



Oxidative Stress and the Epigenome in Human Disease

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Abstract

Epigenetics refers to the study of the changes in gene expression that occur without changes in the DNA sequence. There is growing evidence that epigenetic modifications such as changes in the levels of DNA methylation or post-translational histone modifications are involved in the pathogenesis of many human diseases including cancer. Oxidative stress as a result of metabolic or environmental factors leads to excessive production of reactive oxygen species (ROS). ROS plays a role in many human diseases including cancer and pulmonary and cardiovascular diseases by promoting DNA damage and/or altering signaling pathways. This review article summarizes the most recent reports linking both oxidative stress and epigenetic mechanisms in the pathogenesis of chronic obstructive pulmonary disease (COPD), cardiovascular disease, and lung, prostate and colorectal cancers. Here, we emphasize the importance that future studies should focus on epigenetic intervention strategies to treat diseases associated with oxidative stress.

Keywords

Oxidative stress; Reactive oxygen species; DNA methylation; Post-translational histone modifications; Chronic obstructive pulmonary disease; Atherosclerosis; Non-small cell lung cancer; Prostate cancer; Colorectal cancer

Introduction

The term “oxidative stress” refers to the state of a cell characterized by an imbalance between the production of reactive oxygen species (ROS) and the cell’s detoxification defense system, favoring a ROS-rich environment and/or reduced antioxidant reserves. Under normal circumstances, ROS are produced during physiological processes, such as cellular respiration, the activation of the arachidonic acid cascade and by enzymes, including cytochrome p450, nicotinamide adenine dinucleotide (NADH)/Nicotinamide Adenine Dinucleotide Phosphate (NADPH) oxidase and nitric oxide synthase. Oxidative stress is associated with numerous medical conditions, including pulmonary and cardiovascular diseases, as well as cancer.

At ground state, molecular oxygen (O_2), required for aerobic metabolism, has two unpaired electrons, and can readily accept others. The mitochondria’s electron transport chain uses O_2 as a terminal acceptor for the electrons from NADH and generates a proton motive force. However, if leakage of electrons occurs, O_2 is

rapidly reduced to the superoxide anion ($O_2^{\cdot-}$). Although $O_2^{\cdot-}$ is not reactive itself, it can initiate the generation of ROS. Thus, it is necessary that cells have effective mechanisms for removing $O_2^{\cdot-}$ and ROS. The generation of ROS may cause a wide range of DNA lesions including base modifications, deletions, strand breakage, and chromosomal rearrangements [1,2]. Oxidative stress can either drive genetic mutations or epigenetically regulate the expression of genes. The cellular antioxidants must respond to an overproduction of ROS before these highly reactive molecules adversely alter cellular structures, including DNA, proteins, and lipids. Severe oxidative stress may trigger apoptosis, necrosis, and cell death.

ROS are detoxified within the cell by several kinds of antioxidants [3]. Examples of endogenous defenses against ROS include the antioxidant enzymes glutathione-S-transferase P (GSTP1), glutathione peroxidase, catalase, superoxide dismutase (SOD), peroxiredoxin, and sulfiredoxin [3]. Examples of low molecular weight antioxidants include: glutathione, vitamin C, Vitamin A, and vitamin E [3]. Humans have three separate superoxide dismutases to reduce the $O_2^{\cdot-}$ from the cell: the cytoplasmic Cu/Zn SOD1, the mitochondrial manganese SOD2, and the extracellular SOD3 enzyme.

Most of the chromatin in mammalian cells exists in a condensed, transcriptionally silent heterochromatic form. Euchromatin is less condensed, and contains most of the actively transcribed genes. Epigenetics refers to the study of a stably heritable phenotype that results from changes in the chromatin that alter gene expression without alterations in the DNA sequence [4]. DNA and histone proteins can be chemically modified with epigenetic marks that alter the electrostatic nature of the chromatin or alter the affinity of chromatin-binding proteins. The chromatin structure, or the “epigenome”, is regulated by a large number non-coding RNAs and histone-modifying and DNA methylation enzymes [5]. The three major mechanisms of epigenetic regulation include DNA methylation, post-translational histone modifications, and non-coding RNAs including micro RNAs.

