp. 50 µg/L (www.lfu.bayern.de) [12]. BPA levels in landfill leachates, wastewater treatment plants, sewage plants and paper recycling plants can reach up to 33.5 mg/L [13,14]. Drinking water in China contains up to 6.5 ng/L BPA [15].

BPA can accumulate in sediments to 6 - 30 µg/Kg [16]. BPA uptake in human beings results mostly from food/beverage and thermal receipts, but also from dust. BPA has been rated as an endocrine disruptor, affecting both sexual and thyroidal hormone pathways.

Since 2011 BPA has been forbidden as a precautionary measure in feeding bottles for infants in the EU [1]. However, Japan has banned BPA already 20 years ago in all materials related to human consumption. Canada banned BPA in 2008 [1] after the U.S. National Toxicology Program (NTP) expressed the first concerns [17]. The European Food Safety Authority (EFSA) reduced the tolerable daily intake (TDI) of BPA in 2015 down to 4 µg/Kg. In France, the use of BPA has been banned for all food/beverage packaging in 2015.

Consulting the ecotoxicology database of US-EPA [18], numerous toxicity studies of several BPA-compounds (esp. CAS 79447 and 80057) have been performed on the zebrafish *Danio rerio* and *Daphnia magna* as standard species in ecotoxicology. A rough overview of the US ecotoxicology database shows that BPA does not seem to be highly acutely toxic on survival, growth and behavior of aquatic crustaceans with gammarids showing similar sensitivity as daphnids. However, no studies on groundwater crustaceans have been reported so far.

Groundwater habitats are characterized by lack of light, space and current, constant temperature and usually low pollutant levels. Groundwater invertebrates are mostly small, they show slow growth and long lifespan, thin cuticula, loss of eyes and pigments, slow metabolism and ability to starve long periods of food shortage [19-21]. Therefore, they are expected to react differently to xenobiotics compared to their epigeal relatives. Currently there is a scientific debate if the standard aquatic test species, primarily daphnids, also protect groundwater invertebrate species or if there is a need to find groundwater test species and develop specific toxicity test guidelines for a safe risk assessment

to fulfill the European Groundwater Directive.

The aim of this study is to provide toxicity data on BPA on groundwater crustaceans in comparison to surface water species and standard test species in aquatic ecotoxicology.

Materials & Methods

Test species

Both surface water and groundwater crustaceans were used and compared for the evaluation of BPA-toxicity in aquatic environments.

Gammarus fossarum (Crustacea, Amphipoda) represents a key ecological stream invertebrate in the whole Northern hemisphere due to its; (1) Wide geographical distribution; (2) High abundances; (3) Central position in aquatic food web and importance as decomposer; (4) Important role as bio indicator for saprobic water quality class II according to the European Water Framework Directive and; (5) Increased use as test species in ecotoxicology [19]. Organisms for tests and laboratory culture were originally collected in a small mountainous creek in Kreuzlingen, Switzerland (47.63311 °N, 9.16553 °E) and bred since then in the laboratory.

Daphnia magna (Crustacea, Phyllozoa) represents a key ecological species in lake ecosystems and has been used as standard test species in ecotoxicology for more than four decades. The initial organisms for tests and laboratory culture have been received from the University of Constance. The animals were cultured and bred at 20 °C, 16:8 h photoperiod in medium according to ISO 6341 prepared with water from Lake Constance with *Scenedesmus longispina* as food (3x/week).

Eucyclops serrulatus (Crustacea, Copepoda) is primarily regarded as epigeal benthic freshwater inhabitant, which also lives in the interstitial and in genuine and organically enriched groundwater habitats [22,23]. Organisms for tests were taken from our laboratory culture.

Niphargopsis casparyi (Crustacea, Amphipoda) are hypogean relatives of the epigeal gammaridae. Niphargids have not been used in ecotoxicological testing as laboratory culturing has so far been unsuccessful.

Proasellus slavus (Crustacea, Isopoda) represents a small un-pigmented asellid with a life expectancy of up to 15 years [24]. As hypogean relative to surface water *Asellus* spp. it co-occurred with *N. casparyi* in the groundwater monitoring site in about 50 m depth in the Rhine valley near Neuenburg, Southwest Germany (47.81272 °N, 7.54740 °E).

Experimental setup

Cultures: The culture of *D. magna* was performed according to ISO 6341. 800 mL medium dissolved in pre-filtered (100 µm) water from Lake Constance were filled in 1 L glass beakers with 5 animals at 20 °C with a 16 h:8 h

photoperiod and 5 mL algae suspension of *Scenedesmus longispina* (Helbig Lebendkulturen) culture 3x/week, when the test medium was renewed. The culture was checked weekly for quality by determination of the sex of the animals under the microscope (Motic B3 Professional Series, 4 × 10 × 40).

The culture of *G. fossarum* was performed in a 20 L glass aquarium in a thermostat at both 10 °C (for comparison with the tests of the groundwater species) and at 20 °C (for comparison with *D. magna*), containing water from Lake Constance, pre-conditioned alder leaves and pebbles as substrate. The culture was kept and bred in the dark under constant aeration (oxygen saturation 95%). The water was renewed on a weekly basis and animals then fed with chironomid larvae (Poseidon-Aquaculture). The stygal crustaceans were collected in the field and maintained in Polyethylene boxes (40 × 30 × 20 cm) covered with parafilm at 10 °C in the dark with a few drops of fine detritus and in water from the groundwater sampling site for at least 4 weeks. Evaporating water was replaced on a weekly basis by water from Lake Constance.

Toxicity tests: Acute (24 h, 96 h) and chronic (16 - 28 d) toxicity tests were performed with the abovementioned crustaceans in a thermostat (at 10° and additionally 18° (*N. casparyi*, *P. slavus*, *G. fossarum*) without illumination, and for *D. magna*: 20 °C with 16:8 h photoperiod). For each species five organisms were placed in a beaker (250 mL) filled with 200 mL test water (Lake Constance water, drinking quality: pH 7.0, NH₄: 0.05 mg/L, hardness 1.24 mmol, NO₂: < 0.02 mg/L, NO₃: 0 mg/L, PO₄: 0.25 mg/L) and different BPA concentrations were added. The tests were replicated 3 - 4 times, the animals not fed during the acute tests. All experiments were accompanied by controls (Lake Constance water) and controls with ethanol, functioning as solvent (acute tests: 0.5 ml/L, chronic tests: 0.05 ml/L). The amount leaf eaten by the organisms (amphipods, isopods) was calculated as % leaf loss related to the number of the survivors in each beaker each week.

As the feeding habits of the stygal species are still unknown additionally 1 drop of fine detritus from the groundwater sampling site was added to each beaker. As groundwater organisms live in interstitial habitats 5 g fine pebbles (mixed sizes 1 - 3 mm, after previous incineration for 1 h, 500 °C) were added as substrate

and hiding places. *G. fossarum* demands higher oxygen saturation than groundwater species, therefore gammarid beakers were continuously aerated using standard aquarium pumps (ACO, Green Sun) with fine pipette tips gently dipped into the beakers to achieve 90 - 100% oxygen saturation.

During chronic toxicity tests with *D. magna* molting and reproduction was monitored daily according to the OECD guideline no. 211.

As BPA is poorly soluble in water, a stock solution was prepared with 50 mg/L of granulated BPA, dissolved in 1 mL Ethanol (96%) and 999 mL lake water. Then, the solution was placed inside a water bath (40 °C) and stirred for one hour until the BPA was completely dissolved and the ethanol evaporated. For *D. magna* tests Daphnia medium was used instead of lake water. BPA solutions were prepared and changed every week to ensure a stable concentration during the experiments.

Each concentration level of BPA was tested in 3 - 4 replicates, depending on the amount of available test organisms: the field collected groundwater species were the bottleneck in the study. Beakers were covered with parafilm to minimize evaporation and hence change BPA concentration. Survival and behavior were monitored twice a week. Behavior was quantitatively recorded in lake water in eight randomly chosen organisms from each BPA concentration level, at selected exposure time intervals (acute tests: recorded continuously for 24 h in the test solutions; chronic tests: recorded for 2 h twice a week in lake water/medium) in the MFB (Multispecies Freshwater Biomonitor[®]) [25,26] or in the MSS (Micro-impedance Sensor System[®] for *E. serrulatus*). The MSS is a new recording unit for small organisms of sizes around 1 - 1.5 mm [27]. In the chronic exposures behavior was recorded twice a week for a period of 2 h in lake water, in order to avoid BPA-spilling and ease of handling. Table 1 gives an overview over the toxicity tests.

According to the OECD test guidelines, additionally reproduction (number of neonates per female) and molting was recorded daily in the chronic tests with *D. magna*.

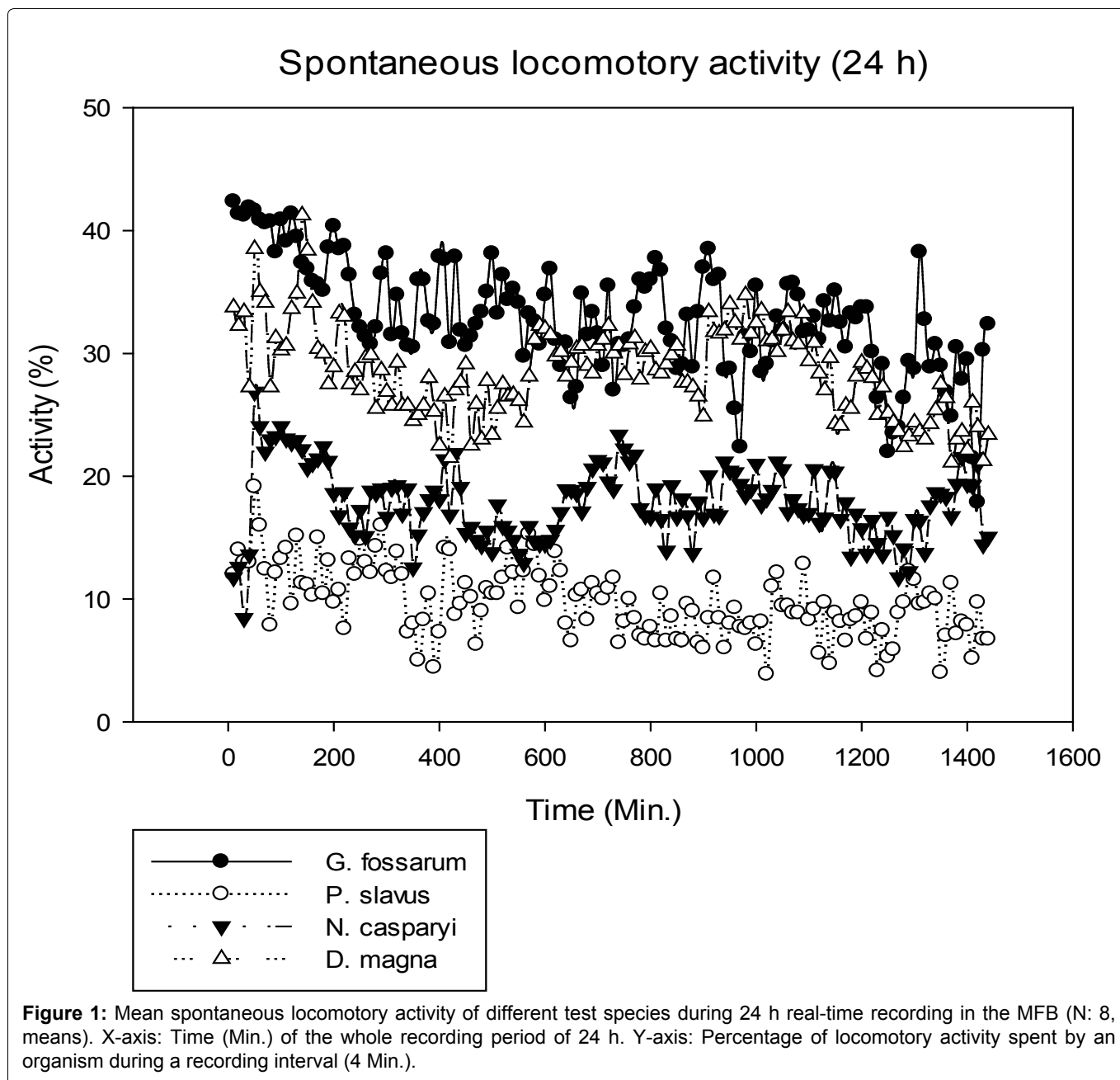
Analytics

BPA (CAS 1980-05-07, 97%, Alfa Aesar) levels were exemplarily verified in a wide range of concentration levels in; (1) The weekly prepared stock solutions and;

Table 1: Overview of the toxicity tests with Bisphenol A (BPA).