DNA methylation plays an important role in embryonic development, genomic imprinting, X-chromosome inactivation, and the preservation of chromosome stability. DNA methylation at the promoter region of genes is associated with repression of gene transcription by maintaining the chromatin in a closed state [6]. During DNA methylation a methyl group is added to the carbon-5 position of the cytosine pyrimidine ring by DNA methyltransferases

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to form 5-methylcytosine (5-MeC) [6]. The chromatin is maintained in a closed state by recruitment of the methyl-CpG-binding-domain protein complexes that also contain HDACs that remove acetyl groups from the histone's N-terminal domains and keep the chromatin in a closed configuration making the chromatin inaccessible to transcription factors and co-activators [7,8]. A family of DNA methyltransferase enzymes (DNMTs) is involved in *de novo* DNA methylation and methylation maintenance. DNMT1 is predominantly responsible for maintaining cellular levels of CpG methylation whereas DNMT3A and DNMT3B are critical for *de novo* methylation during embryogenesis [9]. The absence of 5-MeC in DNA promoters allows acetylation of histones permitting a number of transcription factor complexes to access the chromatin and promote transcription of a specific genomic region [8].

Post-translational histone modifications include lysine acetylation, arginine and lysine methylation, serine phosphorylation, and lysine ubiquitination, and sumoylation. Lysine acetylation is usually associated with transcriptional activation but the functional consequences of lysine and arginine methylation depend on the specific site of the residue within the histone tail [10-13]. For example, methylation of histone H3 at lysine 4 is linked to transcriptional activation, whereas methylation of histone H3 at lysine 9 or lysine 27 is associated with transcriptional repression [11-13]. The post-translational histone modifications allow the chromatin to have a dynamic structure and constitute the docking site for distinct chromatin-binding proteins; for example, the histone acetyltransferases (HATs) and their counterpart, the histone deacetylases (HDACs), or the histone methyltransferases (HMTases), and their opposite, the histone demethylases, direct between a transcriptionally active or transcriptionally silent chromatin [14]. The "histone code" is now also widely accepted and states that specific histone modifications on the same or different histone tails act sequentially or in combination regulate the expression of a specific region within the chromatin [15]. Dysregulated histone post-translational modifications have been shown to be important in both predictive and prognostic value in various diseases such as cancer [16-21].

Non-coding RNAs (ncRNAs) have changed the view of the "central dogma" in that these play fundamental roles in regulating protein levels by modulating transcription and translation to either ultimately increase or decrease protein levels. Small ncRNAs include PIWI-interacting RNAs (piRNAs), transition initiation RNAs (tiRNAs), and microRNAs (miRNAs). Mid-size ncRNAs include small nucleolar RNAs (snoRNAs), promoter upstream transcripts (PROMPTS), and transcription start sites (TSS)-associated RNAs (TSSa-RNAs). Long-ncRNAs include circular RNAs, transcribed ultra-conserved regions (T-URCs) and large intergenic ncRNAs (lincRNAs) [22]. miRNAs can regulate downstream gene expression by binding to the 3' untranslated region (UTR) of an mRNA resulting in mRNA degradation and translational repression [22,23]. Long-ncRNAs are mostly known to modulate the chromatin structure, and thus change DNA condensation, resulting in less transcription [22]. Small and mid-size ncRNA regulate transcription and translation. Some long ncRNAs even regulate the expression of other microRNAs.

There is a growing amount of evidence that both epigenetics and oxidative stress may be linked in the pathology of various human diseases. In this review, we discuss the recent studies on the role that epigenetics and oxidative stress play in chronic obstructive pulmonary disease (COPD), cardiovascular disease, and lung, prostate, and colorectal cancers and how exploring these fields may allow the identification of new therapeutic targets.

Chronic Obstructive Pulmonary Disease (COPD)

The lungs are exposed to numerous sources of endogenous and exogenous sources of oxidants derived from mitochondrial respiration, phagocyte activation, air pollutants, noxious gases, and cigarette smoking [24]. Chronic obstructive pulmonary disease (COPD) represents the fourth cause of mortality worldwide, and

is characterized as a group of disorders with similar respiratory symptoms, including cough, sputum production, systemic inflammation, obstruction of lung airflow, and decreases in respiratory function [25]. COPD patients also have an increased risk of developing lung cancer.