Test species	Acute BPA (µg/L)	Chronic BPA (µg/L)
<i>E. serrulatus</i> (0.8-1 mm)	96 h: 10, 100, 500, 2.500, 5.000	16 d: 10, 50, 100
<i>G. fossarum</i> (5-7 mm)	24 h: 200, 500, 1.000, 1.500, 3.000, 5.000, 6.200, 10.000, 12.500, 25.000, 50.000	28 d: 30, 50, 100, 300, 500, 1.000, 2.000, 3.000
<i>N. casparyi</i> (5-8 mm)	24 h: 1.000, 5.000, 10.000	28 d: 100, 500, 1.000
<i>P. slavus</i> (4-8 mm)	24 h: 200, 500, 5.000, 25.000, 50.000	16 d: 10, 50, 100, 500
<i>D. magna</i> (1-2 mm)	24 h: 1.500, 3.000, 5.000, 6.200, 8.500, 10.000, 12.500, 25.000	28 d: 30, 300, 3.000

Size of test organisms (mm), test duration (d: days) and used nominal concentrations of BPA (µg/L).



(2) Directly after dosing to the experimental beakers and; (3) One week after exposure, i.e. before the weekly change of test solutions in the beakers to prove that (1) Solubility in Ethanol was sufficient to solve the BPA; (2) The experimental dilutions were correctly prepared and; (3) The loss of BPA during one week was negotiable. Stock solutions (50 mg/L) showed high recovery after 1 week (49 mg/L, \pm 1 mg/L). The loss of BPA in the experimental beakers after 1 week reached between 5 and 20% esp. at higher levels concentration levels.

Statistics

Survival data from the acute exposures were estimated by a linear regression of $\log(x)$ and probit (y) transformed data in SigmaPlot. The EC_{50} for locomotory activity were estimated when possible with SigmaPlot with a 4-parameter logistic curve.

Behavior data were analyzed by non-parametric repeated measures ANOVA on ranks (Friedmann test)

for time-dependent data or Kruskal-Wallis ANOVA to compare different groups for not time-dependent data (e.g. reproduction, molting of *D. magna*), followed by post-hoc pair wise comparisons (Tukey, Dunn's test).

Results

Spontaneous locomotory activity

G. fossarum is a highly mobile species and thus shows the highest activity, followed by the planktonic *D. magna*. The groundwater species, *N. casparyi* and esp. *P. slavus* showed lower locomotory activity as they live in the interstitial with limited space (Figure 1). The same holds for *E. serrulatus*, living in fine particulate sediment zones [27]. *D. magna* showed the highest variation in activity during the experiments. Generally, normal activity levels varied up to 15% over time in the controls. A significant BPA-dependent decrease in activity was therefore pre-defined as > 20% permanent difference in activity levels.

Spontaneous locomotory activity of stygal crustaceans at different temperatures.

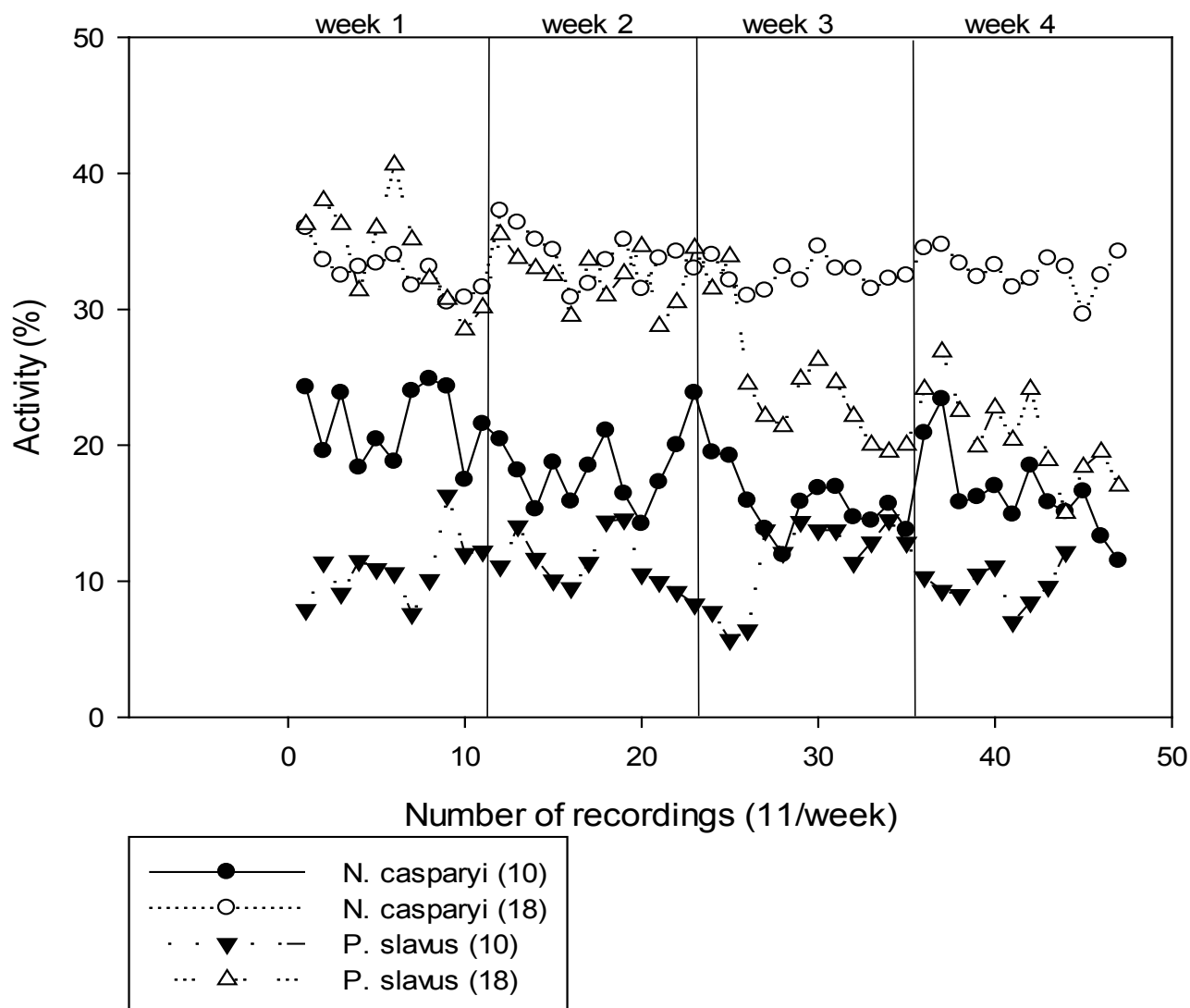


Figure 2: Temperature-dependent (10 versus 18 °C) normal baseline activity of the stygal crustaceans *N. casparyi* and *P. slavus* (Means of 10-18 animals). Y-axis: Percentage of locomotory activity spent by organisms during a recording interval (4 Min.). X-axis: Number of subsequent recordings (11) once a week, over a period of 4 weeks.

During the 28 d exposures *D. magna* grew fast to sexual maturity and their spontaneous locomotory activity significantly ($p < 0.001$, Kruskal Wallis test) increased with age (6 d: mean = 33.33 (SD = 6.33); 12 d: mean = 43.90 (SD = 2.5); 46 d: mean = 48.57 (SD = 3.64)). This was not the case in the slow growing surface and groundwater amphipods and isopod, which have much longer life cycles. Statistically, there is a significant difference ($p < 0.05$) among the organisms of different age: younger daphnids showed lower locomotory activity with a very high variability, whereas older organisms' activity increased and was more stable.

Stygal species are adapted to low and constant temperatures in groundwater habitats. In order to test whether the organisms also tolerated higher temperatures, which would allow to perform all tests

on all species at the same temperature levels survival and activity of stygal species was tested at both 10 °C and 18 °C. The activity of the stygal animals generally increased at higher temperature, however for *P. slavus*, it fell back to low levels after 2 weeks, followed by increased death (Figure 2). Therefore, it was decided to perform the toxicity tests with the stygal species and gammarids at 10 °C, to avoid artifacts of unrealistic conditions and for comparisons, whereas the tests with daphnids were performed at 20 °C according to standard test procedure, and compared with tests with *G. fossarum* at 18 °C (which is the temperature of the long-term culture in the laboratory).

Acute toxicity

After 24 h, the small-sized copepod *E. serrulatus* was

Table 2: LC₅₀ values (R²), EC₅₀ (± SE) and ET₂₀ values generated from the acute toxicity tests.

Species BPA (mg/L)	<i>E. serrulatus</i>	<i>G. fossarum</i>	<i>N. casparyi</i>	<i>P. slavus</i>	<i>D. magna</i>
LC ₅₀ -24 h	0.80 (0.75)	10.60 (0.64)	12.30 (0.68)	6.30 (0.91)	11.20 (0.56)
EC ₅₀ -24 h	-	13.50 (1.50)	5.00 (1.90)	25.00 (2.50)	10.30 (6.10)
ET ₂₀ at x BPA	-	16 h: 5 mg/L	24 h: 5 mg/L	6 h: 2.5 mg/L	24 h: 6 mg/L

LC₅₀: Median lethal concentration; EC₅₀: Median effect concentration; ET₂₀: Time to 20% effect; x: BPA-concentration.

Table 3: Comparison of chronic effects (28 d) of BPA on different crustaceans.

Species	LC ₅₀ (mg/L)	EC ₅₀ (mg/L)	ET ₂₀ (mg/L)
<i>G. fossarum</i> (28 d)	LC ₂₀ : 0.80	EC ₅₀ : 1.00	16 d: 0.10
<i>D. magna</i> (28 d)	LC ₅₀ : 0.74	No decrease	No decrease
<i>N. casparyi</i> (28 d)	LC ₅₀ : 1.00	EC ₅₀ : 1.00	28 d: 1.00
<i>P. slavus</i> (16 d)	LC ₅₀ : 0.10	EC ₂₀ : 0.10	9 d: 0.05

Survival (LC₅₀), Behavior (EC₅₀) and response times (ET₂₀) at the lowest effective concentration.

Table 4: Cumulative number of neonates per female in *D. magna* exposed to different BPA concentrations (mg/L) over 28 d. Means (R = 3, N = 10) and SD (standard deviation).

BPA (mg/L)	Mean	SD
Control	93	18
Control-ethanol	150	20
0.03	187	54
0.3	136	32
3	175	53

the most sensitive species, followed by the groundwater isopod *P. slavus*, then *G. fossarum*, before *D. magna* and the stygal *N. casparyi* (Table 2). A more detailed comparison between the median lethal concentrations, LC₅₀s (at 18 - 20 °C) for several days was calculated from a separate experiment for the two standard species *D. magna* and *G. fossarum*, showing both species' similar sensitivity to BPA after 24 h (*G. fossarum*: 11.86 (8.6 - 16.2) mg/L; *D. magna*: 11.9 (9.8 - 14.8) mg/L, however, increasing sensitivity of *G. fossarum* after already 48 h (*G. fossarum*: 6.2 (4.6 - 8.4) mg/L; *D. magna*: 8.9 (6.9 - 11.5) mg/L, and even without overlap of the confidence intervals (95% CI) after 96 h (*G. fossarum*: 3.4 (2.3 - 5.2) mg/L; *D. magna*: 8.2 (6.2 - 10.9) mg/L. Moreover, the LC₅₀ 24 h values for both species were comparable to those of the 1st test at 10 °C.

Under BPA exposure the activity of most species declined with increasing concentration levels and decreasing response time of the decline. The median effect concentrations, EC₅₀24 h values (i.e. the concentration with a 50% decrease in activity) and the ET₂₀ (i.e. the time of 20% decrease in activity at the lowest effective concentration) are found in (Table 2). *P. slavus* was the most sensitive species regarding mortality within 24 h and rapidly decreasing activity already after 6 h in 2.5 mg/L BPA. However, regarding the EC₅₀ values after 24 h, *N. casparyi* was the most sensitive species with 5 mg/L BPA being the threshold for 20% decrease in activity, compared to 10.3 mg/L in *D. magna*, 13.5 mg/L in *G. fossarum* and 25 mg/L BPA in *P. slavus*.

Chronic toxicity tests

The chronic exposures revealed *P. slavus* to be the most sensitive species regarding survival and behavior effects of BPA (Table 3). Moreover, there was a trend of *D. magna* to be slightly more sensitive than the amphipods *G. fossarum* and *N. casparyi* regarding mortality. However, behavior of *D. magna* could not be evaluated as there was an overall increase in activity with age in all concentration levels and the controls masking potential effects of BPA.

G. fossarum consumed alder leaves and there was a significant decrease in feeding activity in all BPA-exposures and with increasing BPA concentration (Kruskal Wallis test $p < 0.01$; Figure 3). A 50% decline in feeding activity was almost reached at 0.5 mg/L BPA.