Cigarette smoke contains a number of free radicals and chemical compounds, representing the major source of inhaled ROS leading to the deregulated expression of pro-inflammatory genes [26,27]. Recently it was found that cigarette smoke post-translationally modifies histone deacetylase 2 (HDAC2), a class I histone deacetylase, resulting in a reduction of its enzymatic activity [26]. A smoke-dependent HDAC2 inactivation by post-translational phosphorylation via casein protein kinase 2 (CK2) was also reported in macrophages, human bronchial and primary small airway lung epithelial cells and, *in vivo*, in the mouse lung [28]. The inactivation by phosphorylation of HDAC2 results in its ubiquitination and proteosomal degradation. In COPD patients, inflammation and cellular senescence are exacerbated by tobacco smoke [25]. A decreased HDAC2 activity has been associated with inflammation and senescence in COPD patients resulting in an increase in H3 and H4 acetylation, the activation of nuclear factor of kappa light polypeptide gene enhancer in B-cells (NF- κ B) transcription factor, and deregulated expression of proinflammatory genes [28-30]. Levels of the NAD (+) dependent histone deacetylase sirtuin 1 (SIRT1) have also been shown to be reduced in patients with COPD, further demonstrating that HDAC expression and oxidative stress is associated with COPD [31].

Cardiovascular Disease

Cardiovascular diseases are the leading cause of death in industrialized nations [32]. There is a growing body of evidence suggesting that both epigenetic modifications and oxidative stress may play a role in the pathogenesis of many cardiovascular diseases.

Nitric oxide synthases (NOSs) play an important role in cardiovascular diseases and are known to be epigenetically modulated. Nitric Oxide (NO) plays an important cardioprotective role against cardiovascular diseases by regulating blood pressure, vascular tone, and inhibiting platelet aggregation and leukocyte adhesion. NO is produced by three isoforms of NOS encoded by separate genes on different chromosomes: neuronal NOS (*NOS1*), inducible NOS (*iNOS* or *NOS2*), and endothelial NOS (*eNOS* or *NOS3*). *eNOS* is constitutively expressed and responsible for the majority of NOS produced by the vascular endothelium and, therefore, represents the major source of bioactive NO. Methylation plays a role in the expression of the various forms of NOS. For example, *iNOS* is expressed in atherosclerotic plaque, but repressed by methylation in most tissues. The reaction of NO with superoxide forms peroxynitrite and decreases NO bioavailability, which enhances cellular oxidative stress. Peroxynitrite increases endothelial dysfunction and stimulates prothrombotic effects such as increased platelet reactivity and lipid peroxidation. Inactivation of NO by ROS is recognized as a key mechanism underlying the reduced NO availability and the development of endothelial dysfunction, which may be an important contributor to disease pathophysiology.

Cancer

The "epigenetic progenitor" model of human cancer proposed by Feinberg, Ohlsson, and Henikoff states that epigenetic changes in gene expression impact carcinogenesis through aberrant silencing of tumor suppressors genes and the improper activation of oncogenes [33]. Further epigenetic derangements and genetic mutations are acquired as this epigenetically altered progenitor population expands, ultimately leading to carcinogenesis.

Oxidative stress has been clearly linked to the development of various cancers. Oncogenic-driven cancer cells generate increased ROS as byproducts of their augmented metabolism to promote and maintain tumorigenicity [34-36]. Since high levels of ROS can induce cell death, cancer cells adapt to ROS stress by upregulating intracellular antioxidant proteins in order to maintain ROS levels that

allow protumorigenic signaling without resulting in cell death [37-42]. In fact, studies have shown that disabling antioxidant mechanisms triggers ROS-mediated cell death in many forms of human cancers [43-46]. Increasing evidence also has linked the regulation of many pathways associated the homeostasis of oxidative stress to epigenetic mechanisms [47].

Lung cancer is the leading cause of cancer deaths worldwide and only 13% of lung cancer patients survive more than 5 years [47]. Non-small cell lung cancers (NSCLCs) represent 80% of all lung cancers and are often diagnosed at an advanced stage with poor prognosis. SOD1 has been shown to be over expressed at higher levels in lung adenocarcinomas, a subtype of NSCLC [48]. SOD1 converts superoxide to hydrogen peroxide (H_2O_2) and molecular oxygen in the cytosol, the nucleus, and the intermembrane space of the mitochondria. SOD1 protects the cell from oxidative stress and subsequent cell death by maintaining low levels of superoxide in the cytosol. A more recent study reported that inhibition of SOD1 by the small molecule ATN-224 reduced tumor burden in a mouse model of NSCLC suggesting a potential clinical application for the treatment of patients with various forms of NSCLC [49]. ATN-224-dependent SOD1 inhibition in various NSCLC cells increased superoxide, diminished the enzyme activity of the antioxidant glutathione peroxidase, and increased intracellular levels of H_2O_2 .

Elevated levels of HDAC1 mRNA have been reported in more advanced stages of this disease (Stage III or IV) [31,50]. Murine switch-independent 3-associated (mSin3A), a critical scaffold on which the multi-component HDAC co-repressor complex assembles, has also been reported to have decreased expression in NSCLC [31,50]. Additionally, the ATP-dependent SWI/SNF chromatin remodeling complexes members have been reported to be dysregulated in NSCLC [31,50]. In NSCLC, mutations are also found within the lysine acetyltransferase KAT3A in a small subset of patients and polymorphisms have been identified which are associated with an increased risk for lung cancer including the lysine methyltransferases KMT1B and KMT8 [31]. Polymorphisms in the methyl-CpG binding domain 1 (MBD-1) have been associated with an increased risk of developing lung cancer [31].

In the United States, prostate cancer (CaP) is the most commonly diagnosed non-skin cancer and the second-leading cause of cancer deaths [51]. Several studies have reported decreased levels of Erythroid 2p45 (NF-E2)-related factor 2 (NRF2) and members of the glutathione-S-transferase (GST) mu family in human CaP [52]. NRF2 is a basic-region leucine zipper (bZIP) transcription factor that regulates the expression of phase II detoxifying/antioxidant enzymes, including glutathione-S-transferase (GST), UDP-glucuronosyltransferase (UGT), hemeoxygenase-1 (HO-1), NADPH: quinoneoxidoreductase (NQO), glutamate cysteine ligase (CGL) and gamma glutamylcysteine synthase (γ GCS), by binding in combination with small Maf proteins to antioxidant response elements (AREs) in promoter regions [53]. The expression of NRF2 in prostate tumors from TRAMP mice has also been shown to be suppressed epigenetically by promoter CpG methylation and histone modifications [54]. The treatment of TRAMP cells with the cytosine methylation inhibitor, 5-aza-2-deoxycytidine (5-aza-dC), and the histone deacetylase inhibitor trichostatin A (TSA) restored NRF2 expression and increased the expression of NRF2 and its downstream antioxidant and detoxification enzymes [54,55]. Three specific CpG sites in the NRF2 promoter were found to be hypermethylated in clinical CaP samples [56]. CpG sites showed methylation that inhibited the transcriptional activity of NRF2 in LNCaP cells but LNCaP cells treated with 5-aza/TSA restored the expression of NRF2 and its downstream target genes, decreased expression levels of DNMT and HDAC proteins, increased RNA Pol II and H3Ac, and decreased H3K9me3, MBD2, and MeCP2 at CpG sites of the human NRF2 promoter [56]. Moreover, the expression and activity of SOD, catalase, and GPx have also been found to be decreased in plasma, erythrocytes, and CaP tissues confirming the role of NRF2 and its target genes in controlling oxidative stress in CaP and confirming

the existence of an epigenetic mechanism involved in its regulation [57,58].

A recent study reported that ROS silenced the tumor suppressor, RUNX3, by epigenetic regulation and may be associated with the progression of colorectal cancer [59]. The runt-domain transcription factor 3 (RUNX3) is known to be a tumor suppressor involved in various cancers, including gastric cancers [60-63]. Approximately 45-60% of human gastric cancers have been reported to display loss of RUNX3 expression [64]. The Kang et al. [59] study reported that RUNX3 mRNA and protein expressions were down-regulated in response to H_2O_2 in the SNU-407 human colorectal cancer cell line. H_2O_2 treatment increased RUNX3 promoter methylation and the ROS scavenger, N-acetylcysteine (NAC) and 5-aza-dC, decreased it. The downregulation of RUNX3 was also abolished with pretreatment of NAC. 5-aza-dC treatment prevented the decrease in RUNX3 mRNA and protein levels by H_2O_2 treatment. Additionally, this same study also reported that H_2O_2 treatment resulted in DNMT1 and HDAC1 up-regulation with increased expression and activity, increased binding of DNMT1 to HDAC1, and increased DNMT1 binding to the RUNX3 promoter. H_2O_2 treatment also inhibited the nuclear localization of RUNX3, which was also abolished by NAC treatment. When RUNX3 is translocated to the nucleus it acts as a tumor suppressor; however, cytoplasmic RUNX3 does not elicit tumor suppressor activity [65].