Daphnia magna showed no effects in molting frequency under BPA exposure (0.03, 0.3 and 3 mg/L) (Kruskal Wallis, not significant). However, effects of BPA on reproduction in *D. magna* could be seen at the end of the 28 d exposure at ≥ 3 mg/L BPA, even though not significant (Kruskal Wallis, not significant). The cumulative number of neonates was variable within the replicates of one treatment, the highest number of neonates was reached at 0.03 mg BPA/L, the lowest in the control (Table 4).

Discussion

Acute tests

BPA can be found in many polycarbonate plastics and epoxy resins [28-30]. BPA has been reported to be acutely toxic and function as an endocrine disruptor in aquatic organisms [28,31].

In the acute toxicity tests (24 h) the copepod *E. serrulatus* was the most sensitive test species regarding survival and rapid behavioral responses. This might be due to its small size, its rapid generation cycle and its feeding habits as fine particle feeder. However, *D. magna* larvae of similar size as *E. serrulatus* were less sensitive. They feed on planktonic algae and have

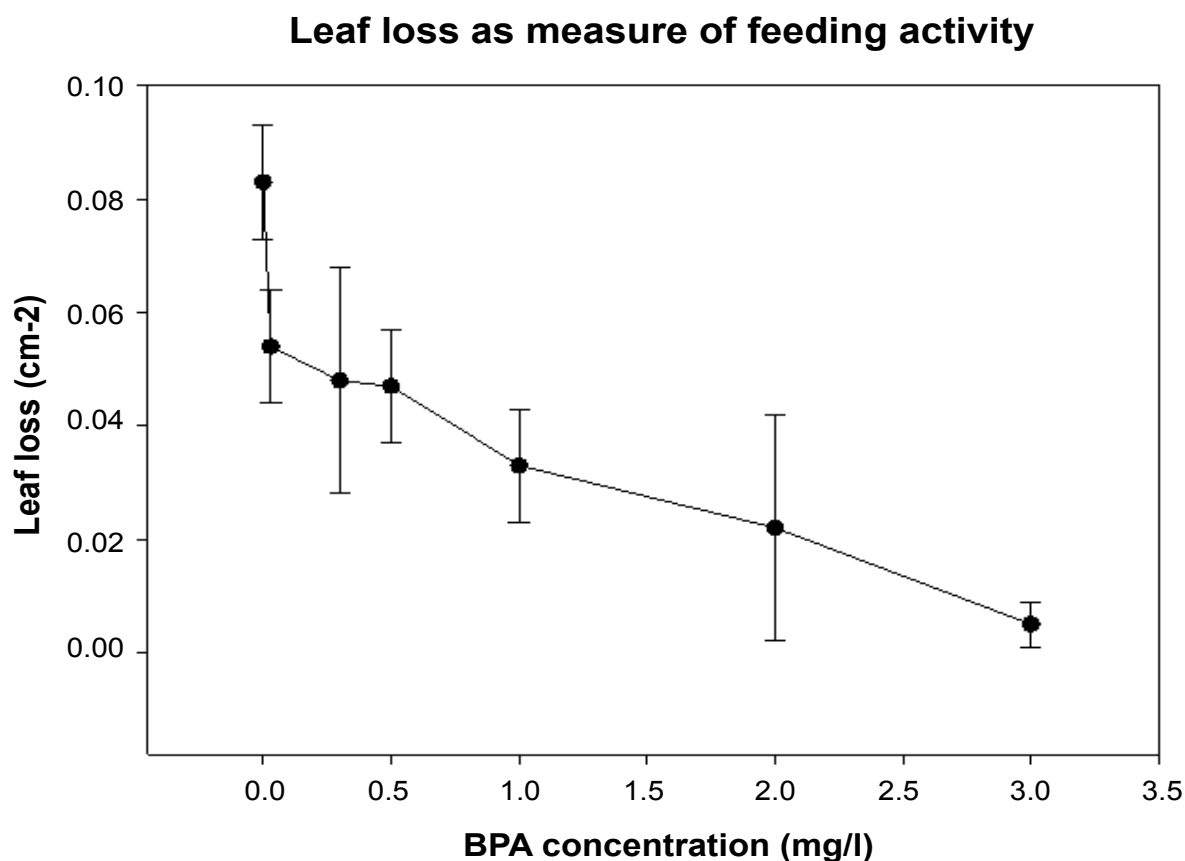


Figure 3: Feeding activity of *Gammarus fossarum*, exposed to BPA (28 d). Y-axis: Leaf loss (cm²). Means (R = 3, N = 5) and SD bars. X-axis: BPA concentrations (mg/L).

slower growth and a longer life cycle than the copepod, hence they might take up BPA less rapidly during short exposure times. Regarding the larger crustacean species, the groundwater species *P. slavus* reacted very rapidly and sensitively to BPA, compared to *N. casparyi* and *G. fossarum*. Regarding the overall effect on locomotory behavior, the EC₅₀-24 h, *N. casparyi* was the most sensitive species. These results show, that groundwater crustaceans tend to react both faster and more sensitive to BPA short-term exposure than the surface water representative *G. fossarum*. Generally, groundwater species have a thinner and transparent cuticula, i.e. less protection from the surrounding aquatic environment.

The surface water crustaceans *G. fossarum* and *D. magna* showed similar sensitivity in the more detailed and direct comparative acute toxicity test (24 - 96 h) at 18 - 20 °C. Whereas the 24 h LC₅₀ for *D. magna* (1 - 2 mm) was 11.9 mg/L (9.58 - 14.8), the value for *G. fossarum* (5 - 7 mm) was 13.41 mg/L (9.7 - 18.43). In the literature, LC₅₀ values for 24 h exposures were reported in *D. magna* neonates as follows: Brennan, et al. [32]: 8.57 mg/L (8.28 - 8.86), Plahuta, et al. [33]: 12.5 mg/L (11.3 - 14.1), Jemec, et al. [34]: 21 mg/L (20.8 - 23.8) for organisms of similar sizes. Moreover, acute toxicity (LC50 24 h) in neonates of *D. magna* required 7.9 mg/l, [35]. Watts, et al. [36] reported a 24 h LC₅₀ of 12.8 mg/L in *G. pulex*. The values of this study supported literature data.