DNA methylation and down-regulation of CDX1 has been observed in a number of colorectal carcinoma derived cell lines and in patient samples. The Zhang et al. [66] study examined whether oxidative stress regulated the expression of the caudal type homeobox-1 (CDX1) tumor suppressor gene in colorectal cancer cells [66]. The results of the study suggested that silencing of CDX1 expression by oxidative stress in colorectal cancer cells may be mediated by epigenetic mechanisms. Additionally, treatment with H_2O_2 down regulated CDX1 mRNA level and protein expression in the T-84 human colorectal cancer cell line. The down regulation of CDX1 at the mRNA and protein level induced by H_2O_2 was further abolished by separate treatment of either NAC or 5-aza-dC. Treatment with H_2O_2 also increased CDX1 promoter methylation and 5-aza-dC reversed this effect. In this same study, H_2O_2 also induced the up regulation of DNMT1 and HDAC1 expression and activity.

ROS induced by DNA hypomethylation is an important factor for the progression of genomic instability and is, in turn, a source of ROS accumulation. One of the main causes of genomic instability is thought to be a result of alterations in oxygen metabolism which can give rise to increased levels of ROS. Genomic instability arises in a few cells capable of sustaining the ROS production. These cells accumulate further changes possibly due to epigenetic factors and to gene mutations induced by the high ROS levels, acquire selective advantage and can proliferate, even with their genomic instability. Progeny of these cells may exhibit memory of genome changes that can lead to a transformed phenotype [67,68].

Conclusions

Oxidative stress, as a consequence of ROS accumulation, increases exponentially with age, in parallel with a decline in the cell repair machinery, resulting in many diseases associated with aging including cancer and respiratory and cardiovascular diseases [69]. It is possible that targeting epigenetic regulators may be an important new therapeutic avenue for suppressing oxidative stress in cancer and other human diseases. A fundamental question now in the field of epigenetics is to understand the biochemical mechanisms underlying ROS-dependent regulation of epigenetic modification, which may open the door to identifying new therapeutic modalities. For example, in oncology, further studies of the epigenetic mark profiles from primary tumor samples will provide important information on the role of methylation of the CpG islands or other epigenetic marks in the promoter regions of tumor suppressor genes. Deciphering the methylation status of tumor suppressor genes may contribute to the regulation of the transcriptional activity of tumor suppressor genes,

which could be used in cancer preventive and therapeutic treatment.