After 24 h of acute exposure, *G. fossarum* became more sensitive to BPA than *D. magna*, as the 48 h LC₅₀ of *G. fossarum* decreased to 6.67 mg/L (4.8 - 9.1) compared to 8.94 mg/L (6.9 - 11.52) in *D. magna*. Similar results were previously found for *G. pulex* (5.6 mg/L, Watts, et al. [36]), and *D. magna* (7.57 mg/L, Brennan, et al. [32]). The few literature studies on freshwater amphipods *G. pulex* and *G. fossarum* revealed effects on survival at 12.8 mg/l (LC₅₀ 24 h), 4 mg/l (LC₅₀ 72 h), 1.9 mg/l (LC₅₀ 96 h) and 1.4 mg/l (LC₅₀ 10 d) [36].

The difference in sensitivity between *G. fossarum* and *D. magna* increased from 48 h to 96 h exposure even further. This shows that test duration, even in acute tests, can drastically affect the test results. Up to now, *D. magna* was always regarded the most sensitive aquatic sentinel species in aquatic ecotoxicology. Moreover, it was always believed that small organisms should be more sensitive than larger organisms due to the larger surface/volume ratio. However, in this study the small daphnids were less sensitive than the larger gammarids towards BPA. Moreover, filter feeders such as daphnids should be more sensitive than detritivores, as they filter great amounts of water (and dissolved toxins) through their body. The moderate toxicity of BPA to *D. magna* (EC₅₀-48 h: 10 mg/L) was also stated by Chen, et al. [37].

The marine rotifer *Brachionuskoreanus* showed effects of BPA, BPF and BPS, such as increased intracellu-

lar ROS (reactive oxygen species) and GST activities and gene expression modulation of cytochrome P450 [10]. Similar biochemical responses might be underlying the observed reduced mortality and locomotory activity in the crustacean species of the current study.

Moreover, fish toxicity studies on BPA revealed an LC_{50} (72 h) for *Poecilla reticulata* of 1.6 mg/L, which is more sensitive than the previously reported toxic range for fish of 6 - 17.9 mg/L [38]. The acute effects (48 h) of BPF in zebrafish larvae *Danio rerio* included disrupted glucose metabolism at > 10 µg/L, showing the rapidity of physiological responses at low concentration levels [8].

However, the acute toxicity levels found in this study are higher than the BPA levels reported in the field. The highest BPA concentrations in surface waters were 21 µg/L [39] and 17.2 mg/L in landfill leachates [40]. BPA levels in surface water in Germany range from 0.5 - 5 µg/l, in a stream in an industrial area of Norway a concentration of 43 µg/l was reported, whereas values in water of Lake Constance taken for drinking water were as low as 2 ng/l [3].

The sensitivity of the studied surface and stygal crustacean species might therefore help to indicate landfill leachate spills or water quality in areas close to landfills or water treatment plants. On the other hand, the copepod *E. serrulatus* might be used to monitor BPA pollution in surface water, e.g. taken for drinking water preparation.

Chronic tests

During chronic BPA exposures the stygal isopod *Proasellus slavus* was the most sensitive species regarding both survival and behavior, followed by *G. fossarum* (esp. feeding behavior), and last *Niphargopsis casparyi*. Regarding survival, *G. fossarum* was as sensitive as *D. magna*. (*G. fossarum* LC_{50} 28 d: 0.80 ± 0.12 mg/L; *D. magna* 0.74 ± 0.25 mg/L). Watts, et al. [36] found an LC_{50} 10 d for *G. pulex* at 1.49 mg/BPA/L. Chronic exposures (21 d) revealed an increased number of daily molting in *D. magna* at 0.02 mg/L, increased dry weight at 2 mg/L, effects on body length at 8.2 mg/L and fecundity at 7.3 mg/L [35].

The No Observed Effect Concentration (NOEC) for stimulation of substance avoidance was found in *G. pulex* at 8.4 mg/L BPA, the Lowest Observed Effect Concentration (LOEC) being 19.4 mg/L (CAS 80057). *G. fossarum* responded sensitively with decreased feeding activity as alder leaf loss of about 40% was found in all BPA exposures, however, significant differences were only found at ≥ 1 mg/L. In BPA concentrations of 2 and 3 mg/L, the reduction of feeding activity was 75% and 95%, respectively, indicating high impact of BPA compared to the control (< 0.05). The feeding behavior of invertebrates is a very sensitive parameter. The reduction of the feeding activity is among the first responses to stress or environmental pollution [41].

N. casparyi and *P. slavus* starve for long periods as an adaptation to stygal environmental conditions. As feeding habits are still not known, both fine detritus from the sampling site and alder leaves were added, however the leaves were not touched by these species.

In arthropods, molting is necessary to allow for growth, being regulated by ecdysteroid hormones [42]. In this study, the cumulative molting of organisms was counted as *D. magna* has a short life cycle with many moltings. This was not possible in the tests with the amphipods and the isopod, as they have longer molting intervals, generating too little data for statistical analysis.

D. magna showed ca. 9 molts per female over 28 d exposure, supporting findings by Brennan, et al. [32], where on average females molted 9 times in a 21- day exposure to BPA. However, no significant differences between the molting of organisms exposed to BPA and the control could be seen. Mu, et al. [42] reported inhibition of molting in *D. magna* in BPA concentrations ranging from 5 to 10 mg/L. This is consistent with the results found in this study, as the molting of *Daphnia magna* exposed to concentrations of 3 mg/L and lower was not affected. However, Li, et al. [35] reported molting in *D. magna* to be inhibited at already 0.02 mg/L.

Another important parameter in the standard test procedures of *D. magna* represents the reproduction, which might be affected by BPA, known as endocrine disruptor for fish and amphibians and esp. snails [43]. *D. magna* has a short life cycle and reproduces several times during the 28 d exposure. The cumulative number of neonates per female was lowest in the control and higher in all other treatments, indicating rather an effect of ethanol than BPA. Ethanol might reduce bacterial and fungal growth in the beakers, hence support growth and reproduction of daphnids. As the number of neonates was similar in all BPA concentrations, no dose-dependent adverse effects of BPA on the organisms were observed. The results agree with those by Brennan, et al. [32] who counted around 70 neonates per female on a 21 d exposure to BPA. Jemec, et al. [34] found BPA affected the reproduction of *D. magna* only above 3.45 mg/L. Li, et al. [35] reported for *D. magna* under chronic exposure to BPA, lignin-derived (LD)-BPA and mixed exposures drastic adverse effects of the mixture at 2 mg/L on fecundity and enzyme activities. Lee, et al. [44] found adverse effects on reproduction in the midge *C. riparius* as well as increased DNA damage at 1 mg/L BPA. The aquatic oligochaete *Lumbriculus variegatus* responded to chronic exposures to BPA or BPS with retarded regeneration ability after fragmentation and increased blood vessel pulse rates at > 10 - 6 M [45].