References

- Valko M, Izakovic M, Mazur M, Rhodes CJ, Telser J (2004) Role of oxygen radicals in DNA damage and cancer incidence. *Mol Cell Biochem* 266: 37-56.
- Valko M, Rhodes CJ, Moncol J, Izakovic M, Mazur M (2006) Free radicals, metals and antioxidants in oxidative stress-induced cancer. *Chem Biol Interact* 160: 1-40.
- Davies KJ (2000) Oxidative stress, antioxidant defenses, and damage removal, repair, and replacement systems. *IUBMB Life* 50: 279-289.
- Berger SL, Kouzarides T, Shiekhattar R, Shilatifard A (2009) An operational definition of epigenetics. *Genes Dev* 23: 781-783.
- Illi B, Colussi C, Grasselli A, Farsetti A, Capogrossi MC, et al. (2009) NO sparks off chromatin: tales of a multifaceted epigenetic regulator. *Pharmacol Ther* 123: 344-352.
- Cedar H, Bergman Y (2012) Programming of DNA methylation patterns. *Annu Rev Biochem* 81: 97-117.
- Williams K, Christensen J, Helin K (2011) DNA methylation: TET proteins-guardians of CpG islands? *EMBO Rep* 13: 28-35.
- Grønbaek K, Hother C, Jones PA (2007) Epigenetic changes in cancer. *APMIS* 115: 1039-1059.
- Okano M, Bell DW, Haber DA, Li E (1999) DNA methyltransferases Dnmt3a and Dnmt3b are essential for de novo methylation and mammalian development. *Cell* 99: 247-257.
- Chahal SS, Matthews HR, Bradbury EM (1980) Acetylation of histone H4 and its role in chromatin structure and function. *Nature* 287: 76-79.
- Lachner M, O'Carroll D, Rea S, Mechtler K, Jenuwein T (2001) Methylation of histone H3 lysine 9 creates a binding site for HP1 proteins. *Nature* 410: 116-120.
- Schotta G, Lachner M, Sarma K, Ebert A, Sengupta R, et al. (2004) A silencing pathway to induce H3-K9 and H4-K20 trimethylation at constitutive heterochromatin. *Genes Dev* 18: 1251-1262.
- Santos-Rosa H, Schneider R, Bannister AJ, Sherriff J, Bernstein BE, et al. (2002) Active genes are tri-methylated at K4 of histone H3. *Nature* 419: 407-411.
- Ruthenburg AJ, Allis CD, Wysocka J (2007) Methylation of lysine 4 on histone H3: intricacy of writing and reading a single epigenetic mark. *Mol Cell* 25: 15-30.
- Strahl BD, Allis CD (2000) The language of covalent histone modifications. *Nature* 403: 41-45.
- Seligson DB, Horvath S, Shi T, Yu H, Tze S, et al. (2005) Global histone modification patterns predict risk of prostate cancer recurrence. *Nature* 435: 1262-1266.
- Minardi D, Lucarini G, Filosa A, Milanese G, Zizzi A, et al. (2009) Prognostic role of global DNA-methylation and histone acetylation in pT1a clear cell renal carcinoma in partial nephrectomy specimens. *J Cell Mol Med* 13: 2115-2121.
- Tzao C, Tung HJ, Jin JS, Sun GH, Hsu HS, et al. (2009) Prognostic significance of global histone modifications in resected squamous cell carcinoma of the esophagus. *Mod Pathol* 22: 252-260.
- Park YS, Jin MY, Kim YJ, Yook JH, Kim BS, et al. (2008) The global histone modification pattern correlates with cancer recurrence and overall survival in gastric adenocarcinoma. *Ann Surg Oncol* 15: 1968-1976.
- Elsheikh SE, Green AR, Rakha EA, Powe DG, Ahmed RA, et al. (2009) Global histone modifications in breast cancer correlate with tumor phenotypes, prognostic factors, and patient outcome. *Cancer Res* 69: 3802-3809.
- Ellinger J, Kahl P, von der Gathen J, Rogenhofer S, Heukamp LC, et al. (2010) Global levels of histone modifications predict prostate cancer recurrence. *Prostate* 70: 61-69.
- Esteller M (2011) Non-coding RNAs in human disease. *Nat Rev Genet* 12: 861-874.
- Pasquinelli AE (2012) MicroRNAs and their targets: recognition, regulation and an emerging reciprocal relationship. *Nat Rev Genet* 13: 271-282.
- Repine JE, Bast A, Lankhorst I (1997) Oxidative stress in chronic obstructive pulmonary disease. Oxidative Stress Study Group. *Am J Respir Crit Care Med* 156: 341-357.
- Tuder RM1, Kern JA, Miller YE (2012) Senescence in chronic obstructive pulmonary disease. *Proc Am Thorac Soc* 9: 62-63.
- Yao H, Rahman I (2012) Role of histone deacetylase 2 in epigenetics and cellular senescence: implications in lung inflammation and COPD. *Am J Physiol Lung Cell Mol Physiol* 303: L557-566.
- Sundar IK, Yao H, Rahman I (2013) Oxidative stress and chromatin remodeling in chronic obstructive pulmonary disease and smoking-related diseases. *Antioxid Redox Signal* 18: 1956-1971.