After 28 d of exposure the snail *P. antipodarum* showed an increased number of embryos in the brood pouch at 0.04 mg/L [43]. Whereas survival of the nematode *C. elegans* was not affected by BPA (up to

10 μM), head thrashes (as a measure for locomotion) was decreased at 0.001 μM BPA already during 10 d of exposure [46]. BPA impaired the reproduction of the oligochaete *Eisenia fetida* as number of juveniles decreased at 1 mg/L, whereas mortality increased at 2 mg/L (14 d) [47].

Endocrine disruptive effects have been reported for fish and amphibians at concentrations around 16 $\mu\text{g/L}$, whereas aquatic insects and crustaceans appear to be more tolerant with effect levels between 100 - 3146 $\mu\text{g/L}$ [3].

Effects on reproduction were found in Japanese Medaka after 60 d of exposure at levels of 1.5 mg/L, such as a reduced number of broods and total number of eggs [48]. No effects of BPA levels up to 1 mg/L were found in *Danio rerio* survival (120 d) [9]. However, Wei, et al. [7] proved adverse effects of BPS on the thyroid endocrine system with altered T3 and T4 levels, delayed development, neurotoxicity and reduction in motility at levels below 0.1 mg/L BPS in a 120 d long study. Such multiple effects of BPS in zebrafish were also reported by Gu, et al. [49], including decreased locomotory behavior, enhanced apoptosis, altered renal structure and increased oxidative stress at > 0.3 mg/L.

Conclusion

This study provides for the 1st time comparative toxicity data (acute, chronic) of BPA on both surface water and groundwater crustaceans.

E. serrulatus proved to be the most sensitive species in acute exposures to BPA regarding survival and locomotory behavior and might be used to monitor acute toxic pulses of bisphenols.

The most sensitive macro-crustacean species was the groundwater isopod *P. slavus*, in both acute and chronic exposures. *P. slavus* might be used for long-term monitoring of chronic low-dose exposures, e.g. in groundwater and drinking water.

G. fossarum (5 - 7 mm) proved to be more sensitive to BPA compared to the standard test species *D. magna* (1 - 2 mm) regarding survival, behavior and esp. feeding as the most sensitive parameter.

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References

- Jalal N, Surendranath AR, Pathak JL, Yu S, Chung CY (2018) Bisphenol A (BPA) the mighty and mutagenic. Toxicol Reports 5: 76-84.
- Murata M, Kang JH (2017) Bisphenol A and all cell signaling pathways. Biotechnology Advances 36: 311-327.
- UBA (2010) BPA, Massenchemikalie mit unerwünschten Nebenwirkungen.
- Liu M, Jia S, Dong T, Han Y, Xue J, et al. (2019) The occurrence of bisphenol plasticizers in paired dust and urine samples and its association with oxidative stress. Chemosphere 216: 472-478.
- Crain DA, Eriksen M, Iguchi T, Jobling S, Laufer H, et al. (2007) An ecological assessment of bisphenole-A: Evidence from comparative biology. Reprod Toxicol 24: 225-239.
- Avissar-Whiting M, Veiga KR, Uhl KM, Maccani MA, Gagne LA, et al. (2010) Bisphenol A exposure leads to specific microRNA alterations in placental cells. Reprod Toxicol 29: 401-406.
- Wei P, Zhao F, Zhang X, Liu W, Jiang G, et al. (2018) Transgenerational thyroid endocrine disruption induced by bisphenol S affects early development of zebrafish offspring. Environ Pollut 243: 800-808.
- Zhao F, Wang H, Wie P, Jiang G, Wang W, et al. (2018) Impairment of bisphenol F on the glucose metabolism of zebrafish larvae. Ecotoxicol Environ Saf 165: 386-392.
- Qiu W, Shao H, Lei P, Zheng C, Qiu C, et al. (2018) Immunotoxicity of bisphenol S and F are similar to that of bisphenol A during zebrafish early development. Chemosphere 194: 1-8.
- Park JC, Lee MC, Yoon DS, Han J, Kim M, et al. (2018) Effects of bisphenol A and its analogs BPF and BPS on life parameters, antioxidant system, and response of defense in the marine rotifer *Brachionus koreanus*. Aquat Toxicol 199: 21-29.
- Ashfaq M, San Q, Zhang H, Le Y, Wang J, et al. (2018) Occurrence and fate of bisphenol A transformation products bisphenol A monomethyl ether and bisphenol A dimethyl ether in wastewater treatment plants and surface water. J Hazard Mater 357: 401-407.
- www.lfu.bayern.de
- Flint S, Markle T, Thompson S, Wallace E (2012) Bisphenol A exposure, effects, and policy: A wildlife perspective. J Environ Manage 104: 19-34.
- Baderna D, Maggioni S, Boriani E, Gemma S, Molteni M, et al. (2011) A combined approach to investigate the toxicity of an industrial landfill's leachate: Chemical analyses, risk assessment and in vitro assays. Environ Res 111: 603-613.
- Zhang H, Zhang Y, Li J, Yang M (2018) Occurrence and

- exposure assessment of bisphenol analogues in source water and drinking water in China. *The Science of the Total Environment*.
16. UBA (2015) Neue grenzwerte für die massenchemikalie Bisphenol A.
 17. Arnich N, Canivenc-Lavier MC, Kolf-Clauw M, Coffigny H, Cravedi JP, et al. (2011) Conclusions of the french food safety agency on the toxicity of bisphenol A. *Int J Hyg Environ Health* 214: 271-275.
 18. <https://www.epa.gov/chemical-research/ecotoxicology-database>.
 19. Grimm C, Gerhardt A (2018) Sensitivity towards copper: Comparison of stygal and surface water species' biomonitoring performance in water quality surveillance. *Intern J Sci Res Environm Sci Toxicol* 3: 15.
 20. Rétaux S, Casane D (2013) Evolution of eye development in the darkness of caves: Adaptation, drift or both? *EvoDevo* 4: 26.
 21. Hahn HJ (2006) The GW-Fauna-Index: A first approach to a quantitative ecological assessment of groundwater habitats. *Limnologica* 36: 119-137.
 22. Di Lorenzo T, Di Marzio WD, Cifoni M, Fiasca B, Baratti M (2015) Temperature effect on the sensitivity of the copepod *Eucyclops serrulatus* to agricultural pollutants in the hyporheic zone. *Current Zoology* 61: 629-640.
 23. Di Lorenzo T, Di Marzio WD, Spigoli D, Baratti M, Messina G, et al. (2015) Metabolic rates of a hypogean and an epigeal species of copepod in an alluvial aquifer. *Freshwater Biology* 60: 426-435.
 24. Bork J, Hahn H J (2008) Groundwater and biodiversity. Presentation at the symposium "Biodiversität von Gewässern, Auen und Grundwasser", Bundesamt für Naturschutz, Bonn, 29-30.
 25. Gerhardt A, Clostermann M, Fridlund B, Svensson E (1994) Monitoring of behavioral patterns of aquatic organisms with an impedance conversion technique. *Environment International* 20: 209-219.
 26. Gerhardt A, Carlsson A, Ressemann C, Stich KP (1998) New online biomonitoring system for *Gammaruspulex* (L.) (Crustacea): In situ test below a copper effluent in South Sweden. *Environmental Science & Technology* 32: 150-156.
 27. Gerhardt A (2018) Suitability of *Eucyclops serrulatus* (Fischer 1851) (Crustacea: Copepoda) for online biomonitoring of water quality in the new microimpedance sensor system®. *Res Trends Ecotoxicol*.
 28. Kang JH, Aasi D, Katayama Y (2007) Bisphenol A in the aquatic environment and its endocrine-disruptive effects on aquatic organisms. *Crit Rev Toxicol* 37: 607-625.
 29. Alexander HC, Dill DC, Smith LW, Guiney PD, Dorn P (1988) Bisphenol A: Acute aquatic toxicity. *Environm Toxicol Chem* 7: 19-26.
 30. Staples CA, Dome PB, Klecka GM, Oblock ST, Harris LR (1998) A review of the environmental fate, effects, and exposures of bisphenol A. *Chemosphere* 36: 2149-2173.
 31. Chapin RE, Adams J, Boekelheide K, Gray LE, Hayward SW, et al. (2008) NTP-CERHR expert panel report on the reproductive and developmental toxicity of bisphenol A. *Birth Defects Res B Dev Reprod Toxicol* 83: 157-395.
 32. Brennan SJ, Brougham CA, Roche JJ, Fogarty AM (2006) Multi-generational effects of four selected environmental estrogens on *Daphnia magna*. *Chemosphere* 64: 49-55.
 33. Plahuta M, Tišler T, Toman MJ, Pintar A (2014) Efficiency of advanced oxidation processes in lowering bisphenol A toxicity and oestrogenic activity in aqueous samples. *Arh Hig Rada Toksikol* 65: 77-87.
 34. Jemec A, Tisler T, Erjavec B, Pintar A (2012) Antioxidant responses and whole-organism changes in *Daphnia magna* acutely and chronically exposed to endocrine disruptor bisphenol A. *Ecotoxicol Environ Saf* 86: 213-218.
 35. Li D, Chen H, Bi R, Xie H, Zhou Y, et al. (2018) Individual and binary mixture effects of bisphenol A and a lignin-derived bisphenol in *Daphnia magna* under chronic exposure. *Chemosphere* 191: 779-786.
 36. Watts MM, Pascoe D, Carroll K (2001) Survival and precopulatory behaviour of *Gammaruspulex* (L.) exposed to two xenoestrogens. *Water Research* 35: 2347-2352.
 37. Chen MY, Ike M, Fujita M (2002) Acute toxicity, mutagenicity and estrogenicity of BPA and other bisphenols. *Environ Toxicol* 17: 80-86.
 38. Silva DCVR, Araujo CVM, Franca FM, Neto MB, Paiva TCB, et al. (2018) Bisphenol risk in fish exposed to a contamination gradient: Triggering of spatial avoidance. *Aquat Toxicol* 197: 1-6.
 39. Belfroid A, Van Velzen M, Van Der Horst B, Vethaak D (2002) Occurrence of bisphenol A in surface water and uptake in fish: Evaluation of field measurements. *Chemosphere* 49: 97-103.
 40. Yamamoto T, Yasuhara A, Shiraishi H, Nakasugi O (2001) Bisphenol A in hazardous waste landfill leachates. *Chemosphere* 42: 415-418.
 41. Alonso Á, De Lange HJ, Peeters ET (2009) Development of a feeding behavioural bioassay using the freshwater amphipod *Gammarus pulex* and the multispecies freshwater biomonitor. *Chemosphere* 75: 341-346.
 42. Mu X, Rider CV, Hwang GS, Hoy H, LeBlanc GA (2005) Covert signal disruption: Anti-ecdysteroidal activity of bisphenol A involves cross talk between signaling pathways. *Environ Toxicol Chem* 24: 146-152.
 43. Sieratowicz A, Stange D, Schulte-Oehlmann U, Oehlmann J (2011) Reproductive toxicity of bisphenol A and cadmium in *Potamopyrgus antipodarum* and modulation of bisphenol A effects by different test temperature. *Environ Pollut* 159: 2766-2774.
 44. Lee SW, Chatterjee N, Im JE, Yoon D, Kim S, et al. (2018) Integrated approach of eco-epigenetics and eco-metabolomics on the stress response of BPA-exposure in the aquatic midge, *Chironomus riparius*. *Ecotoxicol Environ Saf* 163: 111-116.
 45. Vought V, Wang HS (2018) Impact of common environmental chemicals BPA and BPS on the physiology of *Lumbricus variegatus*. *Environ Toxicol Pharmacol* 60: 225-229.
 46. Zhou D, Yang J, Li H, Cui C, Yu Y, et al. (2016) The chronic toxicity of bisphenol A to *Caenorhabditis elegans* after a long-term exposure at environmentally relevant concentrations. *Chemosphere* 154: 546-551.
 47. Verdu L, Trigo D, Martinez-Guitarte JL, Novo M (2018) Bisphenol A in artificial soil: Effects on growth, reproduction and immunity in earthworms. *Chemosphere* 190: 287-295.
 48. Li D, Chen Q, Cao J, Chen H, Li L, et al. (2016) The chronic effects of lignin-derived bisphenol and bisphenol A in Japanese medaka *Oryzias latipes*. *Aquatic Toxicology* 170: 199-207.
 49. Gu J, Zhang J, Chen Y, Wang H, Guo M, et al. (2018) Neurobehavioural effects of BPS exposure in early life stages of zebrafish larvae. *Chemosphere*.