- Adenuga D, Yao H, March TH, Seagrave J, Rahman I (2009) Histone deacetylase 2 is phosphorylated, ubiquitinated, and degraded by cigarette smoke. *Am J Respir Cell Mol Biol* 40: 464-473.
- Rajendrasozhan S, Yang SR, Kinnula VL, Rahman I (2008) SIRT1, an antiinflammatory and antiaging protein, is decreased in lungs of patients with chronic obstructive pulmonary disease. *Am J Respir Crit Care Med* 177: 861-870.
- Yang SR, Chida AS, Bauter MR, Shafiq N, Seweryniak K, et al. (2006) Cigarette smoke induces proinflammatory cytokine release by activation of NF-kappaB and posttranslational modifications of histone deacetylase in macrophages. *Am J Physiol Lung Cell Mol Physiol* 291: L46-57.
- Lawless MW, O'Byrne KJ, Gray SG (2009) Oxidative stress induced lung cancer and COPD: opportunities for epigenetic therapy. *J Cell Mol Med* 13: 2800-2821.
- North BJ, Sinclair DA (2012) The intersection between aging and cardiovascular disease. *Circ Res* 110: 1097-1108.
- Feinberg AP, Ohlsson R, Henikoff S (2006) The epigenetic progenitor origin of human cancer. *Nat Rev Genet* 7: 21-33.
- Weinberg F, Hamanaka R, Wheaton WW, Weinberg S, Joseph J, et al. (2010) Mitochondrial metabolism and ROS generation are essential for Kras-mediated tumorigenicity. *Proc Natl Acad Sci U S A* 107: 8788-8793.
- Wallace DC (2012) Mitochondria and cancer. *Nat Rev Cancer* 12: 685-698.
- Cairns RA, Harris IS, Mak TW (2011) Regulation of cancer cell metabolism. *Nat Rev Cancer* 11: 85-95.
- Shaw AT, Winslow MM, Magendantz M, Ouyang C, Dowdle J, et al. (2011) Selective killing of K-ras mutant cancer cells by small molecule inducers of oxidative stress. *Proc Natl Acad Sci U S A* 108: 8773-8778.
- Ichijo H, Nishida E, Irie K, ten Dijke P, Saitoh M, et al. (1997) Induction of apoptosis by ASK1, a mammalian MAPKKK that activates SAPK/JNK and p38 signaling pathways. *Science* 275: 90-94.
- Hayes JD, McMahon M (2009) NRF2 and KEAP1 mutations: permanent activation of an adaptive response in cancer. *Trends Biochem Sci* 34: 176-188.
- Schafer ZT, Grassian AR, Song L, Jiang Z, Gerhart-Hines Z, et al. (2009) Antioxidant and oncogene rescue of metabolic defects caused by loss of matrix attachment. *Nature* 461: 109-113.
- Mitsuishi Y, Taguchi K, Kawatani Y, Shibata T, Nukiwa T, et al. (2012) Nrf2 redirects glucose and glutamine into anabolic pathways in metabolic reprogramming. *Cancer Cell* 22: 66-79.
- Young TW, Mei FC, Yang G, Thompson-Lanza JA, Liu J, et al. (2004) Activation of antioxidant pathways in ras-mediated oncogenic transformation of human surface ovarian epithelial cells revealed by functional proteomics and mass spectrometry. *Cancer Res* 64: 4577-4584.
- DeNicola GM, Karreth FA, Humpton TJ, Gopinathan A, Wei C, et al. (2011) Oncogene-induced Nrf2 transcription promotes ROS detoxification and tumorigenesis. *Nature* 475: 106-109.
- Ren D, Villeneuve NF, Jiang T, Wu T, Lau A, et al. (2011) Brusatol enhances the efficacy of chemotherapy by inhibiting the Nrf2-mediated defense mechanism. *Proc Natl Acad Sci U S A* 108: 1433-1438.
- Raj L, Ide T, Gurkar AU, Foley M, Schenone M, et al. (2011) Selective killing of cancer cells by a small molecule targeting the stress response to ROS. *Nature* 475: 231-234.
- Trachootham D, Zhou Y, Zhang H, Demizu Y, Chen Z, et al. (2006) Selective killing of oncogenically transformed cells through a ROS-mediated mechanism by beta-phenylethyl isothiocyanate. *Cancer Cell* 10: 241-252.
- Sun S, Schiller JH, Gazdar AF (2007) Lung cancer in never smokers—a different disease. *Nat Rev Cancer* 7: 778-790.
- Somwar R, Erdjument-Bromage H, Larsson E, Shum D, Lockwood WW, et al. (2011) Superoxide dismutase 1 (SOD1) is a target for a small molecule identified in a screen for inhibitors of the growth of lung adenocarcinoma cell lines. *Proc Natl Acad Sci U S A* 108: 16375-16380.
- Glasauer A, Sena LA, Diebold LP, Mazar AP, Chandel NS (2014) Targeting SOD1 reduces experimental nona€small-cell lung cancer. *J Clin Invest* 124: 117-128.
- Lawless MW, Norris S, O'Byrne KJ, Gray SG (2009) Targeting histone deacetylases for the treatment of disease. *J Cell Mol Med* 13: 826-852.
- Cary KC, Cooperberg MR (2013) Biomarkers in prostate cancer surveillance and screening: past, present, and future. *Ther Adv Urol* 5: 318-329.
- Frohlich DA, McCabe MT, Arnold RS, Day ML (2008) The role of Nrf2 in increased reactive oxygen species and DNA damage in prostate tumorigenesis. *Oncogene* 27: 4353-4362.

53. Yu S, Kong AN (2007) Targeting carcinogen metabolism by dietary cancer preventive compounds. *Curr Cancer Drug Targets* 7: 416-424.
54. Yu S, Khor TO, Cheung KL, Li W, Wu TY, et al. (2010) Nrf2 expression is regulated by epigenetic mechanisms in prostate cancer of TRAMP mice. *PLoS One* 5: e8579.
55. Khor TO, Huang Y, Wu TY, Shu L, Lee J, et al. (2011) Pharmacodynamics of curcumin as DNA hypomethylation agent in restoring the expression of Nrf2 via promoter CpGs demethylation. *Biochem Pharmacol* 82: 1073-1078.
56. Khor TO, Fuentes F, Shu L, Paredes-Gonzalez X, Yang AY, et al. (2014) Epigenetic DNA Methylation of Antioxidative Stress Regulator NRF2 in Human Prostate Cancer. *Cancer Prev Res (Phila)* 7: 1186-1197.
57. Bostwick DG, Meiers I, Shanks JH (2007) Glutathione S-transferase: differential expression of alpha, mu, and pi isoenzymes in benign prostate, prostatic intraepithelial neoplasia, and prostatic adenocarcinoma. *Hum Pathol* 38: 1394-1401.
58. Kotrikadze N, Alibegashvili M, Zibzibadze M, Abashidze N, Chigogidze T, et al. (2008) Activity and content of antioxidant enzymes in prostate tumors. *Exp Oncol* 30: 244-247.
59. Kang KA, Zhang R, Kim GY, Bae SC, Hyun JW (2012) Epigenetic changes induced by oxidative stress in colorectal cancer cells: methylation of tumor suppressor RUNX3. *Tumour Biol* 33: 403-412.
60. Bae SC, Choi JK (2004) Tumor suppressor activity of RUNX3. *Oncogene* 23: 4336-4340.
61. Hiramatsu T, Osaki M, Ito Y, Tanji Y, Tokuyasu N, et al. (2005) Expression of RUNX3 protein in human esophageal mucosa and squamous cell carcinoma. *Pathobiology* 72: 316-324.
62. Oshimo Y, Oue N, Mitani Y, Nakayama H, Kitadai Y, et al. (2004) Frequent loss of RUNX3 expression by promoter hypermethylation in gastric carcinoma. *Pathobiology* 71: 137-143.
63. Subramaniam MM, Chan JY, Yeoh KG, Quek T, Ito K, et al. (2009) Molecular pathology of RUNX3 in human carcinogenesis. *Biochim Biophys Acta* 1796: 315-331.
64. Li QL, Ito K, Sakakura C, Fukamachi H, Inoue Ki, et al. (2002) Causal relationship between the loss of RUNX3 expression and gastric cancer. *Cell* 109: 113-124.
65. Ito K, Liu Q, Salto-Tellez M, Yano T, Tada K, et al. (2005) RUNX3, a novel tumor suppressor, is frequently inactivated in gastric cancer by protein mislocalization. *Cancer Res* 65: 7743-7750.
66. Zhang R, Kang KA, Kim KC, Na SY, Chang WY, et al. (2013) Oxidative stress causes epigenetic alteration of CDX1 expression in colorectal cancer cells. *Gene* 524: 214-219.
67. Sciandrello G, Caradonna F, Mauro M, Barbata G (2004) Arsenic-induced DNA hypomethylation affects chromosomal instability in mammalian cells. *Carcinogenesis* 25: 413-417.
68. Sciandrello G, Mauro M, Catanzaro I, Saverini M, Caradonna F, et al. (2011) Long-lasting genomic instability following arsenite exposure in mammalian cells: the role of reactive oxygen species. *Environ Mol Mutagen* 52: 562-568.
69. Sohal RS, Weindruch R (1996) Oxidative stress, caloric restriction, and aging. *Science* 273: 59-63.